Package ‘syntenet’

May 3, 2024

Title Inference And Analysis Of Synteny Networks

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Description syntenet can be used to infer synteny networks from
whole-genome protein sequences and analyze them. Anchor pairs
are detected with the MCScanX algorithm, which was ported to this package
with the Rcpp framework for R and C++ integration.
Anchor pairs from synteny analyses are
treated as an undirected unweighted graph (i.e., a synteny network),
and users can perform: i. network clustering; ii. phylogenomic profiling
(by identifying which species contain which clusters) and;
iii. microsynteny-based phylogeny reconstruction with maximum likelihood.

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BugReports https://support.bioconductor.org/t/syntenet

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Description

syntenet can be used to infer synteny networks from whole-genome protein sequences and analyze them. Anchor pairs are detected with the MCScanX algorithm, which was ported to this package with the Rcpp framework for R and C++ integration. Anchor pairs from synteny analyses are treated as an undirected unweighted graph (i.e., a synteny network), and users can perform: i. network clustering; ii. phylogenomic profiling (by identifying which species contain which clusters) and; iii. microsynteny-based phylogeny reconstruction with maximum likelihood.

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See Also

Useful links:

• https://github.com/almeidasilvaf/syntenet
• Report bugs at https://support.bioconductor.org/t/syntenet
angiosperm_phylogeny  
Microsynteny-based angiosperm phylogeny.

Description
Original tree file obtained from Zhao et al., 2021. The tree is an object of class 'phylo', which can be created by reading the tree file with `treeio::read.tree()`.

Usage
data(angiosperm_phylogeny)

Format
An object of class 'phylo'.

References

Examples
data(angiosperm_phylogeny)

annotation  
Filtered genome annotation for Ostreococcus sp. species

Description
Data obtained from Pico-PLAZA 3.0. Only annotation data for primary transcripts were included, and only genes for chromosomes 1, 2, and 3.

Usage
data((annotation)

Format
A CompressedGRangesList containing the elements Olucimarinus, Osp_RCC809, and Otauri.

References
binarize_and_transpose

Examples
data(annotation)

---

Binarize and transpose the phylogenomic profile matrix

Description
Binarize and transpose the phylogenomic profile matrix

Usage
binarize_and_transpose(profile_matrix = NULL)

Arguments
profile_matrix A matrix with phylogenomic profiles obtained with phylogenomic_profile.

Value
A binary and transposed version of the profiles matrix.

Examples
data(clusters)
profile_matrix <- phylogenomic_profile(clusters)
tmat <- binarize_and_transpose(profile_matrix)

---

blast_list

List of data frames containing BLAST-like tabular output

Description
The object was created by running run_diamond on the protein sequences for the Ostreococcus algae available in the proteomes example data. Hits with <50% identity were filtered out. Code to recreate this data is available at the script/ subdirectory.

Usage
data(blast_list)

Format
A list of data frames containing the pairwise comparisons between proteomes of Ostreococcus species.
check_input

Check if input objects are ready for further analyses

Description
Check if input objects are ready for further analyses

Usage
check_input(seq = NULL, annotation = NULL, gene_field = "gene_id")

Arguments

seq A list of AAStringSet objects, each list element containing protein sequences for a given species. This list must have names (not NULL), and names of each list element must match the names of list elements in annotation.

annotation A GRangesList, CompressedGRangesList, or list of GRanges with the annotation for the sequences in seq. This list must have names (not NULL), and names of each list element must match the names of list elements in seq.

gene_field Character, name of the column in the GRanges objects that contains gene IDs. Default: "gene_id".

Details
This function checks the input data for 3 required conditions:

1. Names of seq list (i.e., names(seq)) match the names of annotation GRangesList/CompressedGRangesList (i.e., names(annotation))

2. For each species (list elements), the number of sequences in seq is not greater than the number of genes in annotation. This is a way to ensure users do not input the translated sequences for multiple isoforms of the same gene (generated by alternative splicing). Ideally, the number of sequences in seq should be equal to the number of genes in annotation, but this may not always stand true because of non-protein-coding genes.

3. For each species, sequence names (i.e., names(seq[[x]]), equivalent to FASTA headers) match gene names in annotation.

Value
TRUE if the objects pass the check.

Examples
data(abbreviation)
data(proteomes)
check_input(proteomes, annotation)
clusters

Synten network clusters of BUSCO genes for 25 eudicot species

Description

Data obtained from Zhao & Schranz, 2019.

