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**git_branch**  RELEASE_3_18  
**git_last_commit**  fb27d0b  
**git_last_commit_date**  2024-02-27  
**Repository**  Bioconductor 3.18  
**Date/Publication**  2024-04-10  
**Author**  Stephen Pederson [aut, cre] (<https://orcid.org/0000-0001-8197-3303>)  
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extraChIPs-package

Description

The package provides three categories of important functions: Range-based, Visualisation and Convenience functions, with most centred around GenomicRanges objects.

Range-based Functions

Many of the range-based functions included in this package have a focus on retaining the mcols information whilst manipulating the ranges, such as `reduceMC()` which not only reduces the Ranges, but collapses the mcols into vectors or IRanges::CompressedList objects. Key function from this group are:

- `reduceMC()`, `setdiffMC()`, `intersectMC()`, `unionMC()`, `distinctMC()` and `chopMC()`
- `bestOverlap()` and `propOverlap()` provide simple output easily able to be added as a column within the mcols element
- `as_tibble()` coerces a GRanges object to a tibble::tibble.
- `colToRanges()` enables parsing of a single column to a GRanges object, setting all other columns as the mcols element.
- `stitchRanges()` merges nearby ranges setting barrier ranges which cannot be crossed when merging
- `partitionRanges()` break apart one set of ranges by another
- `dualFilter()` filters ranges from sliding windows using a guide set of reference ranges where signal is confidently expected
- `mergeByCol()` merges overlapping ranges, as produced by sliding windows
- `mapByFeature()` is able to map a set of GRanges to the most appropriate gene, using any optional combination of promoters, enhancers and HiC interactions
- `grlToSE()` takes selected columns from a GRangesList and sets them as assays within a SummarizedExperiment::RangedSummarizedExperiment object. Used for combining peak intensities or results across multiple ChIP targets.
Visualisation Functions

- `plotHFGC()` is a wrapper to Gviz plotting functions, able to take any combination of HiC, Features, Genes and Coverage (i.e. BigWig) and plot a specified range.
- `plotOverlaps()` visualises overlapping ranges as an UpSet plot or Venn Diagram
- `plotProfileHeatmap()` plots the average signal around a set of ranges, as prepared by `getProfileData()`
- `plotPie()` and `plotSplitDonut()` enable simple comparison across multiple annotation columns within a GRanges object.
- `plotAssayDensities()`, `plotAssayPCA()` and `plotAssayRle()` provide simple interfaces to plotting key values from a `SummarizedExperiment::RangedSummarizedExperiment`.

Convenience Functions

- `fitAssayDiff()` enables differential signal analysis on a `SummarizedExperiment` object
- `collapseGenes()` prints a vector of genes for an rmarkdown document, using italics.
- `importPeaks()` imports large numbers of broadPeak or narrowPeak files
- `makeConsensus()` forms consensus peaks from overlapping ranges within a GRangesList()
- `voomWeightsFromCPM()` allows creation of a `limma::EList` object as would be created from counts by `limma::voom()`, but using `edgeR::cpm()` values as input.

Author(s)

Stevie Pederson

See Also

Useful links:

- [https://github.com/smped/extraChIPs](https://github.com/smped/extraChIPs)
- Report bugs at [https://github.com/smped/extraChIPs/issues](https://github.com/smped/extraChIPs/issues)

---

This is a modified version of harmonicmeanp::p.hmp developed by Prof Daniel Wilson, and hardwired to simply return a combined asymptotically exact HMP. Hardwiring like this gives a 10-fold speed-up. Further modifications may be possible, but this seems enough for now.

Usage

`.ec_HMP(p, w)`
Arguments

\begin{itemize}
  \item \texttt{p} \quad \text{vector of p-values}
  \item \texttt{w} \quad \text{vector of weights}
\end{itemize}

Similar to the above, this produces the FWER-controlled version in a streamlined way.

Description

Similar to the above, this produces the FWER-controlled version in a streamlined way.

Usage

\begin{verbatim}
.ec_HMP_adj(p, w, L)
\end{verbatim}

Arguments

\begin{itemize}
  \item \texttt{p} \quad \text{vector of p-values}
  \item \texttt{w} \quad \text{vector of weights}
  \item \texttt{L} \quad \text{Number of global tests}
\end{itemize}

Make a profile heatmap

Description

Make a profile heatmap with optional summary panel at the top.

Usage

\begin{verbatim}
.makeFinalProfileHeatmap(
  data,
  x = NULL,
  y = NULL,
  fill = NULL,
  colour = NULL,
  linetype = NULL,
  facet_x = NULL,
  facet_y = NULL,
  summary_fun = c("mean", "median", "min", "max", "none"),
  rel_height = 0.3,
  x_lab = NULL,
\end{verbatim}
y_lab = NULL,
fill_lab = NULL,
label_side = c("left", "right", "none"),
...
)

Arguments

data          A data.frame or tibble in long form
x, y          The values mapped to the x & y axes
fill          The column used for heatmap colours
colour, linetype
              Columns used for the summary plot in the top panel
facet_x, facet_y
              Columns used to facet the plot along these axes
summary_fun   Function used to create the summary value at each position
rel_height    The relative height of the top panel
x_lab, y_lab, fill_lab
              _labels added to x/y-axes & the fill legend
...
              Passed to facet_grid

Details

The workhorse function for generating the final heatmap Expects a single data.frame in long form
with requisite columns

Value

A ggplot2 object

.mapFeatures  Map ranges to genes using features as an anchor

Description

Map ranges to genes using features as an anchor

Usage

.mapFeatures(.gr, .feat, .genes, .cols, .gr2feat, .feat2gene, ...)
.mapGi

**Arguments**

- `.gr` The ranges to map onto
- `.feat` Features to use for mapping
- `.genes` GRanges object containing gene-level information
- `.cols` The columns from `.genes` to map onto `.gr`
- `.gr2feat` The maximum distance between ranges and features
- `.feat2gene` The maximum distance between features & genes
- `...` Passed to findOverlaps and subsetByOverlaps

**Value**

A data.frame

---

.mapGi

*Map ranges to genes via Interactions*

**Description**

Map ranges to genes via Interactions

**Usage**

.mapGi(.gr, .gi, .genes, .cols, .gr2gi, .gi2gene, ...)

**Arguments**

- `.gr` The ranges to map onto
- `.gi` GInteractions object
- `.genes` GRanges object containing gene-level information
- `.cols` The columns from `.genes` to map onto `.gr`
- `.gr2gi` The maximum distance between ranges and anchors
- `.gi2gene` The maximum distance between anchors & genes
- `...` Passed to findOverlaps

**Value**

data.frame of mapped ranges
addDiffStatus

Add a status column based on significance and estimated change

Usage

addDiffStatus(x, ...)

## S4 method for signature 'data.frame'
addDiffStatus(
x,
fc_col = "logFC",
sig_col = c("FDR", "hmp_fdr", "p_fdr", "adj.P.Value"),
alpha = 0.05,
cutoff = 0,
up = "Increased",
down = "Decreased",
other = "Unchanged"
addDiffStatus

missing = "Undetected",
new_col = "status",
drop = FALSE,
...
)

## S4 method for signature 'DataFrame'
addDiffStatus(x, new_col = "status", ...)

## S4 method for signature 'GRanges'
addDiffStatus(x, ...)

## S4 method for signature 'GRangesList'
addDiffStatus(x, ...)

## S4 method for signature 'SummarizedExperiment'
addDiffStatus(x, ...)

Arguments

x
Object to be classified
...
Used to pass arguments between methods
fc_col
Name of the fold-change column
sig_col
Name of the column with significance values
alpha
significance threshold
cutoff
minimum estimated change to be considered in either of the up or down categories
up, down, other
factor levels to annotate regions based on the above criteria
missing
Value to add when either fc_col or sig_col has NA values
new_col
name of the new column to be added
drop
logical(1) Drop unused factor levels from the status column

Details

This takes a simple object and adds a new column classifying entries into one of three categories, as specified using up, down or other. Results in the new column will always be returned as a factor with levels in order of the values provided in the arguments other, down and up.

Value

An object of the same type as provided

Examples

## Working with a data.frame
set.seed(101)
df <- data.frame(logFC = rnorm(20), p = rbeta(20, shape1 = 1, shape2 = 20))
```r
df$FDR <- p.adjust(df$p, "fdr")
addDiffStatus(df)

## This works identically with a GRanges object, amongst others
gr <- GRanges(paste0("chr1:", seq_len(20)))
mcols(gr) <- df
addDiffStatus(gr)
```

---

**as_tibble**

Convert to a tibble

---

**Description**

Convert multiple Genomic objects to tibbles

**Usage**

```r
## S3 method for class 'DataFrame'
as_tibble(x, rangeAsChar = TRUE, ...)

## S3 method for class 'GenomicRanges'
as_tibble(x, rangeAsChar = TRUE, name = "range", ...)

## S3 method for class 'Seqinfo'
as_tibble(x, ...)

## S3 method for class 'GInteractions'
as_tibble(x, rangeAsChar = TRUE, suffix = c(".x", ".y"), ...)

## S3 method for class 'SummarizedExperiment'
as_tibble(x, rangeAsChar = TRUE, ...)

## S3 method for class 'TopTags'
as_tibble(x, ...)
```

**Arguments**

- **x** A Genomic Ranges or DataFrame object
- **rangeAsChar** Convert any GRanges element to a character vector
- **...** Passed to `tibble::as_tibble()`
- **name** Name of column to use for ranges. Ignored if rangeAsChar = FALSE
- **suffix** Suffix appended to column names for anchor1 and anchor2 of a GInteractions object. Only used if specifying rangeAsChar = FALSE
Details

Quick and dirty conversion into a tibble.

By default, GenomicRanges will be returned with the range as a character column called `range` and all mcols parsed as the remaining columns. Seqinfo information will be lost during coercion.

