Package ‘macat’

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**Title** MicroArray Chromosome Analysis Tool

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**Suggests** hgu95av2.db, stjudem

**Description** This library contains functions to investigate links between differential gene expression and the chromosomal localization of the genes. MACAT is motivated by the common observation of phenomena involving large chromosomal regions in tumor cells. MACAT is the implementation of a statistical approach for identifying significantly differentially expressed chromosome regions. The functions have been tested on a publicly available data set about acute lymphoblastic leukemia (Yeoh et al.Cancer Cell 2002), which is provided in the library 'stjudem'.

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Auxiliary Computation Functions

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Description

Auxiliary functions for internal usage by other MACAT functions. Normally not called by user.

Details

Internal auxiliary functions. Not called by user.

Author(s)

MACAT development team

See Also

evalScoring
**buildMACAT**

Create MACAT list from objects in workspace

---

**Description**

This is a wrapper around the preprocessedLoader function. Use it, when you want to build a MACAT-list structure from objects already in your workspace.

**Usage**

```
buildMACAT(matrix, chip, labels = NULL, chromLocObj = NULL)
```

**Arguments**

- `matrix`: expression matrix with rows=genes and columns=samples; Rownames have to match chip; Columnnames are not mandatory.
- `chip`: Identifier for used microarray
- `labels`: Classlabels for samples, has to have length=number of columns in matrix
- `chromLocObj`: Object of class chromLocation specifying the genomic position, each probe on the array is mapped to. If not provided, it is build in the function using annotate's function buildChromLocation.

**Details**

This is only a convenience wrapper around the function preprocessedLoader for the case, that you want to build a MACAT-list from objects in your workspace.

**Value**

A MACAT-list structure. For an example and a description of the format see data stjude in package 'stjudem'.

**Author(s)**

MACAT development team

**See Also**

preprocessedLoader, stjude in package 'stjudem'

**Examples**

```
X <- matrix(rnorm(200),nrow=20,ncol=10)
rownames(X) <- c('34916_s_at', '34917_at', '34462_at', '163_at', '35219_at',
                 '31641_s_at', '33300_at', '33301_g_at', '38950_r_at', '41249_at',
                 '294_s_at', '32004_s_at', '33299_at', '41243_at', '33341_at', '362_at',
                 '1918_at', '41499_at', '41500_at', '41282_s_at')
colnames(X) <- paste("Sample",1:10,sep="")
```
```r
ty <- rep(c("A","B"),c(5,5))
toy <- buildMACAT(X,"hgu95av2.db",y)
summary(toy)
```

**compute.sliding**

*Compute and plot smoothing of expression values or scores along the chromosome*

**Description**

'compute.sliding' computes a smoothing of the expression data or scores along the chromosome using the specified kernel function. This function is also used within the 'evalScoring' function. 'plotSliding' creates a plot of the smoothed expression values / scores.

**Usage**

```r
compute.sliding(data, chromosome, sample, kernel, kernelparams=NULL, step.width = 1e+06)
plotSliding(data, chromosome, sample, kernel, kernelparams=NULL, step.width=100000, ...)
```

**Arguments**

- **data**: A MACATData list holding the Expression values and gene locations
- **chromosome**: the chromosome to be smoothed
- **sample**: the sample (patient) whose expression values are smoothed
- **kernel**: a kernel function (one of rbf, kNN, basePairDistance or your own)
- **kernelparams**: a list of named parameters for the kernel (by default estimated from the data)
- **step.width**: the smoothing is computed stepwise every step.width basepairs (default is 100000)
- **...**: further graphical parameters passed on to `plot.default`

**Value**

For `compute.sliding`: a matrix of dimension (steps x 2) with in the first column the locations in basepairs where an interpolation is computed, and in the second column the smoothed values. `plotSliding` does not return anything and is merely called for its side-effect producing the plot.