Usage

data(clusters)

Format

A 2-column data frame containing the following variables:

Gene  Gene ID
Cluster  Cluster ID

References


Examples

data(clusters)

cluster_network

Cluster the synteny network using the Infomap algorithm

Description

Cluster the synteny network using the Infomap algorithm

Usage

cluster_network(
  network = NULL,
  clust_function = igraph::cluster_infomap,
  clust_params = NULL
)
**collapse_protein_ids**

**Arguments**

- **network**
  - A network represented as an edge list, which is a 2-column data frame with node 1 in the first column and node 2 in the second column. In a synteny network, node 1 and node 2 are the anchor pairs.

- **clust.function**
  - Function to be used to cluster the network. It must be one of the functions from the `cluster_*` family in the `igraph` package (e.g., `cluster_infomap`, `cluster_leiden`, etc). Default: `igraph::cluster_infomap`.

- **clust.params**
  - A list with additional parameters (if any) to be passed to the igraph clustering function. Default: `NULL` (no additional parameters).

**Value**

A 2-column data frame with the following variables:

- **Gene** Gene ID.
- **Cluster** Cluster ID as identified by infomap.

**Examples**

```r
data(network)
clusters <- cluster_network(network[1:500,])
```

**collapse_protein_ids**

*Collapse protein IDs into gene IDs in sequence names of AASTringSet objects*

**Description**

This function can be used if the sequence names of the AASTringSet objects contain protein IDs instead of gene IDs (what syntenet requires)

**Usage**

```r
collapse_protein_ids(seq, protein2gene = NULL)
```

**Arguments**

- **seq**
  - A list of AASTringSet objects, each list element containing protein sequences for a given species. This list must have names (not NULL), and names of each list element must match the names of list elements in `protein2gene`.

- **protein2gene**
  - A list of 2-column data frames containing protein-to-gene ID correspondences, where the first column contains protein IDs, and the second column contains gene IDs. Names of list elements must match names of `seq`. 
Details

For each species, this function will replace the protein IDs in sequence names with gene IDs using the protein-to-gene correspondence table in `protein2gene`. After replacing protein IDs with gene IDs, if there are multiple sequences with the same gene ID (indicating different isoforms of the same gene), only the longest sequence is kept, so that the number of sequences is not greater than the number of genes.

Value

A list of AAStringSet objects as in `seq`, but with protein IDs replaced with gene IDs.

Examples

```r
# Load data
seq_path <- system.file("extdata", "RefSeq_parsing_example", package = "syntenet")
seq <- fasta2AAStringSetlist(seq_path)
annot <- gff2GRangesList(seq_path)

# Clean sequence names
names(seq$Aalosa) <- gsub(".*", ",", names(seq$Aalosa))

# Create a correspondence data frame
cor_df <- as.data.frame(annot$Aalosa[annot$Aalosa$type == "CDS", ])
cor_df <- cor_df[, c("Name", "gene")]

# Create a list of correspondence data frames
protein2gene <- list(Aalosa = cor_df)

# Collapse IDs
new_seqs <- collapse_protein_ids(seq, protein2gene)
```

---

create_species_id_table

Create a data frame of species IDs (3-5-character abbreviations)

Description

Create a data frame of species IDs (3-5-character abbreviations)

Usage

```r
create_species_id_table(species_names)
```

Arguments

- `species_names`: A character vector of names extracted from the `seq` or `annotation` lists, which can be extracted with `names(seq)` or `names(annot)`. 
Value
A 2-column data frame with the following variables:

species_id Character, species ID consisting of 3-5 characters.
species_name Character, original names passed as input.

Examples

# Load 'seq' list (list of AAStringSet objects)
data(proteomes)

# Create ID table
create_species_id_table(names(proteomes))

diamond_is_installed  Check if DIAMOND is installed

Description
Check if DIAMOND is installed

Usage
diamond_is_installed()

Value
Logical indicating whether DIAMOND is installed or not.

Examples
diamond_is_installed()

edges  Synteny network of Ostreococcus genomes represented as an edge list

Description
The object was created by running infer_syntenet on the blast_list example data. Code to recreate this data set is available at the script/ subdirectory.

Usage
data(edges)
**export_sequences**

**Format**

A data frame containing anchor pairs between two Ostreococcus proteomes.

**Examples**

```r
data(edges)
```

---

**Description**

Export processed sequences as FASTA files

**Usage**

```r
export_sequences(seq = NULL, outdir = tempdir())
```

**Arguments**

- `seq` A processed list of AAStringSet objects as returned by `process_input()`.
- `outdir` Path to output directory where FASTA files will be stored.

**Value**

Path to exported FASTA files.

