

# Package ‘scanMiR’

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**Description** A set of tools for working with miRNA affinity models (KdModels), efficiently scanning for miRNA binding sites, and predicting target repression. It supports scanning using miRNA seeds, full miRNA sequences (enabling 3' alignment) and KdModels, and includes the prediction of slicing and TDMD sites. Finally, it includes utility and plotting functions (e.g. for the visual representation of miRNA-target alignment).

**License** GPL-3

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---

aggregateMatches	<i>aggregateMatches</i>
------------------	-------------------------

---

**Description**

Aggregates miRNA binding sites with log<sub>k</sub>d values to predict transcript repression. See the vignette for more detail.

**Usage**

```
aggregateMatches(
  m,
  a = 0.007726,
  b = 0.5735,
  c = 0.181,
  p3 = 0.04403,
  coef_utr = -0.28019,
  coef_orf = -0.08622,
  p3.range = c(3L, 8L),
  keepSiteInfo = TRUE,
  toInt = FALSE,
  BP = NULL
)
```

**Arguments**

m	A GRanges or data.frame of matches as returned by 'findSeedMatches'.
a	The relative concentration of unbound AGO-miRNA complexes.
b	Factor specifying the additional repression by a single bound AGO.
c	Penalty for sites that are found within the ORF region.
p3	Factor specifying additional repression due to 3p alignment.
coef_utr	Factor specifying additional repression due to UTR length.
coef_orf	Factor specifying additional repression due to ORF length.
p3.range	Range used for 3p alignment.
keepSiteInfo	Logical; whether to return information about site types (default = TRUE). Ignored if 'm' does not contain 'log_kd' values
toInt	Logical; whether to convert repression scores to integers (default = FALSE).
BP	Pass 'BiocParallel::MulticoreParam(ncores, progressbar=TRUE)' to enable multithreading. Note that in addition, 'aggregateMatches' uses the <a href="#">data.table</a> package, which is often set to use multi-threading by default (which would be multiplied by threads determined by 'BP'). See <a href="#">setDTthreads</a> for more information.

**Value**

a data.frame containing aggregated repression values and/or information about the numbers and types of matches

**Examples**

```
# we create mock RNA sequences and seeds:
seqs <- getRandomSeq(n=10)

# load sample KdModel
data(SampleKdModel)

# find matches
matches <- findSeedMatches(seqs, SampleKdModel)

# aggregate matches
aggregateMatches(matches)
```

---

assignKdType

*assignKdType*


---

**Description**

Assigns a log<sub>k</sub>d and match type to a set of matched sequences.

**Usage**

```
assignKdType(x, mod, mer8 = NULL)
```

**Arguments**

x	A vector of matched sequences, each of 12 nucleotides
mod	An object of class 'KdModel'
mer8	The optional set of 8mers included in the model (for internal use; can be reconstructed from the model).

**Value**

A data.frame with one row for each element of 'x', and the columns 'type' and 'log\_kd'. To save space, the reported log\_kd is multiplied by 1000, rounded and saved as an integer.

**Examples**

```
data(SampleKdModel)
assignKdType(c("CTAGCATTAAGT", "ACGTACGTACGT"), SampleKdModel)
```

---

conservation	<i>conservation</i>
--------------	---------------------

---

**Description**

conservation

**Usage**

```
conservation(x)
```

**Arguments**

x	A KdModelList, or a KdModel
---	-----------------------------

**Value**

A vector of the conservation status for each miRNA

**Examples**

```
data(SampleKdModel)
conservation(SampleKdModel)
```

---

dummyKdData	<i>Create dummy log_kd per 12-mer data</i>
-------------	--

---

**Description**

Create dummy log\_kd per 12-mer data

**Usage**

```
dummyKdData(mod = NULL)
```

**Arguments**

mod                   Optional model from which to create the dummy data

**Value**

A data.frame with 12-mers and log\_kds

**Examples**

```
kd <- dummyKdData()
```

---

findSeedMatches	<i>Predicting and characterizing miRNA binding sites</i>
-----------------	--

---

**Description**

‘findSeedMatches’ takes a set of sequences and a set of miRNAs (given either as target seeds, mature miRNA sequences, or a [KdModelList](#)).