Given that names for ranges are considered as rownames in the mcols element, these can be simply parsed by setting `rownames = "id"` in the call to `as_tibble()`

When coercing a DataFrame, any Compressed/SimpleList columns will be coerced to S3 list columns. Any GRanges columns will be returned as a character column, losing any additional mcols from these secondary ranges

Coercion of SummarizedExperiment objects will be performed on the `rowRanges()` element, whilst for a GInteractions object, both anchors will returned with the default suffixes `.x` and `.y`

Defined as an S3 method for consistency with existing tidy methods

Value

A tibble

Examples

```r
gr <- GRanges("chr1:1-10")
gr$p_value <- runif(1)
names(gr) <- "range1"
gr
as_tibble(gr)
as_tibble(gr, rownames = "id")
as_tibble(mcols(gr))
as_tibble(seqinfo(gr))

hic <- InteractionSet::GInteractions(gr, GRanges("chr1:201-210"))
hic$id <- "interaction1"
as_tibble(hic)
```

---

**bestOverlap**

**Find the best overlap between GRanges**

Description

Find the best overlap between ranges

Usage

```r
bestOverlap(x, y, ...)
```

## S4 method for signature 'GRanges,GRanges'

```r
bestOverlap(
```
Arguments

x a GRanges object

y a named GRangesList or GRanges object with mcol as reference category

... Not used

var The variable to use as the category. Not required if y is a GRangesList

ignore.strand logical(1) Passed to findOverlaps

missing Value to assign to ranges with no overlap

min_prop Threshold below which overlaps are discarded

Details

This finds the category in the subject GRanges (y) which has the best overlap with the query GRanges (x). The aim is to produce a character vector for best classifying the query GRanges using an external set of features (e.g. promoters, enhancers etc). If the subject (y) is a GRanges object, the values in the specified column will be used as the category. If the subject (y) is a GRangesList, the names of the list will be used to provide the best match.

Value

Character vector the same length as the supplied GRanges object

Examples

gr <- GRanges("chr1:1-10")
gr_cat <- GRanges(c("chr1:2-10", "chr1:5-10"))
gr_cat$category <- c("a", "b")
propOverlap(gr, gr_cat)
bestOverlap(gr, gr_cat, var = "category")
`chopMC`  

```r
gr <- splitAsList(gr_cat, gr_cat$category)
lapply(grl, function(x) propOverlap(gr, x))
bestOverlap(gr, grl)
```

---

### Description

Keep unique ranges by 'chopping' mcols

### Usage

```r
chopMC(x, simplify = TRUE)
```

### Arguments

- **x**: A GenomicRanges object
- **simplify**: logical(1)

### Details

This function finds unique ranges and chops all mcols in a manner similar to `chop`. Chopped columns will be returned as CompressedList columns, unless simplify = TRUE (the default). In this case, columns will be returned as vectors where possible.

### Value

A GRanges object

### Examples

```r
gr <- GRanges(rep("chr1:1-10"), 2)
gr$id <- paste0("range", seq_along(gr))
gr$gene <- "gene1"
gr
chopMC(gr)
```
collapseGenes

**Description**

Collapse a vector of gene names

**Usage**

```r
collapseGenes(
  x,
  sort = TRUE,
  dedup = TRUE,
  format = "_",
  sep = "",
  last = " and ",
  numeric = TRUE,
  width = Inf,
  ...
)
```

**Arguments**

- **x**: character vector representing gene names
- **sort**: logical(1) Should the names be sorted alphabetically
- **dedup**: logical(1) Should duplicate names be removed
- **format**: character string for markdown formatting of each element
- **sep**: separator between vector elements
- **last**: character string to place before the last element
- **numeric**: logical(1) sort digits numerically, instead of as strings
- **width**: The maximum width of the string before truncating to ...

**Details**

Convenience function to collapse a vector of gene names into a character/glue object of length 1. By default, symbols are deduplicated, sorted alpha-numerically and italicised with an underscore.

**Value**

- a glue object

**Examples**

```r
genes <- c("FOXP3", "BRCA1", "TP53")
collapseGenes(genes)
```
**colToRanges**

Coerce a column to a GRanges object

**Description**

Coerce a column to a GRanges object from a rectangular object.

**Usage**

```r
colToRanges(x, ...)  
## S4 method for signature 'DataFrame'
colToRanges(x, var, seqinfo = NULL, ...)  
## S4 method for signature 'GRanges'
colToRanges(x, var, ...)  
## S4 method for signature 'data.frame'
colToRanges(x, var, seqinfo = NULL, ...)  
```

**Arguments**

- `x`: A data-frame or GRanges object containing the column to coerce.
- `...`: Used to pass arguments to lower-level functions.
- `var`: The name of the column to coerce.
- `seqinfo`: A seqinfo object to be applied to the new GRanges object. Ignored if the column is already a GRanges object.

**Details**

Take a data.frame-like object and coerce one column to a GRanges object, setting the remainder as the mcols. A particularly useful application of this is when you have a GRanges object with one mcol being a secondary GRanges object.

Alternatively, if you have a data.frame with GRanges represented as a character column, this provides a simple method of coercion. In this case, no Seqinfo element will be applied to the GRanges element.

**Value**

A GenomicRanges object.

**Examples**

```r
set.seed(73)
x <- GRanges(c("chr1:1-10", "chr1:6-15", "chr1:51-60"))
seqinfo(x) <- Seqinfo("chr1", 60, FALSE, "Example")
df <- data.frame(logFC = rnorm(3), logCPM = rnorm(3, 8), p = 10^-rexp(3))
```
cytobands

Cytogenetic bands

Description

Cytogenetic bands for GRCh37/hg19 and GRCh38/hg38

Usage

data(grch37.cytobands)
data(grch38.cytobands)

Format

Cytogenetic bands for standard chromosomes from GRCh37. in the format required by Ideogram-Track. A data.frame with 5 columns:

- **chrom** Chromosome
- **chromStart** Starting position for each cytogenetic band
- **chromEnd** End position for each cytogenetic band
- **name** Name for each band, e.g. p.36.33
- **gieStain** Staining pattern

An object of class data.frame with 862 rows and 5 columns.

Source

https://hgdownload.soe.ucsc.edu/goldenPath/hg19/database/cytoBand.txt.gz
https://hgdownload.soe.ucsc.edu/goldenPath/hg38/database/cytoBand.txt.gz

Examples

data(grch37.cytobands)
head(grch37.cytobands)

data(grch38.cytobands)
head(grch38.cytobands)
defineRegions

Define Genomic Regions Based on Gene Annotations

Description

Use gene, transcript and exon annotations to define genomic regions

Usage

```r
defineRegions(
  genes, transcripts, exons,
  promoter = c(2500, 500),
  upstream = 5000,
  intron = TRUE,
  proximal = 10000,
  simplify = FALSE,
  cols = c("gene_id", "gene_name", "transcript_id", "transcript_name"),
  ...
)
```

Arguments

- `genes, transcripts, exons` GRanges objects defining each level of annotation
- `promoter` Numeric vector defining upstream and/or downstream distances for a promoter. Passing a single value will define a symmetrical promoter. The first value represents the upstream range
- `upstream` The distance from a TSS defining an upstream promoter
- `intron` logical(1) Separate gene bodies into introns and exons. If `intron = FALSE` gene bodies will simply be defined as gene bodies
- `proximal` Distance from a gene to be considered a proximal intergenic region. If set to 0, intergenic regions will simply be considered as uniformly intergenic
- `simplify` Passed internally to `reduceMC` and `setdiffMC`
- `cols` Column names to be retained from the supplied annotations
- `...` Not used

Details

Using GRanges annotated as genes, transcripts and exons this function will define ranges uniquely assigned to a region type using a hierarchical process. By default, these region types will be (in order) 1) Promoters, 2) Upstream Promoters, 3) Exons, 4) Introns, 5) Proximal Intergenic and 6) Distal Intergenic.
Setting `intron = FALSE` will replace introns and exons with a generic "Gene Body" annotation. Setting `proximal = 0` will return all intergenic regions (not previously annotated as promoters or upstream promoters) to an "Intergenic" category.

Notably, once a region has been defined, it is excluded from all subsequent candidate regions.

Any columns matching the names provided in `cols` will be returned, and it is assumed that the gene/transcript/exon ranges will contain informative columns in the `mcols()` element.

**Value**

A GRangesList

**Examples**

```r
## Define two exons for two transcripts
sq <- Seqinfo(seqnames = "chr1", seqlengths = 50000)
e <- GRanges(e, seqinfo = sq)
mcols(e) <- DataFrame(
  gene_id = "Gene1", transcript_id = paste0("Trans", c(1, 1, 2, 2))
)

## Define the transcript ranges
t <- unlist(endoapply(split(e, e$transcript_id), range))
t$gene_id <- "Gene1"
t$transcript_id <- names(t)
names(t) <- NULL

## Summarise to gene level
g <- reduceMC(t)
g$transcript_id <- NA_character_

## Now annotate the regions
regions <- defineRegions(genes = g, transcripts = t, exons = e)
sort(unlist(regions))

## Alternatively, collapse gene body and intergenic ranges
regions <- defineRegions(
  genes = g, transcripts = t, exons = e, intron = FALSE, proximal = 0
)
sort(unlist(regions))
```

---

**defineSeqinfo**

*Use package data to define a Seqinfo object*

**Description**

Use package data to define a Seqinfo object
distinctMC

Usage

defineSeqinfo(
  build = c("GRCh38", "GRCh37", "GRCm39", "GRCm38"),
  chr = TRUE,
  mito,
  ...
)

Arguments

build          The Genome build used
chr            logical(1) Include the prefix "chr"
mito           Specify M or MT to include the mitochondrial chromosome. Omitted by default
...            Not used

Details

This function will create a Seqinfo object from pre-defined data from the Genome Reference Consortium. Returned objects will always be restricted to assembled molecules only. Currently implemented genome builds represent the four most common builds for ChIP-Seq analysis

Value

A Seqinfo object

Examples

defineSeqinfo("GRCh37", TRUE)
defineSeqinfo("GRCh37", FALSE, "MT")

distinctMC

Keep distinct ranges and mcols

Description

Keep distinct ranges by including mcols

Usage

distinctMC(x, ..., .keep_all = FALSE)

Arguments

x            A GenomicRanges object
...          <data-masking> Passed to distinct
.keep_all    If TRUE, keep all columns in x
Details

Wrapper to `distinct` for `GRanges` objects. Finds unique ranges and mcols in combination and retains only the distinct combinations, in keeping with the `dplyr` function.