**Examples**

```r
# Load data
data(proteomes)
data(annotation)

# Process data
pdata <- process_input(proteomes, annotation)

# Export data
outdir <- file.path(tempdir(), "example_test")
export_sequences(pdata$seq, outdir)
```
**fasta2AAStringSetlist**  
*Read FASTA files in a directory as a list of AAStringSet objects*

**Description**

Read FASTA files in a directory as a list of AAStringSet objects

**Usage**

```r
fasta2AAStringSetlist(fasta_dir)
```

**Arguments**

- `fasta_dir` 
  Character indicating the path to the directory containing FASTA files.

**Value**

A list of AAStringSet objects, where each element represents a different FASTA file.

**Examples**

```r
fasta_dir <- system.file("extdata", "sequences", package = "syntenet")
aastringsetlist <- fasta2AAStringSetlist(fasta_dir)
```

---

**find_GS_clusters**  
*Find group-specific clusters based on user-defined species classification*

**Description**

Find group-specific clusters based on user-defined species classification

**Usage**

```r
find_GS_clusters(
  profile_matrix = NULL,
  species_annotation = NULL,
  min_percentage = 50
)
```
Arguments

profile_matrix  A matrix of phylogenomic profiles obtained with phylogenomic_profile.

species_annotation
A 2-column data frame with species IDs in the first column (same as column names of profile matrix), and species annotation (e.g., higher-level taxonomic information) in the second column.

min_percentage Numeric scalar with the minimum percentage of species in a group to consider group specificity. For instance, if a given cluster is present in only 1 group of species, but in less than min_percentage of the species for this group, it will not be considered a group-specific cluster. This filtering criterion is useful to differentiate group-specific clusters (e.g., family-specific) from subgroup-specific clusters (e.g., genus-specific). Default: 50.

Value

A data frame with the following variables:

Group To which group of species the cluster is specific.

Percentage Percentage of species from the group that are represented by the cluster.

Cluster Cluster ID.

Examples

data(clusters)
profile_matrix <- phylogenomic_profile(clusters)

# Species annotation
species_order <- c(
   "Lang", "car", "pmu", "ppe", "ppr", "mdo", "roc", "fve",
   "Mnot", "Zjuj", "hlu", "jcu", "mes", "rco", "lus", "ptr"
)

species_annotation <- data.frame(
   Species = species_order,
   Family = c(rep("Fabaceae", 11), rep("Rosaceae", 6),
             "Moraceae", "Rhamnaceae", "Cannabaceae",
             rep("Euphorbiaceae", 3), "Linaceae", "Salicaceae")
)
gs_clusters <- find_GS_clusters(profile_matrix, species_annotation)

---

**gff2GRangesList**  
*Read GFF/GTF files in a directory as a GRangesList object*

**Description**

Read GFF/GTF files in a directory as a GRangesList object
Usage

```r
gff2GRangesList(gff_dir)
```

Arguments

- `gff_dir` Character indicating the path to the directory containing GFF/GTF files.

Value

A GRangesList object, where each element represents a different GFF/GTF file.

Examples

```r
gff_dir <- system.file("extdata", "annotation", package = "syntenet")
grangeslist <- gff2GRangesList(gff_dir)
```

---

### infer_microsynten PHYLOGENY

Infer microsynten-based phylogeny with IQTREE

**Usage**

```r
infer_microsynten PHYLOGENY(
  transposed_profiles = NULL,
  bootr = 1000,
  alrtboot = 1000,
  threads = "AUTO",
  model = "MK+F0+R",
  outdir = tempdir(),
  outgroup = NULL,
  verbose = FALSE
)
```

**Arguments**

- `transposed_profiles` A binary and transposed profile matrix. The profile matrix can be obtained with `phylogenomic_profile()`.
- `bootr` Numeric scalar with the number of bootstrap replicates. Default: 1000.
- `alrtboot` Numeric scalar with the number of replicates for the SH-like approximate likelihood ratio test. Default: 1000.
infer_syntenet

threads Numeric scalar indicating the number of threads to use or "AUTO", which allows IQTREE to automatically choose the best number of threads to use. Default: "AUTO".

model Substitution model to use. If you are unsure, pick the default. Default: "MK+FO+R".

outdir Path to output directory. By default, files are saved in a temporary directory, so they will be deleted when the R session closes. If you want to keep the files, specify a custom output directory.

outgroup Name of outgroup clade to group the phylogeny. Default: NULL (unrooted phylogeny).

verbose Logical indicating if progress messages should be prompted. Default: FALSE.

Value

A character vector of paths to output files.