**Usage**

```
findSeedMatches(  
  seqs,  
  seeds,  
  shadow = 0L,  
  onlyCanonical = FALSE,  
  maxLogKd = c(-1, -1.5),  
  keepMatchSeq = FALSE,  
  minDist = 7L,  
  p3.extra = FALSE,  
  p3.params = list(maxMirLoop = 5L, maxTargetLoop = 9L, maxLoopDiff = 4L, mismatch =  
    TRUE),  
  agg.params = .defaultAggParams(),  
  ret = c("GRanges", "data.frame", "aggregated"),
```

```

BP = NULL,
verbose = NULL,
n_seeds = NULL,
useTmpFiles = FALSE,
keepTmpFiles = FALSE
)

```

## Arguments

seqs	A character vector or 'DNAStrngSet' of DNA sequences in which to look.
seeds	A character vector of 7-nt seeds to look for. If RNA, will be reversed and complemented before matching. If DNA, they are assumed to be the target sequence to look for. Alternatively, a list of objects of class 'KdModel' or an object of class 'KdModelList' can be given.
shadow	Integer giving the shadow, i.e. the number of nucleotides hidden at the beginning of the sequence (default 0).
onlyCanonical	Logical; whether to restrict the search only to canonical binding sites.
maxLogKd	Maximum log_kd value to keep. This has a major impact on the number of sites returned, and hence on the memory requirements. Set to Inf to disable ( <code>_not_recommended</code> when running large scans!).
keepMatchSeq	Logical; whether to keep the sequence (including flanking dinucleotides) for each seed match (default FALSE).
minDist	Integer specifying the minimum distance between matches of the same miRNA (default 7). Closer matches will be reduced to the highest-affinity. To disable the removal of overlapping features, use <code>'minDist=-Inf'</code> .
p3.extra	Logical; whether to keep extra information about 3' alignment. Disable (default) this when running large scans, otherwise you might hit your system's memory limits.
p3.params	Named list of parameters for 3' alignment with slots <code>'maxMirLoop'</code> (integer, default = 5), <code>'maxTargetLoop'</code> (integer, default = 9), <code>'maxLoopDiff'</code> (integer, default = 4), and <code>'mismatch'</code> (logical, default = TRUE).
agg.params	A named list with slots <code>'a'</code> , <code>'b'</code> , <code>'c'</code> , <code>'p3'</code> , <code>'coef_utr'</code> , <code>'coef_orf'</code> and <code>'keepSite-Info'</code> indicating the parameters for the aggregation. Ignored if <code>'ret!="aggregated"</code> . For further details see documentation of <code>'aggregateMatches'</code> .
ret	The type of data to return, either "GRanges" (default), "data.frame", or "aggregated" (aggregates affinities/sites for each seed-transcript pair).
BP	Pass <code>'BiocParallel::MulticoreParam(ncores, progressBar=TRUE)'</code> to enable multithreading.
verbose	Logical; whether to print additional progress messages (default on if not multithreading)
n_seeds	Integer; the number of seeds that are processed in parallel to avoid memory issues.
useTmpFiles	Logical; whether to write results for single miRNAs in temporary files (ignored when scanning for a single seed). Alternatively, <code>'useTmpFiles'</code> can be a character vector of length 1 indicating the path to the directory in which to write temporary files.

`keepTmpFiles` Logical; whether to keep the temporary files at the end of the process; ignored if `'useTmpFiles=FALSE'`. Temporary files are removed only upon successful completion of the function, meaning that they will not be deleted in case of errors.

### Value

A GRanges of all matches. If `'seeds'` is a `'KdModel'` or `'KdModelList'`, the `'log_kd'` column will report the  $\ln(Kd)$  multiplied by 1000, rounded and saved as an integer. If `'ret!="GRanges'`, returns a data.frame.