Will default to `unique(granges(x))` if no columns are provided

Value

A `GRanges` object

Examples

```r
gr <- GRanges(rep(c("chr1:1-10"), 2))
gr$id <- paste0("range", seq_along(gr))
gr$gene <- "gene1"
gr
distinctMC(gr)
distinctMC(gr, gene)
distinctMC(gr, gene, .keep_all = TRUE)
```

---

**dualFilter**

Apply two filters to sliding windows

Description

Apply two filters to counts generated using sliding windows

Usage

dualFilter(
  x,
  bg = NULL,
  ref,
  q = 0.5,
  logCPM = TRUE,
  keep.totals = TRUE,
  bin.size = NULL,
  prior.count = 2,
  BPPARAM = bpparam()
)

Arguments

- `x` RangedSummarizedExperiment containing sample counts
- `bg` RangedSummarizedExperiment containing background/input counts, or alternate method for selecting samples from within `x`, such as a logical, numeric or character vector
dualFilter

Details

This function will take sliding (or tiling) windows for it's input as a RangedSummarizedExperiment object. The dual strategy of applying filterWindowsControl and filterWindowsProportion will then be applied. A set of reference ranges for which signal is expected is used to refine the filtering criteria.

Cutoff values are found for both signal relative to input and overall signal, such that the 100*q% of the (sliding) windows which overlap a reference range will be returned, along with any others which match the dual filtering criteria. In general, higher values of q will return more windows as those with weak signal and a marginal overlap with a reference range will be returned. Lower values will ensure that fewer windows, generally with the strongest signal, are retained. Cutoff values for both criteria are added to the metadata element of the returned object.

If setting bg = NULL the filterWindowsControl step will be ignored and only the filterWindowsProportion will be used. This should only be performed if no Input sample is available.

Please note that the any .bam files referred to in the supplied objects must be accessible to this function. It will not run on a separate machine or file structure to that which the original sliding windows were prepared. Please see the example/vignette for runnable code.

Value

A RangedSummarizedExperiment which is a filtered subset of the original object. If requested the assay "logCPM" will be added (TRUE by default)

Examples

```r
## Taken from the differential_binding vignette
library(tidyverse)
library(Rsamtools)
library(csaw)
library(BiocParallel)
library(rtracklayer)
## For this function we need a set of counts using sliding windows and the
## original BamFiles from which they were taken
## First we'll set up the bam file list
bfl <- system.file(
    "extdata", "bam", c("ex1.bam", "ex2.bam", "input.bam"), package = "extraChIPs"
)
## Then define the readParam settings for csaw::readParam()

```r
rp <- readParam(
  pe = "none",
  dedup = TRUE,
  restrict = "chr10"
)
```

## Now we can form our sliding window object with the counts.

```r
wincounts <- windowCounts(
  bam.files = bfl,
  spacing = 60,
  width = 180,
  ext = 200,
  filter = 1,
  param = rp
)
```

## As this is a subset of reads, add the initial library sizes for accuracy

```r
wincounts$totals <- c(964076L, 989543L, 1172179L)
```

## We should also update the metadata for our counts

```r
wincounts$sample <- colnames(wincounts)
wincounts$treat <- as.factor(c("ctrl", "treat", NA))
colData(wincounts)
```

The function dualFilter requires a set of peaks which will guide the
filtering step. This indicate where genuine signal is likely to be found
and will perform the filtering based on a) signal above the input, and
b) The overall signal level, using the guide set of peaks to inform the
cutoff values for inclusion

```r
peaks <- import.bed(
  system.file("extdata", "peaks.bed.gz", package = "extraChIPs")
)
```

```r
filtcounts <- dualFilter(
  x = wincounts, bg = "input", ref = peaks,
  q = 0.8 # Better to use q = 0.5 on real data
)
```

```r
colData(filtcounts)
```
**Description**

Various example datasets for demonstrating analysis and visualisation strategies. Generation of all datasets is documented in `system.file("script/ex_datasets.md", package = "extraChIPs")`

- **ex_genes** Simple GRanges object with complete ranges for each gene
- **ex_trans** Exon & transcript level information prepared for plotting with `Gviz` or `plotHFGC()`
- **ex_prom** Regions defined as promoters
- **ex_hic** Example HiC interactions

**Usage**

```r
data(ex_trans)
data(ex_genes)
data(ex_prom)
data(ex_hic)
```

**Format**

GRanges and GInteractions objects

- All annotations are from GRCh37
- An object of class GRanges of length 4.
- An object of class GRanges of length 9.
- An object of class GInteractions of length 1.

**Examples**

```r
data(ex_trans)
ex_trans
```

---

**fitAssayDiff**

*Detect Differential ChIP Signal*

**Description**

Detect differential ChIP signal using one of many approaches
Usage

```r
fitAssayDiff(x, ...)  
```

## S4 method for signature 'SummarizedExperiment'
```r
fitAssayDiff(
  x,
  assay = "counts",
  design = NULL,
  coef = NULL,
  lib.size = "totals",
  method = c("qlf", "lt"),
  norm = c("none", "TMM", "RLE", "TMMwsp", "upperquartile"),
  groups = NULL,
  fc = 1,
  lfc = log2(fc),
  asRanges = FALSE,
  offset = NULL,
  null = c("interval", "worst.case"),
  weighted = FALSE,
  ...
)
```

Arguments

- `x`: a SummarizedExperiment object
- `...`: Passed to `calcNormFactors` and `glmQLFit` when `method = "qlf"`. If `method = "lt"`, instead passed to `lmFit`, `treat`, `eBayes`
- `assay`: The assay to use for analysis
- `design`: The design matrix to use for analysis
- `coef`: The required column from the design matrix
- `lib.size`: The column within the colData element which contains the library size information. If set to NULL, column summaries will be used.
- `method`: the analytic method to be used. Can be 'qlf' which will fit counts using the `glmQLFit` strategy, or 'lt' which fits the limma-trend model on logCPM, or pre-processed logCPM values
- `norm`: The normalisation strategy to use when running the glmQLF models. The value 'none' relies solely on library-size normalisation, and is the default. All methods available in `calcNormFactors` are implemented. Ignored when using `method = "lt"`
- `groups`: character(1) If a column name is supplied here, group-based normalisation will be applied to GLM models treating data in this column as a grouping factor. Ignored when using `method = "lt"`
- `fc, lfc`: Thresholds passed to `treat` or `glmTreat`
- `asRanges`: logical(1). By default, the returned object will be a SummarizedExperiment object with the results added to the rowData element. Setting `asRanges = TRUE` will only return the GRanges object from this element
offset If provided will be used as the offset when the DGEList object is created during model fitting
null Passed to glmTreat
weighted logical(1) Passed to calcNormFactors

Details
Starting with a SummarizedExperiment object this function fits either a glmQLFit model to count data, or the limma-trend model to logCPM data.

If fitting Generalised Linear Models via glmQLFit, options for normalisation are "none", which normalises to library size. Existing library sizes are commonly found in the "totals" column of the colData element and this is attempted by default. All methods provided in calcNormFactors are also implemented, with the added possibility of normalising within groups instead of across the entire dataset. To enable this, the column with the grouping factor is expected to be in the colData element and is simply called by column name. No normalisation is applied when using the limma-trend model, as this allows for previous normalisation strategies to be performed on the data.

Normalising to ChIP Input samples, or using offsets is not yet implemented.

Either range-based hypothesis testing is implemented using glmTreat or treat. Setting fc to 1 (or lfc to 0) will default to a point-based null hypothesis, equivalent to either glmQLFTest (method = "qlf") or eBayes (method = "lt").

It should also be noted that this is primarily a convenience function and if requiring intermediate output from any steps, then these can be run individually as conventionally specified.

Value
A SummarizedExperiment object with results set as the rowData element. Any existing columns not contained in the differential ChIP results will be retained. Results from testing will contain logCPM, logFC, PValue and the t/F statistic as appropriate, along with an FDR-adjusted p-value.

If specifying a range-based H0 by setting lfc != 0, an additional column p_mu0 will be included which is the p-value for the point H0: logFC = 0. These are not used for FDR-adjusted p-values but can be helpful when integrating multiple ChIP targets due to the increase in false-negatives when using a range-based H0, and when requiring more accurate identification of truly unchanged sites, as opposed to those which simply fail to achieve significance using a range-based H0 where arbitrary cutoff values are used.

Examples
```r
nrows <- 200; ncols <- 6
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
colnames(counts) <- paste0("Sample_", seq_len(ncols))
df <- DataFrame(treat = c("A", "A", "A", "B", "B", "B"))
df$treat <- as.factor(df$treat)
se <- SummarizedExperiment(
  assays = SimpleList(counts = counts), colData = df
)
X <- model.matrix(~treat, colData(se))
se <- fitAssayDiff(se, design = X, lib.size = NULL)
rowData(se)
```
fixed_width_datasets  *Datasets for the Fixed-Width Vignette*

**Description**

GRangesList of peaks and SummarizedExperiment of counts. All were saved during initial vignette preparation at https://github.com/smped/extraChIPs_vignette/blob/main/differential_signal_fixed.Rmd

**Usage**

data(se)

data(peaks)

**Format**

An object of class RangedSummarizedExperiment with 188 rows and 6 columns.

An object of class CompressedGRangesList of length 6.

**Examples**

data(se)
se
data(peaks)
peaks

---

**getProfileData**  *Get Profile Data surrounding specified ranges*

**Description**

Get coverage Profile Data surrounding specified ranges

**Usage**

getProfileData(x, gr, ...)