Examples

data(clusters)
profile_matrix <- phylogenomic_profile(clusters)
tmat <- binarize_and_transpose(profile_matrix)

# Leave only some legumes and P. mume as an outgroup for testing purposes
included <- c("gma", "pvu", "vra", "van", "cca", "pmu")
tmat <- tmat[rownames(tmat) %in% included,]

# Remove non-variable sites
tmat <- tmat[, colSums(tmat) != length(included)]

if(iqtree_is_installed()) {
  phylo <- infer_microsynteny_phylogeny(tmat, outgroup = "pmu",
                                        threads = 1)
}

infer_syntenet Infer synteny network

Description

Infer synteny network

Usage

infer_syntenet(
  blast_list = NULL,
  annotation = NULL,
  outdir = tempdir(),
infer_syntenet

anchors = 5,
max_gaps = 25,
is_pairwise = TRUE,
verbose = FALSE,
bp_param = BiocParallel::SerialParam(),
...
)

Arguments

blast_list A list of data frames, each data frame having the tabular output of BLASTp or similar programs, such as DIAMOND. This is the output of the function run_diamond(). If you performed pairwise comparisons on the command line, you can read the tabular output as data frames and combine them in a list. List names must be have species names separated by underscore. For instance, if the first list element is a data frame containing the comparison of speciesA (query) against speciesB (database), its name must be "speciesA_speciesB".

annotation A processed GRangesList, CompressedGRangesList, or list of GRanges as returned by process_input().

outdir Path to the output directory. Default: tempdir().

anchors Numeric indicating the minimum required number of genes to call a syntenic block. Default: 5.

max_gaps Numeric indicating the number of upstream and downstream genes to search for anchors. Default: 25.

is_pairwise specify if only pairwise blocks should be reported Default: TRUE

verbose Logical indicating if log messages should be printed on screen. Default: FALSE.

bp_param BiocParallel back-end to be used. Default: BiocParallel::SerialParam().

... Any additional arguments to the MCScanX algorithm. For a complete list of all available options, see the man page of rcpp_mcsanx_file().

Value

A network represented as an edge list.

Examples

# Load data
data(proteomes)
data(annotation)
data(blast_list)

# Create processed annotation list
annotation <- process_input(proteomes, annotation)$annotation

# Infer the synteny network
net <- infer_syntenet(blast_list, annotation)
Description

Detect interspecies synteny

Usage

interspecies_synteny(
  blast_inter = NULL,
  annotation = NULL,
  inter_dir = file.path(tempdir(), "inter"),
  anchors = 5,
  max_gaps = 25,
  is_pairwise = TRUE,
  verbose = FALSE,
  bp_param = BiocParallel::SerialParam(),
  ...
)

Arguments

- **blast_inter**: A list of BLAST/DIAMOND data frames for interspecies comparisons as returned by `run_diamond()`.
- **annotation**: A processed GRangesList or CompressedGRangesList object as returned by `process_input()`.
- **inter_dir**: Path to output directory where .collinearity files will be stored.
- **anchors**: Numeric indicating the minimum required number of genes to call a syntenic block. Default: 5.
- **max_gaps**: Numeric indicating the number of upstream and downstream genes to search for anchors. Default: 25.
- **is_pairwise**: Specify if only pairwise blocks should be reported. Default: TRUE.
- **verbose**: Logical indicating if log messages should be printed on screen. Default: FALSE.
- **bp_param**: BiocParallel back-end to be used. Default: BiocParallel::SerialParam().
- **...**: Any additional arguments to the MCScanX algorithm. For a complete list of all available options, see the man page of `rcpp_mcscanx_file()`.

Value

Paths to .collinearity files.
Examples

```r
# Load data
data(proteomes)
data(blast_list)
data(annotation)

# Get DIAMOND and processed annotation lists
blast_inter <- blast_list[2]
annotation <- process_input(proteomes, annotation)$annotation

# Detect interspecies synteny
intersyn <- interspecies_synteny(blast_inter, annotation)
```

---

intraspecies_synteny  

**Detect intraspecies synteny**

---

Description

Detect intraspecies synteny

Usage

```r
intraspecies_synteny(
    blast_intra = NULL,
    annotation = NULL,
    intra_dir = file.path(tempdir(), "intra"),
    anchors = 5,
    max_gaps = 25,
    is_pairwise = TRUE,
    verbose = FALSE,
    bp_param = BiocParallel::SerialParam(),
    ...
)
```

Arguments

- **blast_intra**: A list of BLAST/DIAMOND data frames for intraspecies comparisons as returned by `run_diamond()`.
- **annotation**: A processed GRangesList or CompressedGRangesList object as returned by `process_input()`.
- **intra_dir**: Path to output directory where .collinearity files will be stored.
- **anchors**: Numeric indicating the minimum required number of genes to call a syntenic block. Default: 5.
- **max_gaps**: Numeric indicating the number of upstream and downstream genes to search for anchors. Default: 25.
- **is_pairwise**: Logical indicating if only pairwise blocks should be reported. Default: TRUE.
verbose Logical indicating if log messages should be printed on screen. Default: FALSE.
bp_param BiocParallel back-end to be used. Default: BiocParallel::SerialParam().
... Any additional arguments to the MCScanX algorithm. For a complete list of all available options, see the man page of rcpp_mcscanx_file().