**Author(s)**

MACAT development team

**See Also**

`kernelize, evalScoring`
discreteKernelize

Examples

data(stjd)
# just compute smoothed values:
smooth = compute.sliding(stjd, chromosome=3, sample=6, rbf,
    kernelparams=list(gamma=1/10^13))
# compute and plot smoothed values:
plotSliding(stjd, chromosome=3, sample=6, rbf,
            kernelparams=list(gamma=1/10^13),pch=20,
            main="Chromosome 3")

discreteKernelize Discretize and smooth expression values

Description

returns discretized kernelized expression values and saves them to a file if argument ‘saveToFile’ is TRUE. For details on discretization see discretize.

Usage

discreteKernelize(data, chrom, margin = 10, step.width = 1e+05, kernel = rbf,
    kernelparams = list(gamma = 1/10^13), saveToFile = FALSE)

Arguments

data MACATData Object
chrom chromosome to kernelize
margin symmetric quantile in percent
step.width size of the interpolation steps
kernel kernel function one of rbf, kNN, basePairDistance or your own
kernelparams list of named kernel parameters
saveToFile logical value indicating whether to write a flatfile or not; default is FALSE

Details

Filename of the flatfile is: discrete_kernelized_seq_margin_<margin>_chrom_<chrom>.py
where <margin> is the discretization parameter and <chrom> the name of the chromosome.

Value

discretized and kernelized expression matrix

Author(s)

The MACAT Development team
discretize

See Also

pydata.kernelizeAll

Examples

# load datapkgs("stjude")
# data(stjude)
data(stjd)
discretizedKernelized = discretizeKernelize(stjd, 13)

## discretize

Discretize expression values

Description

’discretize’ returns the discretized expression data for all chromosomes in chrom and all samples that have a label listed in label. Discretization is performed by comparing the value gene-wise (location-wise) with the symmetric upper and lower quantile given by margin (in percent margin/2 lower and upper quantile).

Usage

discretize(data, chrom, label, margin = 10)
discretizeChromosome(data, chrom, margin=10)
discretizeOne(data, chrom, sample, margin=10)

Arguments

data MACATData object
chron list of chromosomes
label list of labels
margin symmetric quantile in percent
sample the sample for which you want discretized expression data

Value

returns a discretized expression matrix for all genes on the chromosomes in ‘chrom’ and all samples that have a label in ‘label’.

Author(s)

MACAT development team

See Also

discretizeAll
discretize.tscores  

Discretize regularized t-scores

Description

discretize.tscores returns a discretized version of the scores in the MACATevalScoring object. Discretization is performed by comparing the value gene-wise (location-wise) with the symmetric upper and lower quantile given by margin (in percent margin/2 lower and upper quantile). discretizeAllClasses produces a flatfile readable by PYTHON.

Usage

discretize.tscores(scores)
discretizeAllClasses.tscores(data, chrom, nperms=10, kernel=rbf, kernelparams=NULL, step.width=100000)

Arguments

scores  
a MACATevalScoring object obtained from evalScoring

data  
a MACATData Object containing all expression values, geneLocations and labels (obtained from preprocessedLoader)

chrom  
chromosome that is discretized

nperms  
number of permutations for the computation of empirical p values (evalScoring)

kernel  
kernel function used for smoothing one of rbf, kNN, basePairDistance or your own

kernelparams  
list of parameters for the kernels

step.width  
size of a interpolation step in basepairs

Details

The filename for the python flat files are discrete_chrom_<chrom>_class_<label>.py where <chrom> and <label> are the names of the chromosome and class label.

Value

discretize.tscores  
a vector of discretized tscores

discretizeAllClasses.tscores  
creates python flatfiles (see details)

Author(s)

The MACAT development team

See Also

evalScoring, kernels, pythondata
Examples

```r
#loaddatapkg("stjudem")
data(stjude)
data(stjd)
# simple scoring with short running time
scores = evalScoring(stjd, "T", 1, nperms=100, cross.validate=FALSE)
discrete = discretize.tscores(scores)
```

---

`discretizeAll`  
*Discretize complete expression matrix*

Description

perform discretization on all chromosomes and write python flat files

Usage

```r
discretizeAll(data, margin = 10)
```

Arguments

- `data`: MACATData Object
- `margin`: symmetric quantile in percent

Details

The filename for the python flat files are `discrete_seqs_margin_<margin>_<chrom>_<chrom>.py` where `<chrom>` and `<margin>` are the names of the chromosome and the margin used for discretization. For details on the discretization see `discretize`.