```r
## S4 method for signature 'BigWigFile,GenomicRanges'
getProfileData(
  x,
  gr,
  upstream = 2500,
  downstream = upstream,
```
getProfileData

```r
getProfileData(
  x,
  gr,
  upstream = 2500,
  downstream = upstream,
  bins = 100,
  mean_mode = "w0",
  log = TRUE,
  offset = 1,
  BPPARAM = SerialParam()
)
```

Arguments

- `x` A BigWigFile or BigWigFileList
- `gr` A GRanges object
- `...` Passed to `normalizeToMatrix`
- `upstream` The distance to extend upstream from the centre of each range within `gr`
- `downstream` The distance to extend downstream from the centre of each range within `gr`
- `bins` The total number of bins to break the extended ranges into
- `mean_mode` The method used for calculating the score for each bin. See `normalizeToMatrix` for details
- `log` logical(1) Should the returned values be log2-transformed
offset

Value added to data if log-transforming. Ignored otherwise

n_max

Upper limit on the number of ranges to return profile data for. By default, no limit will be applied.

BPPARAM

Passed internally to bplapply

Details

This will take all provided ranges and set as identical width ranges, extending by the specified amount both up and downstream of the centre of the provided ranges. By default, the ranges extensions are symmetrical and only the upstream range needs to be specified, however this parameterisation allows for non-symmetrical ranges to be generated.

These uniform width ranges will then be used to extract the value contained in the score field from one or more BigWigFiles. Uniform width ranges are then broken into bins of equal width and the average score found within each bin.

The binned profiles are returned as a DataFrameList called profile_data as a column within the resized GRanges object. Column names in each DataFrame are score, position and bp.

If passing a BigWigFileList, profiles will be obtained in series by default. To run in parallel pass a MulticoreParam object to the BPPARAM argument.

Value

GRanges or GrangesList with column profile_data, as described above

Examples

bw <- system.file("tests", "test.bw", package = "rtracklayer")
gr <- GRanges("chr2:1000")
pd <- getProfileData(bw, gr, upstream = 500, bins = 10)
pd
pd$profile_data

---

grlToSE

Set columns from a GRangesList as Assays in a SummarizedExperiment

Description

Move one or more columns from a GRangesList elements into assays in a RangesSummarizedExperiment
Usage

grlToSE(x, ...)

## S4 method for signature 'GRangesList'
grlToSE(
  x,
  assayCols = c(),
  metaCols = c(),
  keyvals = c(),
  by = c("min", "max"),
  ...
)

Arguments

x A GrangesList

... Passed to reduce

assayCols Columns to move to separate assays

metaCols Columns to move to mcols within the rowRanges element

keyvals The value to use when choosing representative values

by How to choose by keyvals

ignore.strand logical(1). Whether the strand of the input ranges should be ignored or not.

Details

Given a GRangesList which would commonly represent multiple samples, reduce any overlapping ranges into a consensus range, setting any metadata columns to be retained as separate assays. These columns may contain values such as coverage, p-values etc.

Additional columns can also be placed asrowData columns where the original values are better suited to information about the consensus range rather than the sample (or GRangesList element).

Only one value for each range will be retained, and these are chosen using the value provided as the keyvals, taking either the min or max value in this column as the representative range.

Value

A RangedSummarizedExperiment

Examples

```r
a <- GRanges("chr1:1-10")
a$feature <- "Gene"
a$p <- 0.1
b <- GRanges(c("chr1:6-15", "chr1:15"))
b$feature <- c("Gene", "Promoter")
b$p <- c(0.5, 0.01)
grl <- GRangesList(a = a, b = b)
```
importPeaks

se <- grlToSE(
  grl, assayCols = "p", metaCols = "feature", keyvals = "p", by = "min"
)
assay(se, "p")
rowRanges(se)

---

**importPeaks**

**Import peaks**

**Description**

Import peaks in narrowPeak, broadPeak or bed format

**Usage**

importPeaks(
  x,
  type = c("narrow", "broad", "bed"),
  blacklist,
  seqinfo,
  pruning.mode = c("coarse", "error"),
  sort = TRUE,
  setNames = TRUE,
  glueNames = "{basename(x)}",
  centre = FALSE,
  nameRanges = TRUE,
  ...
)

**Arguments**

- **x**: One or more files to be imported. All files must be of the same type, i.e. narrow or broad
- **type**: The type of peaks to be imported
- **blacklist**: A set of ranges to be excluded
- **seqinfo**: A seqinfo object to be applied to the GRanges objects
- **pruning.mode**: How to handle conflicts if supplying a seqinfo object. Defaults to pruning.mode = "coarse". Only "coarse" and "error" are implemented. See seqinfo.
- **sort**: logical. Should the ranges be sorted during import
- **setNames**: logical Set basename(x) as the name for each element of the GRangesList
- **glueNames**: glue syntax for naming list elements
- **centre**: Add the estimated peak centre. Ignored unless type = "narrow"
- **nameRanges**: Place any values in the name column as range names within each file.
- **...**: passed to sort
makeConsensus

Details
Peaks are imported from narrowPeak, broadPeak or bed format as GenomicRanges objects. If importing bed files, only the default 3-6 columns will imported.

Value
A GRangesList

Examples
fl <- system.file(
  c("extdata/ER_1.narrowPeak", "extdata/ER_2.narrowPeak"),
  package = "extraChIPs"
)
peaks <- importPeaks(fl)
peaks

makeConsensus  Make a set of consensus peaks

Description
Make a set of consensus peaks based on the number of replicates

Usage
makeConsensus(
  x,
  p = 0,
  var = NULL,
  method = c("union", "coverage"),
  ignore.strand = TRUE,
  simplify = FALSE,
  min_width = 0,
  ...
)

Arguments
x  A GRangesList
p  The minimum proportion of samples (i.e. elements of x) required for a peak to be retained in the output. By default all merged peaks will be returned
var  Additional columns in the mcols element to retain
method  Either return the union of all overlapping ranges, or the regions within the overlapping ranges which are covered by the specified proportion of replicates. When using p = 0, both methods will return identical results
ignore.strand, simplify, ...
    Passed to reduceMC or intersectMC internally

min_width    Discard any regions below this width

Details

This takes a list of GRanges objects and forms a set of consensus peaks.

When using method = "union" the union ranges of all overlapping peaks will be returned, using
the minimum proportion of replicates specified. When using method = "coverage", only the re-
gions within each overlapping range which are 'covered' by the minimum proportion of replicates
specified are returned. This will return narrower peaks in general, although some artefactual very
small ranges may be included (e.g. 10bp). Careful setting of the min_width parameter may be very
helpful for these instances. It is also expected that setting method = "coverage" should return the
region within each range which is more likely to contain the true binding site for the relevant ChIP
targets

Value

GRanges object with mcols containing a logical vector for every element of x, along with the column
n which adds all logical columns. These columns denote which replicates contain an overlapping
peak for each range

If any additional columns have been requested using var, these will be returned as CompressedList
objects as produced by reduceMC() or intersectMC().

See Also

reduceMC intersectMC

Examples

data("peaks")
## The first three replicates are from the same treatment group
grl <- peaks[1:3]
names(grl) <- gsub("_peaks.+", "", names(grl))
makeConsensus(grl)
makeConsensus(grl, p = 2/3, var = "score")

## Using method = 'coverage' finds ranges based on the intersection
makeConsensus(grl, p = 2/3, var = "score", method = "coverage")
mapByFeature

Map Genomic Ranges to genes using defined features

Description

Map Genomic Ranges to genes using defined regulatory features

Usage

mapByFeature(
  gr,
  genes,
  prom,
  enh,
  gi,
  cols = c("gene_id", "gene_name", "symbol"),
  gr2prom = 0,
  gr2enh = 0,
  gr2gi = 0,
  gr2gene = 1e+05,
  prom2gene = 0,
  enh2gene = 1e+05,
  gi2gene = 0,
  ...
)

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>gr</td>
<td>GRanges object with query ranges to be mapped to genes</td>
</tr>
<tr>
<td>genes</td>
<td>GRanges object containing genes (or any other nominal feature) to be assigned</td>
</tr>
<tr>
<td>prom</td>
<td>GRanges object defining promoters</td>
</tr>
<tr>
<td>enh</td>
<td>GRanges object defining Enhancers</td>
</tr>
<tr>
<td>gi</td>
<td>GInteractions object defining interactions. Mappings from interactions to genes should be performed as a separate prior step.</td>
</tr>
<tr>
<td>cols</td>
<td>Column names to be assigned as mcols in the output. Columns must be minimally present in genes. If all requested columns are found in any of prom, enh or gi, these pre-existing mappings will be preferentially used. Any columns not found in utilised reference objects will be ignored.</td>
</tr>
<tr>
<td>gr2prom</td>
<td>The maximum permissible distance between a query range and any ranges defined as promoters</td>
</tr>
<tr>
<td>gr2enh</td>
<td>The maximum permissible distance between a query range and any ranges defined as enhancers</td>
</tr>
<tr>
<td>gr2gi</td>
<td>The maximum permissible distance between a query range and any ranges defined as GInteraction anchors</td>
</tr>
</tbody>
</table>
mapByFeature

gr2gene The maximum permissible distance between a query range and genes (for ranges not otherwise mapped)

prom2gene The maximum permissible distance between a range provided in prom and a gene

enh2gene The maximum permissible distance between a range provided in enh and a gene

gi2gene The maximum permissible distance between a GInteractions anchor (provided in gi) and a gene

Passed to findOverlaps and overlapsAny internally

Details

This function is able to utilise feature-level information and long-range interactions to enable better mapping of regions to genes. If provided, this essentially maps from ranges to genes using the regulatory features as a framework. The following sequential strategy is used:

1. Ranges overlapping a promoter are assigned to that gene
2. Ranges overlapping an enhancer are assigned to all genes within a specified distance
3. Ranges overlapping a long-range interaction are assigned to all genes connected by the interaction
4. Ranges with no gene assignment from the previous steps are assigned to all overlapping genes or the nearest gene within a specified distance

If information is missing for one of these steps, the algorithm will simply proceed to the next step. If no promoter, enhancer or interaction data is provided, all ranges will be simply mapped by step 4. Ranges can be mapped by any or all of the first three steps, but step 4 is mutually exclusive with the first 3 steps.

Distances between each set of features and the query range can be individually specified by modifying the gr2prom, gr2enh, gr2gi or gr2gene parameters. Distances between features and genes can also be set using the parameters prom2gene, enh2gene and gi2gene.

Additionally, if previously defined mappings are included with any of the prom, enh or gi objects, this will be used in preference to any obtained from the genes object.

Value

A GRanges object with added mcols as specified

Examples