Value

Paths to .collinearity files.

Examples

# Load data
data(scerevisiae_annot)
data(scerevisiae_diamond)

# Detect intragenome synteny
intra_syn <- intraspecies_synteny(
    scerevisiae_diamond, scerevisiae_annot
)

iqtree_is_installed Check if IQTREE is installed

Description

Check if IQTREE is installed

Usage

iqtree_is_installed()

Value

Logical indicating whether IQTREE is installed or not.

Examples

iqtree_is_installed()
### iqtree_version

*Get IQ-TREE version*

**Description**

Get IQ-TREE version

**Usage**

iqtree_version()

**Value**

Numeric indicating IQ-TREE version, with either 1 or 2.

**Examples**

iqtree_version()

---

### last_is_installed

*Check if last is installed*

**Description**

Check if last is installed

**Usage**

last_is_installed()

**Value**

Logical indicating whether last is installed or not.

**Examples**

last_is_installed()
**network**

| network | Synten network of BUSCO genes for 25 eudicot species |

**Description**

Data obtained from Zhao & Schranz, 2019.

**Usage**

```r
data(network)
```

**Format**

An edgelist (i.e., a 2-column data frame with node 1 in column 1 and node 2 in column 2).

**References**


**Examples**

```r
data(network)
```

---

**parse_collinearity**

Parse .collinearity files obtained with MCScan

**Description**

The .collinearity files can be obtained with intraspecies_synteny and interspecies_synteny, which execute a native version of the MCScan algorithm.

**Usage**

```r
parse_collinearity(collinearity_paths = NULL, as = "anchors")
```

**Arguments**

- `collinearity_paths`  
  Character vector of paths to .collinearity files.

- `as`  
  Character specifying what to extract. One of "anchors" (default), "blocks", or "all".
phylogenomic_profile

Value

If `as` is "anchors", a data frame with variables "Anchor1", and "Anchor2". If `as` is "blocks", a data frame with variables "Block", "Block_score", "Chr", and "Orientation". If `as` is "all", a data frame with all aforementioned variables, which indicate:

- **Block** Numeric, syntenic block ID
- **Block_score** Numeric, score of syntenic block.
- **Chr** Character, query and target chromosome of the syntenic block formatted as "&".
- **Orientation** Character, the orientation of genes within blocks, with "plus" indicating that genes are in the same direction, and "minus" indicating that genes are in opposite directions.
- **Anchor1** Character, gene ID of anchor 1.
- **Anchor2** Character, gene ID of anchor 2.

Examples

```r
collinearity_paths <- system.file(
  "extdata", "Scerevisiae.collinearity", package = "syntenet"
)
net <- parse_collinearity(collinearity_paths)

phylogenomic_profile(clusters = NULL)
```

Description

Perform phylogenomic profiling for syntenic network clusters

Usage

```r
phylogenomic_profile(clusters = NULL)
```

Arguments

- `clusters` A 2-column data frame with variables `Gene` and `Cluster` as returned by `cluster_network`.

Value

A matrix of i rows and j columns containing the number of genes in cluster i for each species j. The number of rows is equal to the number of clusters in `clusters`, and the number of columns is equal to the number of species in `clusters`.

Examples

```r
data(clusters)
profiles <- phylogenomic_profile(clusters)
```
**plot_network**

**Description**

Plot network

**Usage**

```r
plot_network(
    network = NULL,
    clusters = NULL,
    cluster_id = NULL,
    color_by = "cluster",
    interactive = FALSE,
    dim_interactive = c(600, 600)
)
```

**Arguments**

- `network` The synteny network represented as an edge list, which is a 2-column data frame with each member of the anchor pair in a column.
- `clusters` A 2-column data frame with the variables **Gene** and **Cluster** representing gene ID and cluster ID, respectively, exactly as returned by `cluster_network`.
- `cluster_id` Character scalar or vector with cluster ID. If more than one cluster is passed as input, clusters are colored differently.
- `color_by` Either "cluster" or a 2-column data frame with gene IDs in the first column and variable to be used for coloring (e.g., taxonomic information) in the second column.
- `interactive` Logical scalar indicating whether to display an interactive network or not. Default: FALSE.
- `dim_interactive` Numeric vector of length 2 with the window dimensions of the interactive plot. If `interactive` is set to FALSE, this parameter is ignored.