Value

produces python flat file

Author(s)

The MACAT Development team

See Also

`discretize`
Examples

```r
# !!! takes some time !!!
## Not run:
#loaddatapkg("stjudem")
data(stjude)
data(stjd)
discretizeAll(stjd, margin=10)
## End(Not run)
```

evalScoring

**Score differential expression, assess significance, and smooth scores along the chromosome**

Description

This function computes for all genes on one chromosome the regularized t-statistic to score differential gene expression for two given groups of samples. Additionally these scores are computed for a number of permutations to assess significance. Afterwards these scores are smoothed with a given kernel along the chromosome to give scores for chromosomal regions.

Usage

```r
evalScoring(data, class, chromosome, nperms=1000, permute="labels", pcompute="empirical", subset=NULL, newlabels=NULL, kernel=rbf, kernelparams=NULL, cross.validate=TRUE, paramMultipliers=2^(-4:4), ncross=10, step.width=100000, memory.limit=TRUE, verbose=TRUE)
```

Arguments

data Gene expression data in the MACAT list format. See data(stjude) for an example.

class Which of the given class labels is to be analyzed

chromosome Chromosome to be analyzed

nperms Number of permutations

permute Method to do permutations. Default 'labels' does permutations of the class labels, which is the common and faster way to assess significance of differential expression. The alternative 'locations' does permutations of gene locations, is much slower and right now should be considered preliminary at best.

pcompute Method to determine the p-value for differential expression of each gene. Is only evaluated if the argument permute='labels' and in that case passed on to the function scoring

subset If a subset of samples is to be used, give vector of column-indices of these samples in the original matrix here.
newlabels: If other labels than the ones in the MACAT-list-structure are to be used, give them as character vector/factor here. Make sure argument 'class' is one of them.

kernel: Choose kernel to smooth scores along the chromosome. Available are 'kNN' for k-Nearest-Neighbors, 'rbf' for radial-basis-function (Gaussian), 'basePairDistance' for a kernel, which averages over all genes within a given range of base pairs around a position.

kernelparams: Additional parameters for the kernel as list, e.g., kernelparams=list(k=5) for taking the 5 nearest neighbours in the kNN-kernel. If NULL some defaults are set within the function.

cross.validate: Logical. Should the parameter settings for the kernel function be optimized by a cross-validation?

paramMultipliers: Numeric vector. If you do cross-validation of the kernel parameters, specify the multipliers of the given (standard) parameters to search over for the optimal one.

ncross: Integer. If you do cross-validation, specify how many folds.

step.width: Defines the resolution of smoothed scores on the chromosome, is in fact the distance in base pairs between 2 positions, for which smoothed scores are to be calculated.

memory.limit: If you have a computer with lots of RAM, setting this to FALSE will increase speed of computations.

verbose: logical; should function's progress be reported to STDOUT?; default: TRUE.

Details

Please see the package vignette for more details on this function.

Value

List of class 'MACATevalScoring' with 11 components:

original.geneid: Gene IDs of the genes on the chosen chromosome, sorted according to their position on the chromosome

original.loc: Location of genes on chromosome in base pairs from 5’end

original.score: Regularized t-score of genes on chromosome

original.pvalue: Empirical p-value of genes on chromosome. How often was a higher score observed than this one with random permutations? In other words, how significant seems this score to be?

steps: Positions on the chromosome in bp from 5’, for which smoothed scores have been computed.

sliding.value: Smoothed regularized t-scores at step-positions.

lower.permuted.border: Smoothed scores from permutations, lower significance border, currently 2.5%-quantile of permutation scores.
evaluateParameters

