Package ‘gwascat’

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Title representing and modeling data in the EMBL-EBI GWAS catalog

Version 2.34.0

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Description Represent and model data in the EMBL-EBI GWAS catalog.

Enhances SNPlocs.Hsapiens.dbSNP144.GRCh37

Depends R (>= 3.5.0), methods

Imports S4Vectors (>= 0.9.25), IRanges, GenomeInfoDb, GenomicRanges (>= 1.29.6), GenomicFeatures, readr, Biostrings, AnnotationDbi, BiocFileCache, snpStats, VariantAnnotation, AnnotationHub

Suggests DO.db, DT, knitr, RBGL, testthat, rmarkdown, dplyr, Gviz, Rsamtools, rtracklayer, graph, ggbio, DelayedArray, TxDb.Hsapiens.UCSC.hg19.knownGene, org.Hs.eg.db, BiocStyle

VignetteBuilder knitr

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| as_GRanges | produce a GRanges from gwascat tibble |

### Description

produce a GRanges from gwascat tibble
**bindcadd_snv**

**Usage**

```r
as_GRanges(
  x,
  short = TRUE,
  for_short = c("PUBMEDID", "DATE", "DISEASE/TRAIT", "SNPS"),
  genome_tag = "GRCh38"
)
```

**Arguments**

- **x**: a tibble from `get_cached_gwascat()`
- **short**: logical(1) if TRUE only keep selected columns in mcols
- **for_short**: character() column names to keep in mcols
- **genome_tag**: character(1) defaults to "GRCh38"

**Description**

bind CADD scores of Kircher et al. 2014 to a GRanges instance; by default will use HTTP access at UW

**Usage**

```r
bindcadd_snv(
  gr,
  fn = "http://krishna.gs.washington.edu/download/CADD/v1.0/1000G.tsv.gz"
)
```

**Arguments**

- **gr**: query ranges to which CADD scores should be bound
- **fn**: path to Tabix-indexed bgzipped TSV of CADD as distributed at krishna.gs.washington.edu on 1 April 2014

**Details**

joins CADD fields at addresses that match query; the CADD fields for query ranges that are not matched are set to NA

**Value**

GRanges instance with additional fields as obtained in the CADD resource
chklocs

return TRUE if all named SNPs with locations in both the SNPlocs package and the gwascat agree

Description

return TRUE if all named SNPs with locations in both the SNPlocs package and the gwascat agree

Usage

chklocs(chrtag = "20", gwwl = gwrngs19)

Arguments

chrtag character, chromosome identifier
gwwl instance of {gwaswloc}
ebicat_2020_04_30

**ebicat_2020_04_30**  
serialized gwaswloc instance from april 30 2020, sample of 50000 records

**Description**
serialized gwaswloc instance from april 30 2020, sample of 50000 records

**Usage**
ebicat_2020_04_30

**Format**
gwaswloc instance

---

**g17SM**  
SnpMatrix instance from chr17

**Description**
SnpMatrix instance from chr17

**Usage**
g17SM

**Format**
snpStats SnpMatrix instance

---

**getRsids**  
generic snp name retrieval

**Description**
generic snp name retrieval

**Usage**
getRsids(x)

**Arguments**

\[ x \]  
gwaswloc
getRsids, gwaswloc-method

**Description**

Specific SNP name retrieval

**Usage**

```r
## S4 method for signature 'gwaswloc'
getRsids(x)
```

**Arguments**

- `x`: gwaswloc

---

getTraits

**Description**

Generic trait retrieval

**Usage**

```r
getTraits(x)
```

**Arguments**

- `x`: gwaswloc
getTraits,gwaswloc-method

**Description**

specific trait retrieval

**Usage**

```r
## S4 method for signature 'gwaswloc'
getTraits(x)
```

**Arguments**

- `x` : gwaswloc

---

get_cached_gwascat

**Description**

use BiocFileCache to retrieve and keep an image of the tsv file distributed by EBI

**Usage**

```r
get_cached_gwascat(
  url = "http://www.ebi.ac.uk/gwas/api/search/downloads/alternative",
  cache = BiocFileCache::BiocFileCache(),
  refresh = FALSE,
  ...
)
```

**Arguments**

- `url` : character(1) url to use
- `cache` : BiocFileCache::BiocFileCache instance
- `refresh` : logical(1) force download and recaching
- `...` : passed to bfcadd

**Value**

a tibble as produced by readr::read_tsv, with attributes extractDate (as recorded in cache as ‘access_time’, and problems (a tibble returned by read_tsv).
Note

will If query of cache with ‘ebi.ac.uk/gwas’ returns 0-row tibble, will populate cache with bfcadd. Uses readr::read_tsv on cache content to return tibble. The etag field does not seem to be used at EBI, thus user must check for updates.