### Examples

```
# we create mock RNA sequences and seeds:
seqs <- getRandomSeq(n=10)
seeds <- c("AAACCAC", "AAACCUU")
findSeedMatches(seqs, seeds)
```

---

get3pAlignment      *Finds 3' complementary binding of a miRNA*

---

### Description

Performs a local alignment of the miRNA 3' sequence (determined by `'mir3p.start'`) on given the given sequences.

### Usage

```
get3pAlignment(
  seqs,
  mirseq,
  mir3p.start = 9L,
  allow.mismatch = TRUE,
  maxMirLoop = 5L,
  maxTargetLoop = 9L,
  maxLoopDiff = 4L,
  TGsub = TRUE,
  siteType = NULL
)
```

### Arguments

`seqs` A set of sequences in which to look for 3' matches (i.e. upstream of the seed match)

`mirseq` The sequence of the mature miRNA

`mir3p.start` The position in `'mirseq'` in which to start looking

`allow.mismatch` Logical; whether to allow mismatches

maxMirLoop	Maximum miRNA loop size
maxTargetLoop	Maximum target loop size
maxLoopDiff	Maximum size difference between miRNA and target loops
TGsub	Logical; whether to allow T/G substitutions.
siteType	The optional type of seed-complementarity, as returned by <a href="#">getMatchTypes</a> . This is needed to identify slicing/TDMD sites. If given, should be a vector of the same length as 'seqs'.

**Value**

A data.frame with one row for each element of 'seqs', indicating the size of the miRNA bulge, the size of the target mRNA bulge, the number of mismatches at the 3' end, and the partial 3' alignment score (i.e. roughly the number of consecutive matching nucleotides)

**Examples**

```
get3pAlignment(seqs="NNAGTGTGCCATNN", mirseq="TGGAGTGTGACAATGGTGTTTG")
```

---

`get8merRange`

*get8merRange*

---

**Description**

Returns the minimum and maximum 8-mer log-kd values

**Usage**

```
get8merRange(mod)
```

**Arguments**

mod                    A 'KdModel'

**Value**

A numeric vector of length two

**Examples**

```
data("SampleKdModel")
get8merRange(SampleKdModel)
```



---

 getKdModel

*getKdModel*


---

**Description**

getKdModel

**Usage**

```
getKdModel(kd, mirseq = NULL, name = NULL, conservation = NA_integer_, ...)
```

**Arguments**

kd	A data.frame containing the log_kd per 12-mer sequence, or the path to a text/csv file containing such a table. Should contain the columns 'log_kd', '12mer' (or 'X12mer'), and eventually 'mirseq' (if the 'mirseq' argument is NULL) and 'mir' (if the 'name' argument is NULL).
mirseq	The miRNA (cDNA) sequence.
name	The name of the miRNA.
conservation	The conservation level of the miRNA. See 'scanMiR:::conservation_levels()' for possible values.
...	Any additional information to be saved with the model.

**Value**

An object of class 'KdModel'.

**Examples**

```
kd <- dummyKdData()
mod <- getKdModel(kd=kd, mirseq="TTAATGCTAATCGTGATAGGGTT", name="my-miRNA")
```

---

 getKmers

*getKmers*


---

**Description**

Returns all combinations of 'n' elements of 'from'

**Usage**

```
getKmers(n = 4, from = c("A", "C", "G", "T"))
```

**Arguments**

n	Number of elements
from	Letters sampled

**Value**

A character vector

**Examples**

```
getKmers(3)
```

---

<code>getMatchTypes</code>	<i>getMatchTypes</i>
----------------------------	----------------------

---

**Description**

Given a seed and a set of sequences matching it, returns the type of match.

**Usage**

```
getMatchTypes(x, seed, checkWobble = TRUE)
```

**Arguments**

x	A character vector of short sequences.
seed	A 7 or 8 nucleotides string indicating the seed (5' to 3' sequence of the target RNA). If of length 7, an "A" will be appended.
checkWobble	Whether to flag wobbled sites

**Value**

A factor of match types.