Evaluate Performance of Kernel Parameters by Cross-validation

Description

For a given data set, chromosome, class, and kernel function, this function helps in determining optimal settings for the kernel parameter(s). The performance of individual parameter setting is assessed by cross-validation.

upper.permuted.border

Smoothed scores from permutations, upper significance border, currently 97.5%-quantile of permutation scores.

chromosome

Chromosome, which has been analyzed

class

Class, which has been analyzed

chip

Identifier for used microarray

Author(s)

MACAT development team

See Also

scoring.plot, MACATevalScoring, getResults

Examples

data(stjd) # load example data

# if you have the data package 'stjudem' installed,
# you should work on the full data therein, of which
# the provided example data, is just a piece
#
#loaddatapkg("stjudem")
#data(stjude)

# T-lymphocyte versus B-lymphocyte on chromosome 1,
# smoothed with k-Nearest-Neighbours kernel(k=15),
# few permutations for higher speed
chrom1Tknn <- evalScoring(stjd,"T",chromosome="1",permute="labels",
nperms=100,kernel=kNN,kernelparams=list(k=15),step.width=100000)

# plotting on x11:
if (interactive())
  plot(chrom1Tknn)

# plotting on HTML:
if (interactive())
  plot(chrom1Tknn,"html")
evaluateParameters

Usage

evaluateParameters(data, class, chromosome, kernel, kernelparams = NULL,
paramMultipliers = 2^(-4:4), subset = NULL,
newlabels = NULL, ncross = 10, verbose = TRUE)

Arguments

data          Gene expression data in the MACAT list format. See data(stjude) for an example.
class         Sample class to be analyzed
chromosome    Chromosome to be analyzed
kernel        Choose kernel to smooth scores along the chromosome. Available are 'kNN' for k-Nearest-Neighbors, 'rbf' for radial-basis-function (Gaussian), 'basePairDistance' for a kernel, which averages over all genes within a given range of base pairs around a position.
kernelparams  Additional parameters for the kernel as list, e.g., kernelparams=list(k=5) for taking the 5 nearest neighbours in the kNN-kernel. If NULL some defaults are set within the function.
paramMultipliers Numeric vector. If you do cross-validation of the kernel parameters, specify these as multipliers of the given (standard) kernel parameter, depending on your kernel choice (see page 5 of the vignette). The multiplication results are the kernel argument settings, among which you want to search for the optimal one using cross-validation.
subset        If a subset of samples is to be used, give vector of column-indices of these samples in the original matrix here.
newlabels     If other labels than the ones in the MACAT-list-structure are to be used, give them as character vector/factor here. Make sure argument 'class' is one of them.
ncross         Integer. Specify how many folds in cross-validation.
verbose       Logical. Should progress be reported to STDOUT?

Value

A list of class 'MACATevP' with 4 components:

[parameterName] List of assessed settings for the parameter [parameterName].
avgResid       Average Residual Sum of Squares for the parameter settings in the same order as the first component.
multiplier     Multiplier of the original parameters in the same order as the first components.
best           List of parameter settings considered optimal by cross-validation. Can be directly inserted under the argument 'kernelparams' of the 'evalScoring' function.

Author(s)

MACAT development team
**getResults**

See Also

- evalScoring

Examples

```r
data(stjd)
evalkNN6 <- evaluateParameters(stjd, class="T", chromosome=6,kernel=kNN,
paramMultipliers=c(0.01,seq(0.2,2.0,0.2),2.5))
if (interactive()&&capabilities("X11"))
plot(evalkNN6)
```

getResults  

Access results of 'evalScoring'

Description

This function processes the result of the `evalScoring` function and returns a list of probe sets within chromosome regions deemed significant by MACAT. Additional annotation for these probe sets is provided along with their identifiers.