---

**gg17N**

*genotype matrix from chr17 1000 genomes*

---

**Description**

*genotype matrix from chr17 1000 genomes*

**Usage**

gg17N

**Format**

matrix

**Examples**

data(gg17N)

gg17N[1:4,1:4]

---

**gr6.0_hg38**

*image of locon6 in GRanges, lifted over to hg38*

---

**Description**

*image of locon6 in GRanges, lifted over to hg38*

**Usage**

gr6.0_hg38

**Format**

GRanges instance
Description

character vector of rs numbers for SNP on chr17

Usage

gw6.rs_17

Format

character vector

gwascat_from_AHub

Description

grab an image of EBI GWAS catalog from AnnotationHub

Usage

gwascat_from_AHub(tag = "AH91571", simple = FALSE, fixNonASCII = TRUE)

Arguments

tag character(1) defaults to "AH91571" which is the 3.30.2021 image
simple logical(1) if TRUE, just returns data.frame as retrieved from EBI; defaults to FALSE
fixNonASCII logical(1) if TRUE, use iconv to identify and eliminate non-ASCII content

Value

If ‘simple’, a data.frame is returned based on TSV data produced by EBI. Otherwise, non-ASCII content is processed according to the value of ‘fixNonASCII’ and a ‘gwaswloc’ instance is returned, which has a concise show method. This can be coerced to a simple GRanges instance with as(..., "GRanges"). The reference build is GRCh38.

Examples

gwcat = gwascat_from_AHub()
gwcat
**gwastagger**

*GRanges with LD information on 9998 SNP*

**Description**

GRanges with LD information on 9998 SNP

**Usage**

`gwastagger`

**Format**

GRanges

**gwaswloc-class**

*container for gwas hit data and GRanges for addresses*

**Description**

container for gwas hit data and GRanges for addresses

**gwcatrix\_snapshot**

*use AnnotationHub snapshot as basis for gwaswloc structure creation*

**Description**

use AnnotationHub snapshot as basis for gwaswloc structure creation

**Usage**

`gwcatrix\_snapshot(x, fixNonASCII = TRUE)`

**Arguments**

- `x` inherits from data.frame, with columns consistent with EBI table
- `fixNonASCII` logical(1) if TRUE, use iconv to replace non-ASCII character, important for CMD check but perhaps not important for applied use
**Examples**

```r
ah = AnnotationHub::AnnotationHub()
entitytab = AnnotationHub::query(ah, "gwascatData")
cand = names(entitytab)[1]
stopifnot(nchar(cand)>0)
tab = ah[[cand]]
gww = gwcat_snapshot(tab)
gww
length(gww)
```

---

**gwcex2gviz: Prepare salient components of GWAS catalog for rendering with Gviz**

**Description**

Prepare salient components of GWAS catalog for rendering with Gviz

**Usage**

```r
gwcex2gviz(
  basegr,
  contextGR = GRanges(seqnames = "chr17", IRanges::IRanges(start = 37500000, width = 1e+06)),
  txrefobj = TxDb.Hsapiens.UCSC.hg19.knownGene::TxDb.Hsapiens.UCSC.hg19.knownGene,
  genome = "hg19",
  genesymobj = org.Hs.eg.db::org.Hs.eg.db,
  plot.it = TRUE,
  maxmlp = 25
)
```

**Arguments**

- `basegr`: `gwaswloc` instance containing information about GWAS in catalog
- `contextGR`: A `GRanges` instance delineating the visualization in genomic coordinates
- `txrefobj`: a `TxDb` instance
- `genome`: character tag like 'hg19'
- `genesymobj`: an `OrgDb` instance
- `plot.it`: logical, if `FALSE`, just return list
- `maxmlp`: maximum value of -10 log p – winsorization of all larger values is performed, modifying the contents of `Pvalue_mlogp` in the `elementMetadata` for the call