**Value**

A `ggplot` object with the network.

**Examples**

```r
data(network)
data(clusters)
# Option 1: 1 cluster
cluster_id <- 25
plot_network(network, clusters, cluster_id)
```
# Option 2: 2 clusters
cluster_id <- c(25, 1089)
plot_network(network, clusters, cluster_id)

# Option 3: custom annotation for coloring
"Lang", "car", "pmu", "ppe", "pbr", "mdo", "roc", "fve",
"Mnot", "Zjuj", "jcu", "mes", "roc", "lus", "ptr")

species_annotation <- data.frame(
  Species = species_order,
  Family = c(rep("Fabaceae", 11), rep("Rosaceae", 6),
            "Moraceae", "Rhamnaceae", rep("Euphorbiaceae", 3),
            "Linaceae", "Salicaceae")
)

genes <- unique(c(network$node1, network$node2))
gene_df <- data.frame(
  Gene = genes,
  Species = unlist(lapply(strsplit(genes, "_"), head, 1))
)

gene_df <- merge(gene_df, species_annotation)[, c("Gene", "Family")]
plot_network(network, clusters, cluster_id = 25, color_by = gene_df)

---

plot_profiles  
**Plot a heatmap of phylogenomic profiles**

### Description

Plot a heatmap of phylogenomic profiles

### Usage

```r
plot_profiles(
  profile_matrix = NULL,
  species_annotation = NULL,
  palette = "Greens",
  dist_function = stats::dist,
  dist_params = list(method = "euclidean"),
  clust_function = stats::hclust,
  clust_params = list(method = "ward.D"),
  cluster_species = FALSE,
  show_colnames = FALSE,
  discretize = TRUE,
  ...
)
```
### Arguments

**profile_matrix**  
A matrix of phylogenomic profiles obtained with `phylogenomic_profile`.

**species_annotation**  
A 2-column data frame with species IDs in the first column (same as column names of profile matrix), and species annotation (e.g., higher-level taxonomic information) in the second column.

**palette**  
A character vector of colors or a character scalar with the name of an RColorBrewer palette. Default: "RdYlBu".

**dist_function**  
Function to use to calculate a distance matrix for synteny clusters. Popular examples include `stats::dist`, `labdsv::dsvdis`, and `vegan::vegdist`. Default: `stats::dist`.

**dist_params**  
A list with parameters to be passed to the function specified in parameter `dist_function`. Default: list(method = "euclidean").

**clust_function**  
Function to use to cluster the distance matrix returned by the function specified in `dist_function`. Examples include `stats::hclust` and `Rclusterpp::Rclusterpp.hclust`. Default: `stats::hclust`.

**clust_params**  
A list with additional parameters (if any) to be passed to the function specified in parameter `clust_function`. Default: list(method = "ward.D").

**cluster_species**  
Either a logical scalar (TRUE or FALSE) or a character vector with the order in which species should be arranged. TRUE or FALSE indicate whether hierarchical clustering should be applied to rows (species). Ideally, the character vector should contain the order of species in a phylogenetically meaningful way. If users pass a named vector, vector names will be used to rename species. If users have a species tree, they can read it with `treeio::read.tree()`, plot it with `ggtree::ggtree()`, and get the species order from the ggtree object with `ggtree::get_taxa_name()`. Default: FALSE.

**show_colnames**  
Logical indicating whether to show column names (i.e., cluster IDs) or not. Showing cluster IDs can be useful when visualizing a small subset of them. When visualizing all clusters, cluster IDs are impossible to read. Default: FALSE.

**discretize**  
Logical indicating whether to discretize clusters in 4 categories: 0, 1, 2, and 3+. If FALSE, counts will be log2 transformed. Default: TRUE.

...  
Additional parameters to `pheatmap::pheatmap()`.

### Value

A pheatmap object.

### Examples

```r
data(clusters)
profile_matrix <- phylogenomic_profile(clusters)
species_order <- c(
  "Lang", "car", "pmu", "ppe", "pbr", "mdo", "roc", "fve",
  "Mnot", "Zjuj", "jcu", "mes", "rco", "lus", "ptr"
```

```r
```

names(species_order) <- species_names

species_annotation <- data.frame(
  Species = species_order,
)

p <- plot_profiles(profile_matrix, species_annotation, cluster_species = species_order)

p <- plot_profiles(profile_matrix, species_annotation, cluster_species = species_order, discretize = FALSE)

---

**process_input**

**Process sequence data**

**Description**

Process sequence data

**Usage**

process_input(
  seq = NULL,
  annotation = NULL,
  gene_field = "gene_id",
  filter_annotation = FALSE
)

**Arguments**

- **seq**  
  A list of AAStringSet objects, each list element containing protein sequences for a given species. This list must have names (not NULL), and names of each list element must match the names of list elements in `annotation`.