Usage

```r
getResults(MACATevalScoringOBJ)
```

Arguments

- `MACATevalScoringOBJ`  
  Object of class `MACATevalScoring`, usually the result from `evalScoring`

Details

The p-values have been computed individually for probe sets (genes), not for whole chromosome regions. Thus, regions deemed significant by sliding window approach do not have to consist only of probe sets with low p-values. These probe-set p-values are not used to determine whether a region is considered significant or not. Instead the comparison between actual and interpolated scores to actual and interpolated boundaries determines whether a region is considered significant.

This function is called within the plot function for the results of `evalScoring`, when HTML output is desired.

Value

A list with the following components, describing probe sets within chromosome regions deemed significant:

- `probeID`  
  IDs of probe sets within these chromosome regions
- `cytoband`  
  chromosomal bands these probe sets have been annotated to
- `geneSYM`  
  gene symbols these probe sets have been annotated to
pvalue  p-values for probe sets; see details
locusid  EntrezGene-(formerly LocusLink) IDs of these probe sets
genedescription  Description of genes the probe sets have been annotated to
probeScore  the differential expression scores for the probe sets
chromosome  chromosome, the analysis has been done for
class  sample class, the analysis has been done for

Author(s)
MACAT development team

See Also
evalScoring

Examples
data(stjd)
myevalres <- evalScoring(stjd, class="T", chromosome=6, nperms=10,
cross.validate=FALSE)
results <- getResults(myevalres)
summary(results)
results$probeID[1:20]

---

HTML functions for MACAT.

Description
HTML functions for internal usage by other MACAT functions. Normally not called by user.

Details
Internal HTML functions. Not called by user.

Author(s)
MACAT development team

See Also
plot.MACATevalScoring
**kernelize**

*Smooth expression values or scores*

**Description**

'kernelize' uses a kernel to smooth the data given in geneLocations by computing a weighted sum of the values vector. The weights for each position are given in the kernelweights matrix. A kernelweights matrix can be obtained by using the kernelmatrix function.

**Usage**

getsteps(geneLocations, step.width)
kernelmatrix(steps, geneLocations, kernel, kernelparams)
kernelize(values, kernelweights)

**Arguments**

- **geneLocations**: a list of gene locations (length n)
- **step.width**: the width of steps in basepairs
- **steps**: a list of locations where the kernelization shall be computed
- **kernel**: kernel function one of rbf, kNN or basePairDistance (or your own)
- **kernelparams**: a list of named parameters for the kernel (default is fitted to the data)
- **values**: vector of length n or matrix (m x n) of values that are to be smoothed
- **kernelweights**: a matrix of (n x steps) where n is the length of the values vector and steps is the number of points where you wish to interpolate

**Value**

- getsteps: a list of locations starting at min(geneLocations) going to max(geneLocations) with steps of size step.width
- kernelmatrix: a matrix of (n x steps) containing the kernel weights for each location in steps
- kernelize: a vector of length steps or a matrix (m x steps) containing the smoothed values

**Author(s)**

MACAT Development team

**See Also**

compute.sliding, evalScoring
Examples

```r
data(stjd)
genes = seq(100)
geneLocations = abs(stjd$geneLocation[genes])
geneExpression = stjd$expr[genes,]
step.width = 100000
steps = getsteps(geneLocations, step.width)
weights = kernelmatrix(steps, geneLocations, rbf, list(gamma=1/10^13))
kernelize = kernelize(geneExpression, weights)
plot(steps, kernelize[1,])
```

---

**kernelizeAll**  
*Smooth expression data for all chromosomes*

Description

'kernelizeAll' smoothes complete expression matrix and writes the result into one text file for each chromosome. These text files can be read into Python.

Usage

```r
kernelizeAll(data, step.width = 1e+05, kernel = rbf,
              kernelparams = list(gamma = 1/10^13))
```

Arguments

- **data**: MACATData Object
- **step.width**: size of steps for kernelization
- **kernel**: kernel function one of rbf, kNN, basePairDistance or your own
- **kernelparams**: list of named kernel parameters

Details

filename of the python flatfiles: `kernelize_seq_chrom_<chrom>.py` where `<chrom>` is the name of the chromosome.

