

# Package ‘geneRxCluster’

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**License** GPL (>= 2)

**Description** Detect Differential Clustering of Genomic Sites such as gene therapy integrations. The package provides some functions for exploring genomic insertion sites originating from two different sources. Possibly, the two sources are two different gene therapy vectors. Vectors are preferred that target sensitive regions less frequently, motivating the search for localized clusters of insertions and comparison of the clusters formed by integration of different vectors. Scan statistics allow the discovery of spatial differences in clustering and calculation of False Discovery Rates (FDRs) providing statistical methods for comparing retroviral vectors. A scan statistic for comparing two vectors using multiple window widths to detect clustering differentials and compute FDRs is implemented here.

**Title** gRx Differential Clustering

**Author** Charles Berry

**Maintainer** Charles Berry <ccberry@ucsd.edu>

**Depends** GenomicRanges,IRanges

**Suggests** RUnit, BiocGenerics

**biocViews** Sequencing, Clustering, Genetics

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|               |                         |
|---------------|-------------------------|
| critVal.alpha | <i>critical regions</i> |
|---------------|-------------------------|

---

### Description

critical region cutpoints

### Usage

```
critVal.alpha(k, p0, alpha, posdiff)
```

### Arguments

|         |                              |
|---------|------------------------------|
| k       | - window width(s)            |
| p0      | - length 2 probabilities     |
| alpha   | - two tailed                 |
| posdiff | - position difference matrix |

### Details

This version uses alpha and will find TFD

### Value

list of cutoffs and attributes

### Author(s)

Charles Berry

**See Also**

[gRxCluster](#) for how and why this function is used

**Examples**

```
# symmetric odds:
crit <- critVal.alpha(5:25,c(1,1)/2,alpha=0.05,
                    matrix(1,nr=50,nc=21))

crit[[1]]
sapply(crit,c)
# 5:1 odds
asymmetric.crit <- critVal.alpha(5:25,c(1,5)/6,alpha=0.05,
                                matrix(1,nr=50,nc=21))

# show the critical regions
par(mfrow=c(1,2))
gRxPlot(crit,method="critical")
gRxPlot(asymmetric.crit,method="critical")
rm(crit,asymmetric.crit)
```

---

critVal.power

*critical regions*

---

**Description**

critical region cutpoints

**Usage**

```
critVal.power(k, p0, target, pwr = 0.8, odds = 7)
```

**Arguments**

|        |                            |
|--------|----------------------------|
| k      | - window width(s)          |
| p0     | - length 2 probabilities   |
| target | - false discoveries wanted |
| pwr    | - desired power            |
| odds   | - alternative odds ratio   |

**Details**

This version uses power and TFD and will limit windows screened

**Value**

list of cutoffs and attributes

**Author(s)**

Charles Berry

**See Also**[gRxCluster](#) for how and why this function is used**Examples**

```
# symmetric odds:
crit <-
  critVal.power(5:25,c(1,1),5,pwr=0.8,odds=7)
crit[[1]]
sapply(crit,c)
# 5:1 odds
asymmetric.crit <-
  critVal.power(5:25,c(1,5),5,pwr=0.8,odds=7)
# show the critical regions
par(mfrow=c(1,2))
gRxPlot(crit,method="critical")
gRxPlot(asymmetric.crit,method="critical")
rm(crit,asymmetric.crit)
```

---

critVal.target

*critical regions*


---

**Description**

critical region cutpoints

**Usage**

```
critVal.target(k, p0, target, posdiff = NULL, ns)
```

**Arguments**

|         |  |
|---------|--|
| k       | window width(s)                                |
| p0      | length 2 probabilities                         |
| target  | - two tailed                                   |
| posdiff | - position difference matrix                   |
| ns      | the number of windows passing filter at each k |

**Details**

This version uses TFD and will find alpha implicitly

**Value**

list of cutoffs and attributes

**Author(s)**

Charles Berry

**See Also**

[gRxCluster](#) for how and why this function is used

**Examples**

```
# symmetric odds:
crit <- critVal.target(5:25,c(1,1),1,ns=rep(10,21))
crit[[1]]
sapply(crit,c)
# 5:1 odds
asymmetric.crit <- critVal.target(5:25,c(1,5),1,ns=rep(10,21))
# show the critical regions
par(mfrow=c(1,2))
gRxPlot(crit,method="critical")
gRxPlot(asymmetric.crit,method="critical")
rm(crit,asymmetric.crit)
```

**Description**

geneRxCluster provides the function [gRxCluster](#) and friends.