```r
## Define some genes
genes <- GRanges(c("chr1:2-10:*", "chr1:25-30:-", "chr1:31-40:+"))
genes$gene_id <- paste0("gene", seq_along(genes))
genes

## Add a promoter for each gene
prom <- promoters(genes, upstream = 1, downstream = 1)
prom

## Some ranges to map
gr <- GRanges(paste0("chr1:", seq(0, 60, by = 15)))
gr
```

Passed to findOverlaps and overlapsAny internally
## Map so that any gene within 25bp of the range is assigned
mapByFeature(gr, genes, gr2gene = 25)

## Now use promoters to be more accurate in the gene assignment
## Given that the first range overlaps the promoter of gene1, this is a
## more targeted approach. Similarly for the third range
mapByFeature(gr, genes, prom, gr2gene = 25)

---

### mapGrlCols

**Collapse a GRangesList adding multiple columns from each element**

#### Description

Make consensus peaks and add individual columns from each original GRangesList element

#### Usage

```r
mapGrlCols(x, var = NULL, collapse = NULL, collapse_sep = "; ", name_sep = ", ", include = FALSE, ...)
```

#### Arguments

- **x** | GRangesList
- **var** | Column(s) to map onto the set of consensus peaks
- **collapse** | Columns specified here will be simplified into a single column. Should only be character or factor columns
- **collapse_sep** | String to separate values when collapsing columns
- **name_sep** | String to separate values when adding column names
- **include** | logical(1) Include the original ranges as character columns
- **...** | Passed to makeConsensus

#### Details

Starting with a GRangesList, make a single GRanges object with select columns from each element added to the new object
mergeByCol

Merge sliding windows using a specified column

Description

Merge sliding windows using a specified column

Usage

mergeByCol(x, ...)

## S4 method for signature 'GenomicRanges'
mergeByCol(
  x,

mergeByCol

```r
df = NULL,
col,
by = c("max", "median", "mean", "min"),
logfc = "logFC",
pval = "p",
inc_cols,
p_adj_method = "fdr",
merge_within = 1L,
ignore_strand = TRUE,
min_win = 1,
...
)

## S4 method for signature 'RangedSummarizedExperiment'
mergeByCol(
  x,
  df = NULL,
col,
by = c("max", "median", "mean", "min"),
logfc = "logFC",
pval = "p",
inc_cols,
p_adj_method = "fdr",
merge_within = 1L,
ignore_strand = FALSE,
...
)
```

### Arguments

- **x**
  - A GenomicRanges or SummarizedExperiment object
- **...**
  - Not used
- **df**
  - A data.frame-like object containing the columns of interest. If not provided, any columns in the mcols() slot will be used.
- **col**
  - The column to select as representative of the merged ranges
- **by**
  - The method for selecting representative values
- **logfc**
  - Column containing logFC values
- **pval**
  - Column containing p-values
- **inc_cols**
  - Any additional columns to return. Output will always include columns specified in the arguments `col`, `logfc` and `pval`. Note that values from any additional columns will correspond to the selected range returned in `keyval_range`
- **p_adj_method**
  - Any of `p.adjust.methods`
- **merge_within**
  - Merge any ranges within this distance
- **ignore_strand**
  - Passed internally to `reduce` and `findOverlaps`
- **min_win**
  - Only keep merged windows derived from at least this number
Details
This merges sliding windows using the values in a given column to select representative values for the subsequent merged windows. Values can be chosen from the specified column using any of \( \text{min}() \), \( \text{max}() \), \( \text{mean}() \) or \( \text{median}() \), although \( \text{max}() \) is strongly recommended when specifying values like logCPM. Once a representative range is selected using the specified column, values from columns specified using \( \text{inc_cols} \) are also returned. In addition to these columns, the range from the representative window is returned in the \text{mcols} element as a GRanges object in the column \text{keyval_range}.

Merging windows using either the logFC or p-value columns is not implemented.

If adjusted p-values are requested an additional column names the same as the initial p-value, but tagged with the adjustment method, will be added. In addition, using the p-value from the selected window, the number of windows with lower p-values are counted by direction and returned in the final object. The selected window will always be counted as up/down regardless of significance as the p-value for this column is taken as the threshold. This is a not dissimilar approach to \text{cluster-direction}.

If called on a SummarizedExperiment object, the function will be applied to the \text{rowRanges} element.

Value
A Genomic Ranges object

Examples
```r
x <- GRanges(c("chr1:1-10", "chr1:6-15", "chr1:51-60"))
set.seed(1001)
df <- DataFrame(logFC = rnorm(3), logCPM = rnorm(3,8), p = rexp(3, 10))
mergeByCol(x, df, col = "logCPM", pval = "p")
mcols(x) <- df
x
mergeByCol(x, col = "logCPM", pval = "p")
```

mergeByHMP

Merge Sliding Windows using the Harmonic Mean P

Description
Merge overlapping windows using harmonic mean p-values from significance testing

Usage
```r
mergeByHMP(x, ...)
```

## S4 method for signature 'GenomicRanges'
mergeByHMP(
x,  
...  
)
mergeByHMP

```r
df = NULL,
w = NULL,
logfc = "logFC",
pval = "P",
cpm = "logCPM",
inc_cols = NULL,
p_adj_method = "fdr",
merge_within = 1L,
ignore_strand = TRUE,
min_win = 1,
keyval = c("min", "merged"),
...
"
)
```

```r
## S4 method for signature 'RangedSummarizedExperiment'
mergeByHMP(
  x,
  df = NULL,
w = NULL,
logfc = "logFC",
pval = "P",
cpm = "logCPM",
inc_cols = NULL,
p_adj_method = "fdr",
merge_within = 1L,
ignore_strand = FALSE,
...
)
```

### Arguments

- **x**
  - GenomicRanges object
- **df**
  - data.frame with results of differential binding analysis performed using a sliding window strategy. If not provided, the columns in the `mcols()` element of x will be used
- **w**
  - vector of weights to applied when calculating harmonic mean p-values
- **logfc, pval, cpm**
  - Column names for the values holding window specific estimates of change in binding (logfc), overall signal intensity (cpm) and the significance from statistical testing (pval)
- **inc_cols**
  - (Optional) Character vector of any additional columns in df to return. Values will correspond to the range in the keyval_range column
- **p_adj_method**
  - One of `p.adjust.methods` or "fwer". If "fwer" is specified the adjusted harmonic-mean p-value will be returned in a form which strictly controls the experiment-wide FWER. Please see vignette("harmonicmeanz") for more details
- **merge_within**
  - Merge any non-overlapping windows within this distance
mergeByHMP

ignore_strand  Passed internally to reduce and findOverlaps
min_win       Only keep merged windows derived from at least this number
keyval        Return the key-value range as the window associated with the minimum p-value,
or by merging the ranges from all windows with raw p-values below the merged harmonic-mean p-value

Details

When using sliding windows to test for differential signal, overlapping windows can be merged based on the significance of results. mergeByHMP() merges overlapping windows using the asymptotically exact harmonic mean p-value p.hmp from the individual, window-level tests. This tests the Null Hypothesis that there is no significance amongst the initial set of p-values, and returns a summarised value which controls the FDR within a set of tests (Wilson, PNAS, 2019). Multilevel testing across the set of results is currently implemented using p_adj_method = "fwer"

Given that the harmonic mean p-value is calculated from the inverse p-values, these are used to provide a weighted average of expression and logFC values in the returned object. Any weights provided in w are ignored for these values as they are simple representative estimates. The representative range returned in keyval_range corresponds to the window with the lowest p-value.

The total number of windows is also returned in the final object, with the summarised values n_up and n_down indicating the number of windows with raw p-values below the calculated harmonic mean p-value, and with the corresponding direction of change.

The column containing the harmonic mean p-values is returned as 'hmp'. An additional column with adjusted hmp-values is returned with the suffix '_*' added where the p-value adjustment method is added after the underscore.

Value

A GenomicRanges object with merged ranges from the original object along with summarised or representative values from the relevant columns. The range corresponding to a representative values is also returned as described above.

Examples