**Examples**

```
x <- c("AACACTCCAG", "GACACTCCGC", "GTACTCCAT", "ACGTACGTAC")
getMatchTypes(x, seed="ACACTCCA")
```

---

getRandomSeq	<i>getRandomSeq</i>
--------------	---------------------

---

**Description**

Produces a random sequence of the given letters

**Usage**

```
getRandomSeq(length = 3000, alphabet = c("A", "C", "G", "T"), n = 1)
```

**Arguments**

length	Length of the sequence
alphabet	Letters from which to sample
n	The number of sequences to generate

**Value**

A character vector of length 1

**Examples**

```
getRandomSeq(100)
```

---

getSeed8mers	<i>getSeed8mers</i>
--------------	---------------------

---

**Description**

Generates all possible 8mers with 4 consecutive and positioned matches to a given seed.

**Usage**

```
getSeed8mers(seed, addNs = FALSE)
```

**Arguments**

seed	The miRNA seed (target DNA sequence), a character vector of length 8 (if of length 7, a "A" will be added on the right)
addNs	Logical; whether to include 8mers with one flanking N

**Value**

A vector of 1024 8mers.

**Examples**

```
head(getSeed8mers("ACACTCCA"))
```

---

KdModel

*miRNA affinity models*

---

**Description**

Methods for the [KdModel](#) class

**Usage**

```
## S4 method for signature 'KdModel'  
show(object)
```

```
## S4 method for signature 'KdModel'  
summary(object)
```

```
## S4 method for signature 'KdModel'  
c(x, ...)
```

**Arguments**

object, x, ... An object of class [KdModel](#)

**Value**

Depends on the method.

**See Also**

[KdModel](#), [KdModelList](#)

**Examples**

```
data(SampleKdModel)  
SampleKdModel  
summary(SampleKdModel)
```

---

KdModelList-class      *KdModelList*

---

**Description**

KdModelList

**Usage**

```
KdModelList(..., description = NULL, makeUnique = FALSE)
```

**Arguments**

...                    Any number of [KdModel](#) objects or lists thereof.  
description            A description for the collection.  
makeUnique            Logical; whether to rename models if names are duplicated.

**Value**

A KdModelList

**Examples**

```
data(SampleKdModel)
mods <- KdModelList(SampleKdModel, SampleKdModel, makeUnique = TRUE)
mods
```

---

KdModelList-methods      *Methods for the [KdModelList](#) classes*

---

**Description**

Methods for the [KdModelList](#) classes

**Usage**

```
## S4 method for signature 'KdModelList'
summary(object)

## S4 method for signature 'KdModelList,ANY'
x[i, j = NULL, ..., drop = TRUE]
```

**Arguments**

object, x            An object of class [KdModelList](#)  
i                    the index of item(s) to select  
j, drop, ...        ignored

**Value**

Depends on the method.

**See Also**

[KdModel](#), [KdModelList](#)

**Examples**

```
# create a KdModelList :
data(SampleKdModel)
kml <- KdModelList( SampleKdModel, SampleKdModel, makeUnique=TRUE )
summary(kml)
kml[1] # returns a KdModelList
kml[[2]] # returns a KdModel
conservation(kml)
```

---

plotKdModel

*plotKdModel*

---

**Description**

Plots the summary of an affinity model.

**Usage**

```
plotKdModel(mod, what = c("both", "seeds", "logo"), n = 10)
```

**Arguments**

mod	A 'KdModel'
what	Either 'seeds', 'logo', or 'both' (default). 'logo' and 'both'
n	The number of top 7-mers to plot (when 'what='seeds')

**Details**

'what='seeds' plots the  $-\log(Kd)$  values of the top 'n' 7-mers (including both canonical and non-canonical sites), with or without the final "A" vis-a-vis the first miRNA nucleotide. 'what='logo' plots a 'seqLogo' (requires the [seqLogo](<https://bioconductor.org/packages/release/bioc/html/seqLogo.html>) package) showing the nucleotide-wise information content and preferences for all 12-mers (centered around the seed). 'what="both"' plots both.

**Value**

If 'what="logo"', returns nothing a plots a position weight matrix. Otherwise returns a ggplot.