- **annotation**  
  A GRangesList, CompressedGRangesList, or list of GRanges with the annotation for the sequences in `seq`. This list must have names (not NULL), and names of each list element must match the names of list elements in `seq`.

- **gene_field**  
  Character, name of the column in the GRanges objects that contains gene IDs. Default: "gene_id".
filter_annotation

Logical indicating whether annotation should be filtered to keep only genes that are also in seq. This is particularly useful if users want to remove information on non-protein coding genes from annotation, since such genes are typically not present in sets of whole-genome protein sequences. Default: FALSE.

Details

This function processes the input sequences and annotation to:

1. Remove whitespace and anything after it in sequence names (i.e., names(seq[[x]]), which is equivalent to FASTA headers), if there is any.
2. Add a unique species identifier to sequence names. The species identifier consists of the first 3-5 strings of the element name. For instance, if the first element of the seq list is named "Athaliana", each sequence in it will have an identifier "Atha_" added to the beginning of each gene name (e.g., Atha_AT1G01010).
3. If sequences have an asterisk (*) representing stop codon, remove it.
4. Add a unique species identifier (same as above) to gene and chromosome names of each element of the annotation GRangesList/CompressedGRangesList.
5. Filter each element of the annotation GRangesList/CompressedGRangesList to keep only seqnames, ranges, and gene ID.

Value

A list of 2 elements:

- seq The processed list of AAStringSet objects from seq.
- annotation The processed GRangesList or CompressedGRangesList object from annotation.

Examples

data(translation)
data(proteomes)
seq <- proteomes
clean_data <- process_input(seq, annotation)

profiles2phylip

Save the transposed binary profiles matrix to a file in PHYLIP format

Description

Save the transposed binary profiles matrix to a file in PHYLIP format

Usage

profiles2phylip(transposed_profiles = NULL, outdir = tempdir())
Arguments

transposed_profiles
A binary and transposed profile matrix. The profile matrix can be obtained with phylogenomic_profile().

outdir
Path to output directory. By default, files are saved in a temporary directory, so they will be deleted when the R session closes. If you want to keep the files, specify a custom output directory.

Value
Character specifying the path to the PHYLIP file.

Examples

data(clusters)
profile_matrix <- phylogenomic_profile(clusters)
tmat <- binarize_and_transpose(profile_matrix)
profiles2phylip(tmat)

-------
proteomes
Filtered proteomes of Ostreococcus sp. species
-------

Description
Data obtained from Pico-PLAZA 3.0. Only the translated sequences of primary transcripts were included, and only genes from chromosomes 1, 2, and 3.

Usage

data(proteomes)

Format
A list of AAStringSet objects containing the elements Olucimarinus, Osp_RCC809, and Otauri.

References

Examples

data(proteomes)
Description

MCSCanX provides a clustering module for viewing the relationship of collinear segments in multiple genomes (or heavily redundant genomes). It takes the predicted pairwise segments from dynamic programming (DAGchainer in particular) and then tries to build consensus segments from a set of related, overlapping segments.

Usage

rcpp_mcscanx_file(
  blast_file,
  gff_file,
  prefix = "out",
  outdir = "",
  match_score = 50L,
  gap_penalty = -1L,
  match_size = 5L,
  e_value = 1e-05,
  max_gaps = 25L,
  overlap_window = 5L,
  is_pairwise = FALSE,
  in_synteny = 0L,
  species_id_length = 3L,
  verbose = FALSE
)

Arguments

- **blast_file**: Character indicating the path to the BLAST/DIAMOND output file.
- **gff_file**: Character indicating the path to the "gff" file, which is a tab-delimited file with 4 columns indicating the chromosome name, gene id, gene start position, and gene end position, respectively.
- **prefix**: Character indicating the prefix to output files. Default: "out".
- **outdir**: Character indicating the path to the output directory. Default: "".
- **match_score**: Numeric indicating the match score. Default: 50.
- **gap_penalty**: Numeric indicating the gap penalty. Default: -1.
- **match_size**: Numeric indicating the minimum number of genes required to call synteny. Default: 5.
- **e_value**: Numeric indicating the minimum e-value allowed. Default: 1e-5.
- **max_gaps**: Numeric indicating the maximum number of gaps between genes allowed. The unit measure of gaps is number of genes, so max_gaps = 20 indicates that a maximum of 20 genes can exist between two homologous genes for synteny to be called. Default: 25.
read_diamond

**overlap_window**  Numeric indicating the overlap window. Default: 5.

**is_pairwise**  Logical indicating whether only pairwise blocks should be reported. Default: FALSE.

**in_synteny**  Numeric indicating the patterns of collinear blocks, where 0 indicates intra and interspecies comparisons, 1 indicates intraspecies comparisons, and 2 indicates interspecies comparisons. Default: 0.