**Examples**

```r
data(ebicat_2020_04_30)
# GenomeInfoDb::seqlevelsStyle(ebicat_2020_04_30) = "UCSC" # no more
GenomeInfoDb::seqlevels(ebicat_2020_04_30) = paste0("chr", GenomeInfoDb::seqlevels(ebicat_2020_04_30))
gwcex2gviz(ebicat_2020_04_30)
```
ldtagr

expand a list of variants by including those in a VCF with LD exceeding some threshold; uses snpStats ld()

Usage

```r
ldtagr(
    snprng,
    tf,
    samples,
    genome = "hg19",
    lbmaf = 0.05,
    lbR2 = 0.8,
    radius = 1e+05
)
```

Arguments

- **snprng**: a named GRanges for a single SNP. The name must correspond to the name that will be assigned by genotypeToSnpMatrix (from VariantTools) to the corresponding column of a SnpMatrix.
- **tf**: TabixFile instance pointing to a bgzipped tabix-indexed VCF file
- **samples**: a vector of sample identifiers, if excluded, all samples used
- **genome**: tag like 'hg19'
- **lbmaf**: lower bound on variant MAF to allow consideration
- **lbR2**: lower bound on R squared for regarding SNP to be incorporated
- **radius**: radius of search in bp around the input range

Value

a GRanges with names corresponding to 'new' variants and mcols fields 'paramRangeID' (base variant input) and 'R2'

Note

slow but safe approach. probably a matrix method could be substituted using the nice sparse approach already in snpStats

Author(s)

VJ Carey
Examples

cand = GenomicRanges::GRanges("1", IRanges::IRanges(113038694, width=1))
names(cand) = "rs883593"
requireNamespace("VariantAnnotation")
expath = dir(system.file("vcf", package="gwascat"), patt=".*exon.*gz", full=TRUE)
tf = Rsamtools::TabixFile(expath)
ldtagr( cand, tf, lbR2 = .8)

Description

location data for 10000 SNP

Usage

locon6

Format

data.frame, coordinates are hg19

locx4trait

get locations for SNP affecting a selected trait

Description

get locations for SNP affecting a selected trait

Usage

locs4trait(gwwl, trait, tag = "DISEASE/TRAIT")

Arguments

gwwl instance of \{gwaswloc\}
trait character, name of trait
tag character, name of field to be used for trait enumeration
10w17  

SnpMatrix instance from chr17

Description

SnpMatrix instance from chr17

Usage

10w17

Format

snpStats SnpMatrix instance

makeCurrentGwascat  

read NHGRI GWAS catalog table and construct associated GRanges instance records for which clear genomic position cannot be determined are dropped from the ranges instance an effort is made to use reasonable data types for GRanges metadata, so some qualifying characters such as (EA) in Risk allele frequency field will simply be omitted during coercion of contents of that field to numeric.

Description

read NHGRI GWAS catalog table and construct associated GRanges instance records for which clear genomic position cannot be determined are dropped from the ranges instance an effort is made to use reasonable data types for GRanges metadata, so some qualifying characters such as (EA) in Risk allele frequency field will simply be omitted during coercion of contents of that field to numeric.

Usage

makeCurrentGwascat(
    table.url = "http://www.ebi.ac.uk/gwas/api/search/downloads/alternative",
    fixNonASCII = TRUE,
    genome = "GRCh38",
    withOnt = TRUE
)
obo2graphNEL

Arguments

- `table.url` string identifying the .txt file curated at EBI/EMBL
- `fixNonASCII` logical, if TRUE, non-ASCII characters as identified by iconv will be replaced by asterisk
- `genome` character string: 'GRCh38' is default and yields current image as provided by EMBL/EBI; 'GRCh37' yields a realtime liftOver to hg19 coordinates, via AnnotationHub storage of the chain files. Any other value yields an error.
- `withOnt` logical indicating whether 'alternative' (ontology-present, includes repetition of loci with one:many ontological mapping) or 'full' (ontology-absent, one record per locus report) version of distributed table

Value

a slightly extended GRanges instance, with class name 'gwaswloc'; the purpose of the introduction of this class is to support a concise show method that does not produce very long lines owing to large numbers of fields in the mcols component.