Value

does not return anything; called for its side-effect that is to produce Python-readable text files

Author(s)

The MACAT Development Team

See Also

[pydata](#), [kernelizeToPython](#)
Examples

## Not run:
# !!! takes quite some time !!!
load datapkg("stjude")
data(stjude)
kernelizeAll(stjude)

## End(Not run)

kerneizeToPython

Smooth expression values and write to file

Description

Smoothes expression data for one chromosome and writes the result into a text file, which can be read into PYTHON, or returns it without writing. `kerneizeToPython` is the one-chromosome version of `kerneizeAll`.

Usage

```r
erkerneizeToPython(data, chrom, step.width = 1e+05, kernel = rbf,
  kernelparams = list(gamma = 1/10^13), saveToFile = TRUE)
```

Arguments

- **data**: MACATData Object
- **chrom**: kernelize all genes that are on this chromosome
- **step.width**: width of interploation steps
- **kernel**: kernel function one of rbf, kNN, basePairDistance or your own
- **kernelparams**: list of named kernel parameters
- **saveToFile**: logical indicating whether to save as flat file or not

Details

filename of the flatfile: `kerneized_seq_chrom_<chrom>.py` where `<chrom>` is the name of the chromosome.

Value

returns kernelized expression matrix

Author(s)

The MACAT Development team
Kernels

See Also

pydata.kernelizeAll

Examples

```r
## Not run:
data(stjd)
kernelized = kernelizeToPython(stjd, 3)
## End(Not run)
```

---

### Kernels

**Various kernel functions for computations in MACAT**

Description

Various Kernel functions for computations in MACAT. Normally not called by user. All kernel functions have the same arguments in the same order!!!

Usage

```r
kNN(geneLocations, position, params)
rbf(geneLocations, position, params = list(gamma=1/10^13))
basePairDistance(geneLocations, position, params = list(distance = 1e+06))
```

Arguments

- **geneLocations**: Location of genes
- **position**: Position on chromosome
- **params**: special kernel parameters
  - **kNN**: k = number of nearest genes
  - **rbf**: gamma = kernel width
  - **basePairDistance**: distance = distance within which all genes are averaged

Details

For internal use by other MACAT-functions. Not called by user.

Value

returns kernel weight for position, computed from the geneLocations

Author(s)

MACAT development team
loaddatapkg

See Also
evalScoring, compute.sliding, kernelize

Examples

data(stjd)
genes = seq(100)
geneLocations = abs(stjd$geneLocation[genes])
position = c(1000) # location for which you want the kernelweights
kernelweights = rbf(as.matrix(geneLocations), as.matrix(position),
list(gamma=1/10e13))
hist(kernelweights)

loaddatapkg  Load data package

Description

This function loads the data package, you need for seeing the demo and the examples. If you have
already installed the data package, it will simply attach it via library. Otherwise it will try to
download and install the package using functions from the package

Usage

loaddatapkg(mydatapkg,installDir=.libPaths()[1])

Arguments

mydatapkg  Name of the data package to load as String
installDir  Directory, into which the new package will be installed, if is not already in-
            stalled. Defaults to the first entry of the standard installation paths.

Note

The package stjudem by now is a Bioconductor example data package, too. Thus, you can also
install with the following command: BiocManager::install('stjudem')

Author(s)

Joern Toedling

See Also

install.packages

Examples

## Not run: loaddatapkg("stjudem") # to load the data package "stjudem"
### Description

Function plots scores, 0.025 and 0.975 quantiles of the permuted scores (grey lines), and sliding average score (red line) along the chromosome. Yellow dots highlight regions, in which the smoothed absolute scores exceed the permutation-derived quantile boundaries.

### Usage

```r
## S3 method for class 'MACATevalScoring'
plot(x, output = "x11",
     HTMLfilename = NULL, mytitle = NULL, new.device = TRUE, ...)
```

### Arguments

- **x**  
  MACATevalScoring object.

- **output**  
  plot "x11" or create a "html" -file with further information. HTML-page will open automatically.