```r
x <- GRanges(c("chr1:1-10", "chr1:6-15", "chr1:51-60"))
set.seed(1001)

df <- DataFrame(logFC = rnorm(3), logCPM = rnorm(3, 8), p = rexp(3, 10))
mergeByHMP(x, df, pval = "p")
mcols(x) <- df
x
mergeByHMP(x, pval = "p", p_adj_method = "fwer")
```
mergeBySig

mergeBySig (GenomicRanges object)

Description
Merge overlapping windows using p-values from significance testing

Usage

mergeBySig(x, ...)

## S4 method for signature 'GenomicRanges'
mergeBySig(
  x,
  df = NULL,
  logfc = "logFC",
  pval = "P",
  cpm = "logCPM",
  inc_cols,
  p_adj_method = "fdr",
  alpha = 0.05,
  method = c("combine", "best", "minimal"),
  merge_within = 1L,
  ignore_strand = TRUE,
  min_win = 1,
  ...)

## S4 method for signature 'RangedSummarizedExperiment'
mergeBySig(
  x,
  df = NULL,
  logfc = "logFC",
  pval = "P",
  cpm = "logCPM",
  inc_cols,
  p_adj_method = "fdr",
  alpha = 0.05,
  method = c("combine", "best", "minimal"),
  merge_within = 1L,
  ignore_strand = TRUE,
  ...)

Arguments

x GenomicRanges object
mergeBySig

... Passed to all csaw functions being wrapped

df data.frame with results of differential binding analysis performed using a sliding window strategy. If not provided, the columns in the mcols() element of x will be used

logfc, pval, cpm Column names for the values holding window specific estimates of change in binding (logfc), overall signal intensity (cpm) and the significance from statistical testing (pval)

inc_cols (Optional) Character vector of any additional columns in df to return

p_adj_method One of p.adjust.methods

alpha Significance threshold to apply during internal calculations

method Shorthand versions for which csaw strategy to use for merging windows. Choose from 'combine' (combineTests), 'best' (getBestTest) or 'minimal' (minimalTests).

merge_within Merge any non-overlapping windows within this distance

ignore_strand Passed internally to reduce and findOverlaps

min_win Only keep merged windows derived from at least this number

Details

When using sliding windows to test for differential signal, overlapping windows can be merged based on the significance of results. mergeBySig() is a wrapper to the functions combineTests, getBestTest and minimalTests, using each function’s approach to finding a representative window. The returned object differs from those returned by the original functions in that the description of windows as 'up', 'down' or mixed is omitted and the genomic range corresponding to the representative window is also returned. Column names also correspond to those in the original object.

An additional column with adjusted p-values is returned. This column retains the same name as the original but with the suffix '_*' added where the p-value adjustment method is added after the underscore.

Value

A GenomicRanges object with overlapping ranges from the original object merged and representative values returned. The range corresponding to the representative values is also returned

Examples

```r
x <- GRanges(c("chr1:1-10", "chr1:6-15", "chr1:51-60"))
set.seed(1001)
df <- DataFrame(logFC = rnorm(3), logCPM = rnorm(3,8), p = rexp(3, 10))
mcols(x) <- df
mergeBySig(x, pval = "p", method = "combine")
mergeBySig(x, pval = "p", method = "best")
mergeBySig(x, pval = "p", method = "min")
```
partitionRanges

Description

Partition a set of Genomic Ranges by another

Usage

partitionRanges(x, y, ...)

## S4 method for signature 'GRanges,GRanges'
partitionRanges(
  x,
  y,
  y_as_both = TRUE,
  ignore.strand = FALSE,
  simplify = TRUE,
  suffix = c(".x", ".y"),
  ...
)

Arguments

x, y                GenomicRanges objects
...                  Not used
y_as_both           logical(1) If there are any unstranded regions in y, should these be assigned to both strands. If TRUE unstranded regions can be used to partition stranded regions
ignore.strand       If set to TRUE, then the strand of x and y is set to "*" prior to any computation.
simplify            Pass to chopMC and simplify mcols in the output
suffix               Added to any shared column names in the provided objects

Details

The query set of ranges can be broken in regions which strictly overlap a second set of ranges. The complete set of mcols from both initial objects will included in the set of partitioned ranges

Value

A GRanges object
Examples
x <- GRanges(c("chr1:1-10", "chr1:6-15"))
x$id <- paste0("range", seq_along(x))
x
y <- GRanges(c("chr1:2-5", "chr1:6-12"))
y$id <- paste0("range", seq_along(y))
y
partitionRanges(x, y)

plotAssayDensities

Plot Densities for any assay within a SummarizedExperiment

Description
Plot Densities for any assay within a SummarizedExperiment

Usage
plotAssayDensities(x, ...)

## S4 method for signature 'SummarizedExperiment'
plotAssayDensities(
  x,
  assay = "counts",
  colour,
  linetype,
  group,
  trans = NULL,
  n_max = Inf,
  ...
)

Arguments
x A SummarizedExperiment object
...
Passed to density
assay An assay within x
colour Optional column in colData to colour lines by. To remove any colours, set this
linetype Any optional column in colData used to determine linetype
group Used by geom_line. Defaults to the sample names but setting to NULL will
trans character(1). Any transformative function to be applied to the data before cal-
n_max Maximum number of points to use when calculating densities

plotAssayHeatmap

Draw a heatmap from a single SummarizedExperiment assay

Description

Use ggplot2 to create a heatmap from a SummarizedExperiment object

Usage

plotAssayHeatmap(x, ...)

## S4 method for signature 'SummarizedExperiment'
plotAssayHeatmap(
  x,
  assay = "counts",
  by_x = "colnames",
  facet_x = NULL,
  ysideline = FALSE,
  yside_col = NULL,
  trans = NULL,
  n_max = 100,
  ...
)
Arguments

- **x**: a SummarizedExperiment object
- **...**: Not used
- **assay**: the assay to take values from
- **by_x**: the parameter to use for the x-axis. Will default to column names but should be one value per sample, such as an additional column containing shortened sample labels.
- **facet_x**: column from colData(x) which will be used to group samples along the x-axis
- **ysideline**: logical(1) Draw a line across the side of the y-axis summarising values for each range
- **yside_col**: column from colData(x) to group and colour the lines drawn on the side of the y-axis. If grouping by treatment or replicate, the mean values will be shown
- **trans**: character(1). Any transformative function to be applied to the data before calculating the density, e.g. `trans = "log2"`
- **n_max**: Maximum number of ranges to draw

Details

Draw a heatmap containing selected values from an assay within a SummarizedExperiment object. Columns within the colData element of the object can be used to facet along the x-axis (e.g. treatment groups). The maximum number of points is set to be 100, although this can be changed easily should the plot require more ranges to be drawn.

The averages across any grouping of samples can be drawn as a line plot on the side of the y-axis by setting `ysideline = TRUE`, with groups as specified in `yside_col`. This feature is added for the specific context of neighbouring or overlapping ranges, and as such may be less informative in any other scenario.

The returned object is a ggplot2 object so scales can easily be added after heatmap creation using `scale_fill_*` for the main heatmap, and `scale_colour_*` for any groupings along the y-axis.

Value

A ggplot2 object. Scales and labels can be added using conventional ggplot2 syntax.

Examples

```r
nrows <- 10; ncols <- 4
counts <- matrix(runif(nrows * ncols, 1, 1e4), nrows)
colnames(counts) <- paste0("Sample", seq_len(ncols))
df <- DataFrame(treat = c("A", "A", "B", "B"))
se <- SummarizedExperiment(
  assays = SimpleList(counts = counts),
  colData = df
)
rowRanges(se) <- GRanges(paste0("chr1:", seq_len(nrows)))
plotAssayHeatmap(se, facet_x = "treat")
```
plotAssayPCA

Plot PCA For any assay within a SummarizedExperiment

Description
Plot PCA for any assay within a SummarizedExperiment object

Usage
plotAssayPCA(x, ...)

## S4 method for signature 'SummarizedExperiment'
plotAssayPCA(
x,
assay = "counts",
colour,
shape,
size,
label,
show_points = TRUE,
pc_x = 1,
pc_y = 2,
trans = NULL,
n_max = Inf,
tol = sqrt(.Machine$double.eps),
rank = NULL,
...
)

Arguments

x An object containing an assay slot

... Passed to geom_text

assay The assay to perform PCA on

colour The column name to be used for colours

shape, size The column name(s) to be used for determining the shape or size of points

label The column name to be used for labels

show_points logical(1). Display the points. If TRUE any labels will repel. If FALSE, labels will appear at the exact points

pc_x numeric(1) The PC to plot on the x-axis

pc_y numeric(1) The PC to plot on the y-axis

trans character(1). Any transformative function to be applied to the data before performing the PCA, e.g. trans = "log2"

n_max Subsample the data to this many points before performing PCA
Any rows with variance below this value will be excluded prior to passing to `prcomp`. All rows are scaled and centred by default.

Details
Uses ggplot2 to create a PCA plot for the selected assay. Any numerical transformation prior to performing the PCA can be specified using the `trans` argument.

Value
A ggplot2 object

Examples
```r
data("se")
se$treatment <- c("E2", "E2", "E2", "E2DHT", "E2DHT", "E2DHT")
se$sample <- colnames(se)
plotAssayPCA(se, trans = "log1p", colour = "treatment", label = "sample")
plotAssayPCA(
  se, trans = "log1p", colour = "treatment", label = "sample",
  size = totals / 1e3
)
plotAssayPCA(
  se, trans = "log1p", colour = "treatment", label = "sample",
  show_points = FALSE
)
```
Arguments

x A SummarizedExperiment object

... Passed to geom_boxplot

assay The assay to plot

colour Column from colData(x) to outline the boxplots

fill Column from colData(x) to fill the boxplots

rle_group Column from colData(x) to calculate RLE within groups. Commonly an alternative sample label.

by_x Boxplots will be drawn by this grouping variable from colData(x). If not specified, the default values will be colnames(x)

n_max Maximum number of points to plot

trans character(1). Numerical transformation to apply to the data prior to RLE calculation

Details

Uses ggplot2 to create an RLE plot for the selected assay. Any numerical transformation prior to performing the RLE can be specified using the trans argument.

Value

A ggplot2 object

Examples

data("se")
se$treatment <- c("E2", "E2", "E2", "E2DHT", "E2DHT", "E2DHT")
se$sample <- colnames(se)
## A conventional RLE Plot using all samples
plotAssayRle(se, trans = "log1p", fill = "treatment")
## Calculate RLE within groups
plotAssayRle(se, trans = "log1p", fill = "treatment", rle_group = "treatment")
# Or show groups combined
plotAssayRle(se, trans = "log1p", fill = "treatment", by_x = "treatment")
**plotGrlCol**  
*Draw a plot from a GRangesList column*

---

**Description**

Draw a plot from a GRangesList column using ggplot2

**Usage**