**Details**

Windows defined by  $k$  consecutive integration sites are scanned. A two class indicator is tallied to determine whether one class dominates. If one does, a flag is set and the window is retained. Various values of  $k$  are used. Conflicts between overlapping windows with the same value of  $k$  can occur — two windows are dominated by the two different classes. In that case, the sites of overlap are marked and neither window is retained. Conflicts can also arise between windows differing in their values of  $k$ . In that case, the window having the smaller value of  $k$  is retained and the other is discarded.

Permutation tests and permutation based false discovery rates are available.

Filtering of windows is allowed so that regions which are sparsely populated need not be studied.

gRxCluster

*gRxCluster***Description**

cluster integration sites - optionally perform the permutations needed to estimate the discoveries expected under a null hypothesis

**Usage**

```
gRxCluster(object, starts, group, kvals, nperm = 0L,
  pruneFun = prune.loglik, ..., cutpt.filter.expr, cutpt.tail.expr, tmp.env,
  sample.id, sample.tab)
```

**Arguments**

|                   |   |
|-------------------|---|
| object            | chromosome names or other grouping of starts  |
| starts            | ordered chromosome position or ordered integer vector   |
| group             | logical vector separating two groups  |
| kvals             | integer vector of window widths   |
| nperm             | number of permutations for FDR calculation  |
| pruneFun          | a function like <code>prune.loglik</code> .   |
| ...               | other args  |
| cutpt.filter.expr | (optional) R object or call (or variable naming a call) with (optional) var <code>x</code> (window widths in base pairs) to filter windows. It must evaluate to mode "double". If not specified, <code>as.double(apply(x, 2, median, na.rm=TRUE))</code> is used. If an atomic vector of length one is supplied it is expanded to the proper length and coerced to double. If this arg is the name of a variable provided in <code>tmp.env</code> , it must be protected with <code>quote(...)</code> .   |
| cutpt.tail.expr   | R object or call (or variable naming a call) with (optional) vars: <code>k</code> , <code>n</code> , and <code>x</code> (as above). Returns list like <code>critVal.target</code> . <code>k</code> is a vector of the number of sites in a collection of windows, and <code>n</code> is a vector of counts or proportions for the two classes of insertion. If not supplied, <code>critVal.target(k, n, target=5, posdiff=x)</code> is used. If this arg is the name of a variable provided in <code>tmp.env</code> , it must be protected with <code>quote(...)</code> . |
| tmp.env           | (optional) environment to use in evaluation of <code>cutpt.*</code> expressions. This is usually needed for <code>critVal.power</code> , which is first calculated and placed in the environment, and the supplied object is used in the expression for <code>cutpt.filter.expr</code> .  |
| sample.id         | (optional) integer vector indexing cells in <code>sample.tab</code> to be looked up to determine group under permutation. A factor can be used, too, but will be coerced to integer.  |

`sample.tab` (optional) integer vector containing 0 or 1 in each cell. Its length is the same as `max(sample.id)`. Both or neither `sample.id` and `sample.tab` should be supplied. When supplied `sample.tab[sample.id]` must equal `group`. If the arguments are supplied, permutations are of the form `sample(sample.tab)[sample.id]`. Otherwise they are of the form `sample(group)`.

### Value

a GRanges object with a special metadata slot, see [gRxCluster-object](#)

### Author(s)

Charles Berry example inst/ex-gRxCluster.R

---

`gRxCluster-object`      *gRxCluster object*

---

### Description

Overview of the result of `gRxCluster(...)`

### Details

The object returned is a [GRanges](#) object.

If the object is `x`, `seqnames(x)` and `ranges(x)` slots demarcate the clusters discovered. There will be one element for each cluster (aka 'clump') discovered.