- **HTMLfilename**  
  HTML-filename, default: Results<CHROMOSOME>_<CLASS>.html

- **mytitle**  
  Title of HTML-page, default: "Results of class <CLASS> on chromosome <CHROMOSOME>"

- **new.device**  
  if FALSE: Possibility to plot several plots in one device

- **...**  
  further arguments passed on to generic function `plot`

### Details

One can create a HTML-page on-the-fly if argument output='html'. The HTML-page provides informations about highlighted regions in the plot. Furthermore there are click-able Entrezgene-IDs for further analysis.

### Author(s)

MACAT development team

### See Also

evalScoring, getResults

### Examples

```r
# see function 'evalScoring' for an example
```
**Description**

This function reads expression data either from a saved R-file (.RData,.rda), or from a tab-separated text-file (.xls). For building a MACAT-list structure from objects in your workspace, you can either use this function or the convenience wrapper ‘buildMACAT’.

**Usage**

```r
preprocessedLoader(rdatafile, chip, labels = NULL, chromLocObj = NULL,
   rdafile = TRUE, tabfile = FALSE, labelfile = FALSE)
```

**Arguments**

- `rdatafile`: Complete name of the expression data file, or the expression matrix
- `chip`: Identifier of the used microarray. To date only commercial Affymetrix microarrays are supported by MACAT
- `labels`: Classlabels of the samples, vector of same length as number of columns in expression matrix; alternatively complete name of textfile with one label per line
- `chromLocObj`: Object of class `chromLocation` specifying the genomic position, each probe on the array is mapped to. If not provided, it is build in the function using annotate’s function `buildChromLocation`.
- `rdafile`: Logical; is first argument a saved R-file?
- `tabfile`: Logical; is first argument a tab-separated text file?
- `labelfile`: Logical; is third argument a file with one label per line?

**Value**

List of class ’MACATData’ with 6 components:

- `geneName`: Identifiers of genes/probe sets in expression data
- `geneLocation`: Location of genes on their chromosome as distance from 5’end in base pairs Negative numbers denote genes on the antisense strand.
- `chromosome`: Chromosome of the respective gene. Components ’geneName’, ’geneLocation’, and ’chromosome’ are in the same order.
- `expr`: expression matrix with rows = genes and columns = samples/patients
- `labels`: (disease) subtype of each sample, has length = number of columns of expression matrix
- `chip`: Identifier for Microarray used for the experiments
Python data format

Note

At present, macat can only work with Affymetrix microarrays, for which an annotation package is installed on your system. Such annotation packages can either be obtained from the Bioconductor annotation packages repository or be constructed using the Bioconductor package AnnBuilder. For an example, see the common annotation package hgu95av2.

Author(s)

MACAT development team

See Also

buildMACAT, read.table, stjd, stjude in package 'stjudem'

Examples

## Not run:
# assume you have your HG-U95Av2 expression values in a
tab-separated text file, called 'foo.txt'
mydata <- preprocessedLoader("foo.txt","hgu95av2",rdafile=FALSE,tabfile=TRUE)

## End(Not run)

Python data format

Flat file format

Description

This is a description of the text files, usually written by MACAT functions such as kernelizeToPython. These text files can be read into PYTHON. Each flat file contains a list of matrices. The outer list has one entry for each class in the data set, the inner matrices consist of all the sequences from the specified chromosome belonging to that class. Each row represents one sample, each column a gene from the respective chromosome.

The lists are delimited by '[' and ']' brackets. The matrices are written as lists of lists (e.g. [[],[],[...]]).

Examples

# The files look like this:
# [ [[1.11, 1.32, 0.92, ...],[...],[...]],
#  [[0.45, 0.91, 1.84, ...],[...],[...]],
#  [[1.06, 1.59, 0.73, ...],[...],[...]],
#  ]
scoring

Compute (regularized) t-scores for gene expression data

Description

This function computes for all genes in an expression matrix the (regularized) t-scores (statistics) with the given class labels and a number of permutations of these labels. Each gene is also assigned a p-value either empirically from the permutation scores or from a t-distribution.