**species_id_length**  Integer indicating the length of the species IDs. Default: 3. 0: intra- and inter-species (default); 1: intra-species; 2: inter-species

**verbose**  Logical indicating whether to print progress messages to the screen. Default: FALSE.

**Value**

NULL, and a `.collinearity` file is created in the directory specified in `outdir`.

**Author(s)**

Kristian K Ullrich and Fabricio Almeida-Silva

**References**


---

**read_diamond**  *Read DIAMOND/BLAST tables as a list of data frames*

**Description**

Read DIAMOND/BLAST tables as a list of data frames

**Usage**

```r
read_diamond(diamond_dir = NULL)
```

**Arguments**

**diamond_dir**  Path to directory containing the tabular output of DIAMOND or similar programs (e.g., BLAST).

**Value**

A list of data frames with the tabular DIAMOND output.
Examples

```r
# Path to output directory
diamond_dir <- system.file("extdata", package = "syntenet")

# Read output
l <- read_diamond(diamond_dir)
```

---

**Description**

Wrapper to run DIAMOND from an R session

**Usage**

```r
run_diamond(
  seq = NULL,
  top_hits = 5,
  verbose = FALSE,
  outdir = tempdir(),
  threads = NULL,
  compare = "all",
  ...
)
```

**Arguments**

- `seq`: A processed list of AAStrongSet objects as returned by `process_input()`.
- `top_hits`: Number of top hits to keep in DIAMOND search. Default: 5.
- `verbose`: Logical indicating if progress messages should be printed. Default: FALSE.
- `outdir`: Output directory for DIAMOND results. By default, output files are saved to a temporary directory.
- `threads`: Number of threads to use. Default: let DIAMOND auto-detect and use all available virtual cores on the machine.
- `compare`: Character scalar indicating which comparisons should be made when running DIAMOND. Possible modes are "all" (all-vs-all comparisons), "intraspecies" (intraspecies comparisons only), or "interspecies" (interspecies comparisons only). Alternatively, users can pass a 2-column data frame as input with the names of species to be compared.
- `...`: Any additional arguments to `diamond blastp`.

**Value**

A list of data frames containing DIAMOND’s tabular output for each pairwise combination of species. For n species, the list length will be $n^2$. 
Examples

```r
data(proteomes)
data(annotation)
seq <- process_input(proteomes, annotation)$seq[1:2]
if(diamond_is_installed()) {
  diamond_results <- run_diamond(seq)
}
```

run_last

Wrapper to run last from an R session

Description

Wrapper to run last from an R session

Usage

```r
run_last(
  seq = NULL,
  verbose = FALSE,
  outdir = tempdir(),
  threads = 1,
  compare = "all",
  lastD = 1e+06,
  ...
)
```

Arguments

- **seq**: A processed list of AAStringSet objects as returned by `process_input()`.
- **verbose**: Logical indicating if progress messages should be printed. Default: FALSE.
- **outdir**: Output directory for last results. By default, output files are saved to a temporary directory.
- **threads**: Number of threads to use. Default: 1.
- **compare**: Character scalar indicating which comparisons should be made when running last. Possible modes are "all" (all-vs-all comparisons), "intraspecies" (intraspecies comparisons only), or "interspecies" (interspecies comparisons only). Alternatively, users can pass a 2-column data frame as input with the names of species to be compared.
- **lastD**: last option D: query letters per random alignment. Default: 1e6.
- **...**: Any additional arguments to `lastal`.

Value

A list of data frames containing last’s tabular output for each pairwise combination of species. For n species, the list length will be $n^2$. 

Examples

data(proteomes)
data(annotations)
seq <- process_input(proteomes, annotation)$seq[1:2]
if(last_is_installed()) {
  last_results <- run_last(seq)
}

scerevisiae_annot  Genome annotation of the yeast species S. cerevisiae

Description
Data obtained from Ensembl Fungi. Only annotation data for primary transcripts were included.

Usage

data(scerevisiae_annot)

Format
A GRangesList as returned by process_input() containing the element **Scerevisiae**.

Examples

data(scerevisiae_annot)

scerevisiae_diamond  Intraspecies DIAMOND output for S. cerevisiae

Description
List obtained with run_diamond().

Usage

data(scerevisiae_diamond)

Format
A list of data frames (length 1) containing the whole paranome of S. cerevisiae resulting from intragenome similarity searches.

Examples

data(scerevisiae_diamond)
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