**Examples**

```
data(SampleKdModel)
plotKdModel(SampleKdModel, what="seeds")
```

---

```
removeOverlappingRanges
```

*removeOverlappingRanges*

---

**Description**

Removes elements from a GRanges that overlap (or are within a given distance of) other elements higher up in the list (i.e. assumes that the ranges are sorted in order of priority). The function handles overlaps between more than two ranges by successively removing those that overlap higher-priority ones.

**Usage**

```
removeOverlappingRanges(  
  x,  
  minDist = 7L,  
  retIndices = FALSE,  
  ignore.strand = FALSE  
)
```

**Arguments**

<code>x</code>	A GRanges, sorted by (decreasing) importance.
<code>minDist</code>	Minimum distance between ranges.
<code>retIndices</code>	Logical; whether to return the indices of entries to remove, rather than the filtered GRanges.
<code>ignore.strand</code>	Logical. Whether the strand of the input ranges should be ignored or not.

**Value**

A filtered GRanges, or an integer vector of indices to be removed if `'retIndices==TRUE'`.

**Examples**

```
library(GenomicRanges)
gr <- GRanges(seqnames=rep("A",4), IRanges(start=c(10,25,45,35), width=6))
removeOverlappingRanges(gr, minDist=7)
```

---

SampleKdModel	<i>Example KdModel (hsa-miR-155-5p)</i>
---------------	---

---

**Description**

'KdModel' for hsa-miR-155-5p, based on Kd predictions from the CNN of [McGeary, Lin et al. (2019)](<https://dx.doi.org/10.1126/science.aav1741>).

**Value**

a 'KdModel' object

**Examples**

```
data(SampleKdModel)
SampleKdModel
```

---

SampleTranscript	<i>Example transcript sequence</i>
------------------	------------------------------------

---

**Description**

An artificial transcript sequence used for examples.

**Value**

a named character vector of length 1.

---

viewTargetAlignment	<i>viewTargetAlignment</i>
---------------------	----------------------------

---

**Description**

viewTargetAlignment



**Usage**

```
viewTargetAlignment(
  m,
  miRNA,
  seqs = NULL,
  flagBulgeMatches = FALSE,
  maxBulgeSize = 9L,
  maxBulgeDiff = 4L,
  min3pMatch = 3L,
  hideSingletons = FALSE,
  UGsub = TRUE,
  ...,
  outputType = c("print", "data.frame", "plot", "ggplot")
)
```

**Arguments**

<code>m</code>	A GRanges of length 1 giving the information for a given match, as produced by <code>findSeedMatches</code> .
<code>miRNA</code>	A miRNA sequence, or a <code>KdModel</code> object of the miRNA corresponding to the match in 'm'; alternatively, a <code>KdModelList</code> including the model.
<code>seqs</code>	The sequences corresponding to the seqnames of 'm'. Not needed if 'm' contains the target sequences.
<code>flagBulgeMatches</code>	Logical; whether to flag matches inside the bulge (default FALSE)
<code>maxBulgeSize</code>	The maximum bulge size to consider (default none)
<code>maxBulgeDiff</code>	The maximum difference between miRNA and target bulges
<code>min3pMatch</code>	The minimum 3' alignment for any to be plotted
<code>hideSingletons</code>	Logical; whether to hide isolated single base-pair matches
<code>UGsub</code>	Logical; whether to show U-G matches
<code>...</code>	Passed to 'text' if 'outputType="plot"'.
<code>outputType</code>	Either 'print' (default, prints to console), 'data.frame', or 'plot'.

**Value**

Returns nothing 'outputType="print"'. If 'outputType="data.frame"', returns a data.frame containing the alignment strings; if 'outputType="plot"' returns a 'ggplot' object.

**Examples**

```
data(SampleKdModel)
seq <- c(seq1="CGACCCCTATCACGTCCGCAGCATTAAT")
m <- findSeedMatches(seq, SampleKdModel, verbose=FALSE)
viewTargetAlignment(m, miRNA=SampleKdModel, seqs=seq)
```

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