Usage

scoring(data, labels, method = "SAM", pcompute = "tdist", nperms = 1000, memory.limit = TRUE, verbose = TRUE)

Arguments

data          Expression matrix with rows = genes and columns = samples
labels        Vector or factor of class labels; Scoring works only with two classes!
method        Either "SAM" to compute regularized t-scores, or "t.test" to compute Student’s t-statistic
pcompute      Method to compute p-values for each genes, either "empirical" to do permutations and compute p-values from them, or "tdist" to compute p-values based on respective t-distribution
nperms        Number of permutations of the labels to be investigated, if argument 'pcompute="empirical"
memory.limit  Logical, if you have a really good computer (>2GB RAM), setting this FALSE will increase speed of computations
verbose       Logical, if progress should be reported to STDOUT

Details

If 'pcompute="empirical"', the statistic is computed based on the given class labels, afterwards for 'nperms' permutations of the labels. The p-value for each gene is then the proportion of permutation statistics that are higher or equal than the statistic from the real labels. For each gene the 2.5%- and the 97.5%-quantile of the permutation statistics are also returned as lower and upper 'significance threshold'.

If 'pcompute="tdist", the statistic is computed only based on the given class labels, and the p-value is computed from the t-distribution with (Number of samples - 2) degrees of freedom.

Value

A list, with four components:

observed      (Regularized) t-scores for all genes based on the given labels
pvalues       P-values for all genes, either from permutations or t-distribution
expected.lower 2.5%-quantile of permutation test-statistics, supposed to be a lower 'significance border' for the gene; or NULL if p-values were computed from t-distribution

expected.upper 97.5%-quantile of permutation test-statistics, supposed to be an upper 'significance border' for the gene; or NULL if p-values were computed from t-distribution

Note

In package macat, this function is only called internally by the function evalScoring

Author(s)

MACAT development team

References

Regarding the regularized t-score please see the macat vignette.

See Also

evalScoring

Examples

data(stjd)
# compute gene-wise regularized t-statistics for
# T- vs. B-lymphocyte ALL:
isT <- as.numeric(stjd$labels=="T")
TvsB <- scoring(stjd$expr,isT,method="SAM",pcompute="none")
summary(TvsB$observed)

stjd

Subset Microarray Data from St.Jude Children Research Hospital
(USA)

Description

Example for list-structure used by many functions in MACAT. It’s based on the gene expression data published by Yeoh et al. (2002) The data has been preprocessed using ‘vsn’ on probe level and the probe values have been summed up to probe set values using the ‘median polish’ procedure. This is a subset of the data, containing only the data for the 5000 probe sets with the highest variance across the samples and for 10 exemplary samples, 5 from T-lymphocytic Acute Lymphocytic Leukemia (ALL) and 5 from B-lymphocytic ALL.

Usage

data(stjd)
Format

List of class 'MACATData' with 6 components:

- **geneName**: Identifiers of genes/probe sets in expression data
- **geneLocation**: Location of genes on their chromosome as distance from 5’end in base pairs Negative numbers denote genes on the antisense strand.
- **chromosome**: Chromosome of the respective gene. Components ‘geneName’, ‘geneLocation’, and ‘chromosome’ are in the same order.
- **expr**: expression matrix with rows = genes and columns = samples/patients
- **labels**: (disease) subtype of each sample, has length = number of columns of expression matrix
- **chip**: Identifier for Microarray used for the experiments (here for the Affymetrix HG-U95av2 Oligonucleotide GeneChip)

Note

For the full data package see the Bioconductor data package stjudem. If it is not already installed on your system, try if (!requireNamespace("BiocManager", quietly=TRUE)); install.packages("BiocManager"); BiocManager::install("stjudem")

References


See Also

- buildMACAT, stjude in package 'stjudem' for the complete expression data

Examples

```r
data(stjd)
summary(stjd)
```
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