Using the default argument `pruneFun=prune.loglik` or `pruneFun=noprune`, `mcols(x)` will have these columns:

`value1` **and** `value2` are the counts of the two classes of insertion sites for the clusters of object `x`  
`clump.id` numbers each cluster.

If the user supplies a custom `pruneFun`, it should return a GRanges with those columns and one element for each unique `clump.id`. The column `target.min` has the smallest nominal False Discoveries Expected for each cluster and is added to (or replaces) the `mcols(x)` produced by the argument supplied as `pruneFun`.

`metadata(x)` will include these components:

**criticalValues** A list object such as supplied by `critVal.target` whose elements each give the cutpoints to be used for a window with `k` sites. `attributes(metadata(object)$criticalValues[[i]])` will contain elements

**fdr** with dimension `c(k+1, 4)` of target false discovery expectations and and the one-sided p-values

**target** the target for false discovery which sometimes is specified a priori and sometimes results from calculation

**n** an upper bound on the number of windows to screen, if this number is needed.

In some cases, an attribute is attached to `metadata(object)$criticalValues`, see `critVal.power` for an example.

**kvals** the number of sites,  $k$ , to include in a window

**perm\_cluster\_best** a list whose canonical element is a vector of values like `x$target.min` obtained from a permutation of the class indicators

**summary\_matrix** a matrix giving the start, end, depth, and counts in each class for every cluster and depth in sequential order

**call** the call invoking `gRxCluster` which may include some arguments added by default.

### Author(s)

Charles Berry <ccberry@ucsd.edu>

---

gRxPlot

*gRxPlot*

---

### Description

Plot Clumps and/or Critical Regions

### Usage

```
gRxPlot(object, pi.0 = NULL, method = c("odds", "criticalRegions"),
        xlim = NULL, main = NULL, xlab = "log odds ratio", breaks = "Sturges",
        kvals = NULL, ...)
```

### Arguments

|                     |  |
|---------------------|--|
| <code>object</code> | either the results of <code>gRxCluster</code> or a list containing cutpoints for critical regions. |
| <code>pi.0</code>   | the background proportion for vector 2   |
| <code>method</code> | character vector of "odds" and/or "criticalRegions"  |
| <code>xlim</code>   | limits of the log odds histogram   |
| <code>main</code>   | a title for the panel(s)   |
| <code>xlab</code>   | label for the x-axis of the log odds plot  |
| <code>breaks</code> | see <code>hist</code>  |
| <code>kvals</code>  | values to use in selecting a subset of the critical regions to display                             |
| <code>...</code>    | other args to pass to the plotting routine(s)  |

### Details

The results of a call to `gRxCluster` are plotted. Optionally, with `method="criticalRegions"` only the critical regions are plotted or with `method="odds"` the log odds only are plotted.



**Value**

see [hist](#)

**Author(s)**

Charles Berry

**See Also**

[gRxPlotClumps](#) for a more fine grained display

**Examples**

```
x.seqnames <- rep(letters[1:3],each=500)
x.starts <- c(seq(1,length=500),seq(1,by=2,length=500),seq(1,by=3,length=500))
x.lens <- rep(c(5,10,15,20,25),each=20)
x.group <- rep(rep(c(TRUE,FALSE),length=length(x.lens)),x.lens)
## add a bit of fuzz:
x.group <- 1==rbinom(length(x.group),1,pr=ifelse(x.group,.8,.2))
x.kvals <- as.integer(sort(unique(x.lens)))
x.res <- gRxCluster(x.seqnames,x.starts,x.group,x.kvals)
gRxPlot(x.res)
rm( x.seqnames, x.starts, x.lens, x.group, x.kvals, x.res)
```

---

`gRxPlotClumps`

*gRxPlotClumps*

---

**Description**

Plot gRxCluster object clumps

**Usage**

```
gRxPlotClumps(object, data, seqLens, panelExpr = quote(grid()))
```

**Arguments**

|                        |   |
|------------------------|---|
| <code>object</code>    | result of gRxCluster  |
| <code>data</code>      | (optional) GRanges like that from which args to gRxCluster were derived |
| <code>seqLens</code>   | (optional) seqlengths(data) or similar. Can be given if data is missing |
| <code>panelExpr</code> | - an expression to evaluate after drawing each panel                    |

**Details**

Plot Relative Frequencies of the two classes according to region. Regions typically alternate between clusters and non-clusters on each chromosome.