Note

'readr::read_tsv' records problems when some records have field contents that are inconsistent with the column specification. This information can be retrieved from the metadata slot of the returned object, as noted in a message produced when this function is run.

Author(s)

VJ Carey

Examples

```r
# if you have good internet access
if (interactive()) {
  newcatr = makeCurrentGwascat()
  newcatr
}
```

obo2graphNEL

convert a typical OBO text file to a graphNEL instance (using Term elements)

Description

convert a typical OBO text file to a graphNEL instance (using Term elements)
Usage

obo2graphNEL(
  obo = "human-phenotype-ontology.obo",
  kill = "\[Typedef\]",
  killTrailSp = TRUE
)

Arguments

obo  string naming a file in OBO format
kill entity types to be excluded from processing – probably this should be in a ’keep’ form, but for now this works.
killTrailSp In the textual version of EFO ca. Aug 2015, there is a trailing blank in the tag field defining EFO:0000001, which is not present in references to this term. Set this to TRUE to eliminate this, or graphNEL construction will fail to validate.

Details

Very rudimentary list and grep operations are used to retain sufficient information to map the DAG to a graphNEL, using formal term identifiers as node names and ’is-a’ relationships as edges, and term names and other metadata are assigned to nodeData components.

Value

a graphNEL instance

Note

The OBO for Human Disease ontology is serialized as text with this package.

Author(s)

VJ Carey <stvjc@channing.harvard.edu>

References

For use with human disease ontology, http://www.obofoundry.org/cgi-bin/detail.cgi?id=disease_ontology

Examples

data(efo.obo.g)
requireNamespace("graph")
hn = graph::nodes(efo.obo.g)[1:5]
hn
graph::nodeData(efo.obo.g, hn[5])
process_gwas_dataframe

convert GWAS catalog data.frame to gwaswloc, a GRanges extension with simple show method

Description

convert GWAS catalog data.frame to gwaswloc, a GRanges extension with simple show method

Usage

process_gwas_dataframe(df)

Arguments

df data.frame

riskyAlleleCount given a matrix of subjects x SNP calls, count number of risky alleles

Description

given a matrix of subjects x SNP calls, count number of risky alleles for various conditions, relative to NHGRI GWAS catalog

Usage

riskyAlleleCount(
callmat,
matIsAB = TRUE,
chr,
gwwl,
snpap = "SNPlocs.Hsapiens.dbSNP144.GRCh37",
gencode = c("A/A", "A/B", "B/B")
)

Arguments

callmat matrix with subjects as rows, SNPs as columns; entries can be generic A/A, A/B, B/B, or specific nucleotide calls
matIsAB logical, FALSE if nucleotide codes are present, TRUE if generic call codes are present; in the latter case, gwascat:::ABmat2nuc will be run
chr code for chromosome, should work with the SNP annotation getSNPlocs function, so likely "ch[nn]"
gwwl an instance of {gwaswloc}
snpap name of a Bioconductor SNPlocs.Hsapiens.dbSNP.* package
genencode codes used for generic SNP call
Value
matrix with rows corresponding to subjects, columns corresponding to SNP

Examples
## Not run:
data(gg17N) # translated from GGdata chr 17 calls using ABmat2nuc
data(ebicat37)
library(GenomeInfoDb)
seqlevelsStyle(ebicat37) = "UCSC"
h17 = riskyAlleleCount(gg17N, matIsAB=FALSE, chr="ch17", gwwl=ebicat37)
h17[1:5,1:5]
table(as.numeric(h17))
## End(Not run)

---

si.hs.37  Seqinfo for GRCh37

Description
Seqinfo for GRCh37

Usage
si.hs.37

Format
GenomeInfoDb Seqinfo instance

---

si.hs.38  Seqinfo for GRCh38

Description
Seqinfo for GRCh38

Usage
si.hs.38

Format
GenomeInfoDb Seqinfo instance
subsetByChromosome  generic trait subsetting

Description
generic trait subsetting

Usage
subsetByChromosome(x, ch)