```r
plotGrlCol(
    x,
    var = "width",
    geom = c("boxplot", "violin", "point", "jitter"),
    .id = "sample",
    df,
    fill,
    colour,
    q = 0.1,
    q_size = 3.5,
    qline_type = 2,
    qline_col = "blue",
    total = "{comma(n)}",
    total_geom = c("label", "text", "none"),
    total_pos = c("median", "top", "bottom"),
    total_size = 3.5,
    total_alpha = 1,
    total_adj = 0.025,
    ...,  
    digits = 0
)
```

**Arguments**

- **x**: A GRangesList  
  - The variable to plot. Either a column in the mcols element or width. Can be quoted or unquoted  
- **var**: Choose between different geoms, or even provide a geom_*() function  
- **.id**: The column name to place the element names. Passed internally to the same argument in bind_rows  
- **df**: Optional data.frame with columns to be passed to the colour or fill parameters. Must contain a column with the same name as the value passed to the .id argument.  
- **fill, colour**: Optional column names found in the df. Can be quoted or unquoted  
- **q**: The overall percentile to be drawn as a labelled, horizontal line. Set q = 0 to hide this line
plotGrlCol

q_size: Text size of percentile label
qline_type, qline_col: Linetype and colour arguments for the horizontal line showing the specified percentile(s)
total: Glue syntax for totals, representing the length of each GRangesList element
total_geom: Passed to annotate. Set to none to hide totals
total_pos: Position for placing totals
total_size, total_alpha: Size and transparency of totals
total_adj: Adjustment for labels
...: Passed to the geom if selecting via character string. Ignored otherwise
digits: Number of decimal places for the horizontal line label

Details

Using a common column or the width of the ranges, produces a boxplot or violinplot from each element of the provided GRangesList. The names of the GRangesList will be passed to the x-axis using the .id argument. A data frame containing annotations corresponding to each element can be supplied, ensuring that the column associated with each elements is the name passed to the .id argument.

If q is > 0, a horizontal line will be draw corresponding to this percentile across the complete dataset, with parameters for this line able to be set using the qline_* arguments. The digits argument controls how many decimal points will be shown for the associated label.

The total length of each element will be added by default as a total, and is able to be placed across the median values, or at the top and bottom extremes of the plot.

Examples

```r
## Load some peaks
data('peaks')
names(peaks) <- gsub("._peaks.+", "", names(peaks))

## The default boxplot
plotGrlCol(peaks)

## A customised violin plot
df <- data.frame(sample = names(peaks), treat = rep(c("A", "B"), each = 3))
plotGrlCol(
    peaks, geom = "violin", total_pos = "bottom", total_adj = 0.05,
    df = df, fill = "treat",
    draw_quantiles = 0.5, trim = FALSE, width = 0.7, alpha = 0.7
) +
scale_y_log10()
plotGrlCol(
    peaks, var = score, geom = "jitter", total_pos = "bottom", total_adj = 0.05,
    df = df, colour = treat, width = 0.2, height = 0
)
```
plotHFGC

Plot a Genomic Region showing HiC, Features, Genes and Coverage

Description

Plot a region with showing HiC, Features, Genes and Coverage

Usage

plotHFGC(gr, hic, features, genes, coverage, annotation, zoom = 1, shift = 0, max = 1e+07, axistrack = TRUE, cytobands, covtype = c("l", "heatmap"), linecol = c(), gradient = hcl.colors(101, "viridis"), hiccol = list(anchors = "lightblue", interactions = "red"), featcol, genecol, annotcol, highlight = "blue", hicsize = 1, featsize = 1, genesize = 1, covsize = 4, annotsize = 0.5, hicname = "HiC", featname = "Features", featstack = c("full", "hide", "dense", "squish", "pack"), collapseTranscripts = "auto", maxTrans = 12)
Arguments

gr

The range(s) of interest. Must be on a single chromosome

hic

Any HiC interactions to be included as a GenomicInteractions object. If not supplied, no HiC track will be drawn.

features

A named GRangesList or list of GRangesList objects. Each GRangesList should contain features in each element which will drawn on the same track. If providing a list, each GRangesList within the list will drawn on a separate track. If this argument is not specified, no feature track will be drawn. Features will be drawn with colours provided in featcol.

genes

A GRanges object with exon structure for each transcript/gene. If not included, no track will be drawn for gene/transcript structure. The expected mcols in this object are type, gene, exon transcript and symbol. See data(ex_trans) for an example.

coverage

A named list of BigWigFileList objects containing the primary tracks to show coverage for. Each list element will be drawn on a separate track, with elements within each BigWigFileList shown on the same track. List names will become track names. Alternatively, a single BigWigFileList will plot all individual files on separate tracks. If not included, no coverage tracks will be drawn.

annotation

Annotations for the coverage track(s). A single GRangesList if coverage is a BigWigListList. If coverage is supplied as a list of BigWigFileLists, a named list of GRangesList objects for each coverage track being annotatated. Names must match those given for coverage.

zoom

Multiplicative factor for zooming in and out

shift

Shift the plot. Applied after zooming

max

The maximum width of the plotting region. Given that the width of the final plotting window will be determined by any HiC interactions, this argument excludes any interactions beyond this distance. Plotting can be somewhat slow if any long range interactions are included. Ignored if no HiC interactions are supplied.

axistrack

logical. Add an AxisTrack()

cytobands

Cytogenetic bands to be displayed on each chromosome. See data('grch37.cytobands') for the correct format. Only drawn if a cytobands data.frame is provided.

covtype

The plot type for coverage. Currently only lines ("l") and heatmaps ("heatmap") are supported
linecol  If passing a BigWigFileList to coverage, a vector of colours. If passing a list of BigWigFileList objects to coverage, a list of colours with structure that matches the object being passed to coverage, i.e. a named list of the same length, with elements who’s length matches each BigWigFileList. Only used if covtype = "l".

gradient Colour gradient for heatmaps

hiccol  list with names "anchors" and "interactions". Colours are passed to these elements

featcol  Named vector (or list) of colours for each feature. Must be provided if drawing features

genecol  Named vector (or list) of colours for each gene category

annotcol  Colours matching the coverage annotations

highlight Outline colour for the highlight track. Setting this to NULL will remove the highlight

hicsize, featsize, genesize, covsize, annotsize  Relative sizes for each track (hic, features, genes, coverage & annotation)

hicname, featname  Names displayed in the LHS panel

featstack  Stacking for the feature track

collapseTranscripts  Passed to GeneRegionTrack for the genes track. Defaults to "auto" for automatic setting. If the number of transcripts to be plotted is > maxtrans, the argument will be automatically set to "meta", otherwise this will be passed as FALSE which will show all transcripts.

maxTrans  Only used if collapseTranscripts is set to "auto".

ylim  If a numeric vector, this will be passed to all coverage tracks. Alternatively, a named list of y-limits for each coverage track with names that match those in each element of the coverage list.

...  Passed to DataTrack for the coverage tracks only. Useful arguments may be things like legend

fontsize  Applied across all tracks

cex.title  Passed to all tracks

rotation.title  Passed to all tracks

col.title  Passed to all tracks

background.title  Passed to all tracks

title.width  Expansion factor passed to plotTracks, and used to widen the panels on the LHS of all tracks. Can have unpredictable effects on the font size of y-axis limits due to the algorithm applied by plotTracks
Details

Convenience function for plotting a common set of tracks. All tracks are optional. For more fine control, users are advised to simply use Gviz directly.

The primary tracks defined in this function are H (HiC), F (features), G (genes), and C (coverage). Axis and Ideogram tracks are an additional part of this visualisation, with the Ideogram also being optional.

Use all tracks specific to this dataset to generate a simple visualisation. In descending order the tracks displayed will be:

1. HiC Interactions (if supplied)
2. Regulatory features
3. Genes/genes
4. Coverage tracks as supplied

All tracks are optional and will simply be omitted if no data is supplied. See individual sections below for a more detailed explanation of each track.

If wanting a single track of genes, simply pass a GRanges object in the format specified for a GeneRegionTrack. Passing a GRangesList with the same format will yield an individual track for each list element, with each track shown by default as a separate colour. This can be used for showing Up/Down-regulated genes, or Detected/Undetected genes.

If passing a BigWigFileList for the coverage track, each file within the object will be drawn on a separate track. If specified, the same y-limits will be applied to each track. If passing a list of BigWigFileList objects, each list element will be drawn as a single track with the individual files within each BigWigFileList overlaid within each track.

Cytogenetic band information must be in the structure required by IdeogramTrack, with data for both GRCh37 and GRCh38 provided in this package (grch37.cytobands, grch38.cytobands).

A highlight overlay over the GRanges provided as the gr argument will be added if a colour is provided. If set to NULL, no highlight will be added.

Value

A Gviz object

Displaying HiC Interactions

The available arguments for displaying HiC Interactions are defined below. If hic is supplied, a single InteractionTrack will be added displaying all interactions with an anchor within the range specified by gr. Only interactions with an anchor explicitly overlapping gr will be shown. If no interactions are found within gr, the track will not be displayed. The plotting range will expand to incorporate these interactions, with the parameter max providing an upper limit on the displayed range.

hic This is the GInteractions object required for inclusion of a HiC track in the final output. Will be ignored if not supplied

hiccol Determines the colours used for display of anchors and interactions

hicsize Relative size of the track compared to others
hicname  The name to display on the LHS panel

max  The maximum width of the plotted region. If multiple long-range interactions are identified, this provides an upper limit for the display. This defaults to 10Mb.

Displaying Features

If wanting to add an AnnotationTrack with regions defined as ‘features’, the following arguments are highly relevant. All are ignored if features is not provided.

features  A named GRangesList. Each element will be considered as a separate feature and drawn as a block in a distinct colour. Any mcols data will be ignored.

featcol  A named vector (or list) providing a colour for each element of features

featname  The name to display on the LHS panel

featstack  Stacking to be applied to all supplied features

featsize  Relative size of the track compared to others

Displaying Genes And Transcripts

To display genes or transcripts, simply provide a single GRanges object if you wish to display all genes on a single track. The mcols element of this object should contain the columns feature, gene, exon, transcript and symbol as seen on the GeneRegionTrack help page.

Alternatively, a GRangesList can be provided to display genes on separate tracks based on their category. This can be useful for separating and colouring Up/Down regulated genes in a precise way. All elements should be as described above. Again, all parameters associated with this track-set will be ignored of no object is supplied to this argument.

genes  A GRanges or GRangesList object as described above

genecol  A single colour if supplying a GRanges object, or a named vector/list of colours matching the GRangesList

geneseize  Relative size of the track compared to others

collapseTranscripts  Passed to all tracks. See the GeneRegionTrack section in settings for detail regarding possible arguments. If genes is a GRangesList, can be a named vector/list with names matching the names of the genes object.

Displaying Coverage Tracks

This section contains the most flexibility and can take two types of input. The first option is a BigWigFileList, which will lead to each BigWig file being plotted on it's own track. An alternative is a list of BigWigFileList objects. In this case, each list element will be plotted as a separate track, with all individual BigWig files within each list element overlaid within the relevant track.

In addition to the coverage tracks, annotations can be added to each BigWigFileList in the form of coloured ranges, indicating anything of the users choice. Common usage may be to indicate regions with binding of a ChIP target is found to be detected, unchanged, gained or lost.

coverage  A BigWigFileList or list of BigWigFileList objects. A single BigWigFileList will be displayed with each individual file on a separate track with independent y-axes. Each element of the BigWigFileList must be named and these names will be displayed on the LHS
panels A list of BigWigFileList objects will be displayed with each list element as a separate track, with any BigWig files overlaid using the same y-axis. The list must be named with these names displayed on the LHS panel. Each internal BigWig within a BigWigFileList must also be named.

covtype Currently only lines (covtype = "l") and heatmaps (covtype = "heatmap") are supported. Colours can be specified using the arguments below

linecol Can be a single colour applied to all tracks, or a named vector (or list) of colours. If coverage is a single BigWigFileList, these names should match the names of this object exactly. If coverage is a list of BigWigFileList objects, linecol should be a list with matching names. Each element of this list should also be a named vector with names that exactly match those of each corresponding BigWigFileList.

gradient A colour gradient applied to all heatmap tracks. No specific structure is required beyond a vector of colours.

covsize Relative size of the tracks compared to others

ylim Can be a vector of length 2 applied to all coverage tracks. Alternatively, if passing a list of BigWigFileList objects to the coverage argument, this can be a named list of numeric vectors with names matching coverage

annotation Each BigWigFileList needs annotations to be passed to this argument as a named GRangesList, with names being used to associate unique colours with that set of ranges. If coverage is a BigWigFileList a simple GRangesList would be supplied and a single 'annotation' track will appear at the top of the set of coverage tracks. If coverage is a list, then a named list of GRangesList objects should be supplied, with each being displayed above the corresponding track from the coverage object.

annotcol A vector of colours corresponding to all names within all GRangesList elements supplied as annotation. It is assumed that the same colour scheme will be applied to all annotation tracks and, as such, the colours should not be provided as a list which matches the coverage tracks. Instead, every named element anywhere in the annotation GRanges, across all of the tracks must be included as a colour

annotsize Relative size of the tracks compared to others

Examples

library(rtracklayer)
## Make sure we have the cytobands active
data(grch37.cytobands)

## Prepare the HiC, promoter & transcript information
data(ex_hic, ex_trans, ex_prom)
ex_features <- GRangesList(Promoter = ex_prom)
featcol <- c(Promoter = "red")

## Prepare the coverage
fl <- system.file("extdata\bigwig", c("ex1.bw", "ex2.bw"), package = "extraChIPs")
 bwfl <- BigWigFileList(fl)
names(bwfl) <- c("ex1", "ex2")
bw_col <- c(ex1 = "#4B0055", ex2 = "#007094")
## Define the plotting range
gr <- GRanges("chr10:103862000-103900000")

## Now create the basic plot
plotHFGC(
gr, hic = ex_hic, features = ex_features, genes = ex_trans, coverage = bwfl, featcol = featcol, linecol = bw_col, cytobands = grch37.cytobands)

plotHFGC(
gr, hic = ex_hic, features = ex_features, genes = ex_trans, coverage = bwfl, featcol = featcol, linecol = bw_col, cytobands = grch37.cytobands, maxTrans = 1)

---

### plotOverlaps

**Plot Overlaps Between List Elements**

#### Description

Plot Overlaps between list elements as an upset or Venn diagram

#### Usage

plotOverlaps(x, ...)

#### S4 method for signature 'GRangesList'

plotOverlaps(
x, type = c("auto", "venn", "upset"),
var = NULL,
f = c("mean", "median", "max", "min", "sd"),
set_col = NULL,
...,
.sort_sets = "ascending",
hj_sets = 1.15,
sz_sets = 3.5,
exp_sets = 0.25,
min-gapwidth = 1L,
ignore.strand = TRUE
)

#### S4 method for signature 'list'

plotOverlaps(
  x,
  type = c("auto", "venn", "upset"),
  set_col = NULL,
  ..., 
  .sort_sets = "ascending",
  hj_sets = 1.15,
  sz_sets = 3.5,
  exp_sets = 0.25
)

Arguments

x  GRangesList of S3 list able to be coerced to character vectors
...
Passed to draw.pairwise.venn (or draw.single/triple.venn) for Venn Diagrams, and to upset for UpSet plots
type  The type of plot to be produced
var  Column to summarised as a boxplot in an upper panel (UpSet plot only)
f  Summarisation function. Must return a single value from any numeric vector
set_col  Colours to be assigned to each set
.sort_sets  passed to sort_sets in upset
hj_sets  Horizontal adjustment of set size labels
sz_sets  Text size for set size labels. Passed internally to geom_text(size = sz_sets)
exp_sets  X-axis expansion for set size panel
min.gapwidth, ignore.strand  Passed to reduce

Details

This function should give the capability to show overlaps for any number of replicates or groups, or a list of items such as gene names. For n = 2, a scaled Venn Diagram will be produced, however no scaling is implemented for n = 3

UpSet plots are possible for any lists with length > 1, and are the only implemented possibility for lists > 3.

If the input is a GRangesList an additional boxplot can be requested using any numeric column within the existing mcols() element. Values will be summarised across all elements using the requested function and the boxplot will be included as an upper panel above the intersections

Value

Either a VennDiagram (i.e. grid) object, or a ComplexUpset plot
Examples