**Author(s)**

Charles Berry

**Examples**

```
x.seqnames <- rep(letters[1:3],each=50)
x.starts <- c(seq(1,length=50),seq(1,by=2,length=50),seq(1,by=3,length=50))
x.lens <- rep(c(5,10,15,20,25),each=2)
x.group <- rep(rep(c(TRUE,FALSE),length=length(x.lens)),x.lens)
## add a bit of fuzz:
x.group <- 1==rbinom(length(x.group),1,pr=ifelse(x.group,.8,.2))
x.kvals <- as.integer(sort(unique(x.lens)))
x.res <- gRxCluster(x.seqnames,x.starts,x.group,x.kvals)
gRxPlotClumps(x.res)
rm( x.seqnames, x.starts, x.lens, x.group, x.kvals, x.res)
```

gRxSummary

*gRxSummary***Description**

Summarize gRxCluster Results

**Usage**

```
gRxSummary(object, targetFD = NULL)
```

**Arguments**

|          |  |
|----------|--|
| object   | the result of gRxCluster               |
| targetFD | the critical value target in each tail |

**Details**

Get the FDR and related data for a run of gRxCluster. By selecting a value for targetFD that is smaller than what was used in constructing the object, fewer clumps will be included in the computation for the False Discovery Rate - akin to what would have been obtained from the object if it had been constructed using that value.

**Value**

a list containing the summarized results

**Author(s)**

Charles Berry

**Examples**

```
x.seqnames <- rep(letters[1:3],each=50)
x.starts <- c(seq(1,length=50),seq(1,by=2,length=50),seq(1,by=3,length=50))
x.lens <- rep(c(5,10,15,20,25),each=2)
x.group <- rep(rep(c(TRUE,FALSE),length=length(x.lens)),x.lens)
x.kvals <- as.integer(sort(unique(x.lens)))
x.res <- gRxCluster(x.seqnames,x.starts,x.group,x.kvals,nperm=100L)
gRxSummary(x.res)
rm( x.seqnames, x.starts, x.lens, x.group, x.kvals, x.res)
```

---

noprune

*noprune*

---

**Description**

join contiguous windows

**Usage**

```
noprune(x, ...)
```

**Arguments**

|     |                  |
|-----|------------------|
| x   | a GRanges object |
| ... | currently unused |

**Details**

return all the candidate sites in a clump without pruning  
this is to be used as the pruneFun are of gRxCluster

**Value**

same as [gRxCluster](#) less the metadata

**Author(s)**

Charles Berry

**See Also**

[gRxCluster-object](#) for more details on what this function returns.

plot.cutpoints      *plot.cutpoints*

---

**Description**

Plot a set of cutpoints - Utility

**Usage**

```
## S3 method for class 'cutpoints'  
plot(crit, pi.0 = NULL, kvals = NULL, ...)
```

**Arguments**

crit                    - a cutpoint object see [gRxCluster](#)  
pi.0                    - optional null value to plot  
kvals                   - which cutpoints to include in the plot  
...                     passed to barplot

**Details**

NOT FOR USERS. Not exported.

**Value**

list with components of “bar.x” (the value of hist()), “kvals” (window widths plotted), and “pi.0” (the input value of pi.0)

**Author(s)**

Charles Berry

---

prune.loglik      *prune.loglik*

---

**Description**

best contiguous region

**Usage**

```
prune.loglik(x, p.null = 0.5)
```

**Arguments**

|        |                                       |
|--------|---------------------------------------|
| x      | a GRanges object                      |
| p.null | the probability of category 1 (FALSE) |

**Details**

prune each end of the region using loglik criterion  
this is to be used as the pruneFun are of gRxCluster

**Author(s)**

Charles Berry

**See Also**

[gRxCluster-object](#) for details on what this function returns.

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