Arguments
x  gwaswloc
ch  character vector of chromosomes

subsetByChromosome,gwaswloc-method  specific trait subsetting

Description
specific trait subsetting

Usage
## S4 method for signature 'gwaswloc'
subsetByChromosome(x, ch)

Arguments
x  gwaswloc
ch  character vector of chromosomes
subsetByTraits\_{\text{gwaswloc-method}}

\begin{itemize}
  \item \textit{subsetByTraits} \hspace{1cm} \textit{generic trait subsetting}
  \item \textit{subsetByTraits, gwaswloc-method} \hspace{1cm} \textit{specific trait subsetting}
\end{itemize}

\section*{Description}

generic trait subsetting

\section*{Usage}

\begin{verbatim}
subsetByTraits(x, tr)
\end{verbatim}

\section*{Arguments}

\begin{itemize}
  \item \textit{x} \hspace{1cm} \textit{gwaswloc}
  \item \textit{tr} \hspace{1cm} \textit{character vector of traits}
\end{itemize}

\section*{subsetByTraits, gwaswloc-method}

\begin{itemize}
  \item \textit{subsetByTraits, gwaswloc-method} \hspace{1cm} \textit{specific trait subsetting}
\end{itemize}

\section*{Description}

specific trait subsetting

\section*{Usage}

\begin{verbatim}
## S4 method for signature 'gwaswloc'
subsetByTraits(x, tr)
\end{verbatim}

\section*{Arguments}

\begin{itemize}
  \item \textit{x} \hspace{1cm} \textit{gwaswloc}
  \item \textit{tr} \hspace{1cm} \textit{character vector of traits}
\end{itemize}
### topTraits

**Description**

operations on GWAS catalog

**Usage**

```r
topTraits(gwwl, n = 10, tag = "DISEASE/TRAIT")
```

**Arguments**

- `gwwl` instance of `{gwaswloc}`
- `n` numeric, number of traits to report
- `tag` character, name of field to be used for trait enumeration

**Value**

topTraits returns a character vector of most frequently occurring traits in the database

locs4trait returns a `{gwaswloc}` object with records defining associations to the specified trait

chklocs returns a logical that is TRUE when the asserted locations of SNP in the GWAS catalog agree with the locations given in the dbSNP package SNPlocs.Hsapiens.dbSNP144.GRCh37

**Author(s)**

VJ Carey <stvjc@channing.harvard.edu>

**Examples**

```r
data(ebicat_2020_04_30)
topTraits(ebicat_2020_04_30)
```

### traitsManh

**Description**

use ggbio facilities to display GWAS results for selected traits in genomic coordinates
traitsManh

Usage

traitsManh(
  gwr,
  selr = GRanges(seqnames = "chr17", IRanges(3e+07, 5e+07)),
  traits = c("Asthma", "Parkinson's disease", "Height", "Crohn's disease"),
  truncmlp = 25,
  ...
)

Arguments

  gwr  GRanges instance as managed by the gwaswloc container design, with Disease.Trait and Pvalue\_mlog among elementMetadata columns
  selr A GRanges instance to restrict the gwr for visualization. Not tested for noncontiguous regions.
  traits Character vector of traits to be exhibited; GWAS results with traits not among these will be labeled "other".
  truncmlp Maximum value of -log10 p to be displayed; in the raw data this ranges to the hundreds and can cause bad compression.
  ...

Details

  uses a ggbio autoplot

Value

  autoplot value

Note

  An xlab is added, concatenating genome tag with seqnames tag.

Author(s)

  VJ Carey <stvjc@channing.harvard.edu>

Examples

  # do a p-value truncation if you want to reduce compression
  ## Not run:  # ggbio July 2018
  data(ebicat_2020_04_30)
  library(GenomeInfoDb)
  seqlevelsStyle(ebicat_2020_04_30) = "UCSC"
  traitsManh(ebicat_2020_04_30)

  ## End(Not run)
Description
extractor for gwaswloc

Usage
## S4 method for signature 'gwaswloc,ANY,ANY,ANY'

x[i, j, ..., drop = FALSE]

Arguments

- **x**: gwaswloc
- **i**: index
- **j**: index
- **...**: addtl indices
- **drop**: logical(1)
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