```r
## Examples using a list of character vectors
ex <- list(
    x = letters[1:5],
    y = letters[c(6:15, 26)],
    z = letters[c(2, 10:25)]
)
plotOverlaps(ex, type = "upset")
plotOverlaps(ex, type = "venn", set_col = 1:3, alpha = 0.3)
plotOverlaps(ex, type = "upset", set_col = 1:3, labeller = stringr::str_to_title)
plotOverlaps(ex[1:2])

## GRangesList object will produce a boxplot of summarised values in the upper panel
data("peaks")
grl <- peaks[1:3]
names(grl) <- gsub("_peaks.+", "", names(grl))
plotOverlaps(grl, type = "upset", var = "score", f = "max")

## If only two samples are present, a VennDiagram will be produced
plotOverlaps(grl[1:2], set_col = c("green", "blue"))
```

---

**plotPairwise**

*Plot Pairwise Values from a GRangesList*

**Description**

Plot Pairwise Values from a GRangesList by overlapping GRanges

**Usage**

```r
plotPairwise(
    x,
    var,
    colour,
    label,
    index = c(1, 2),
    p = 0,
    method = "union",
    ignore.strand = TRUE,
    min_width = 0,
    xside = c("boxplot", "density", "violin", "none"),
    yside = c("boxplot", "density", "violin", "none"),
    side_panel_width = c(0.3, 0.4),
    xside_axis_pos = "right",
    yside_axis_label = scales::label_wrap(10),
    smooth = TRUE,
    rho_geom = c("text", "label", "none"),
    rho_col = "black",
)```
Arguments

x A GRangesList

var The column to compare between list elements

colour Optional column to use for combining across elements and setting point colour

label Optional column to use for labelling ranges with the most extreme changes

index Which list elements to compare

p, method, ignore.strand, min_width
Passed to makeConsensus()

xside, yside Will call geom_(x/y)side* from the package ggside and show additional panels on the top and right of the plot respectively

side_panel_width Set the relative widths of the side panels

xside_axis_pos Position for axis_labels in the top panel when using a discrete axis

yside_axis_label Wrapping for axis labels on the right-side panel when using a discrete axis. Set to waiver() to turn off wrapping

smooth logical(1). If TRUE a regression line will be drawn using geom_smooth. To add this manually, set to FALSE and call this geom with any custom parameters after creating the plot

rho.geom Used to add correlation coefficients for the two values

rho_col, rho_size, rho_alpha Parameters for displaying the correlation

rho_pos Place the correlation coefficient within the plotting region

label.geom Used to add labels from the column specified in label

label_width Label text will be truncated to this length

label.sep If multiple values (e.g. genes) are mapped to a range, separate values using this string
label_size, label_alpha
Passed to the geom used for adding labels

min_d
Labels will only be added if the points lie circle beyond a circle of this radius

group_sep
Text separator used to separate categories when specifying colour

simplify_equal
logical(1) When combining columns from both elements for the colour categories, should shared values be annotated as 'Both ...' instead of having longer, more difficult to read annotations.

name_sep
Character string to separate names of the GRangesList and the selected column. Will appear as axis-labels

plot_theme
Sets the initial theme by using the default theme for the current R session via get_theme()

... Passed to geom_point() for the main panel

Details
This function enables pairwise plotting of two elements within a GRangesList. All elements of the GRangesList will contain the same columns, so a set of consensus ranges are first formed, before then taking all values from each GRangesList element which overlap the range and producing a pairwise plot.

Given that not all ranges will have values in both elements, side panels are produced which can show the distribution of plotted values, along with those which are only found in one of the foundational GRanges. These can take the form of density, violin or boxplots.

Addition columns, such as Differential Signal status can also be used to form pairwise groups and colour the points.

If a column in the GRangesList is suitable for labelling points, such as a column with genes mapped to each range, this can be specified using the argument label = "col_to_label". Only the furthest point from the origin will be labelled within each group used to colour the points. Labels will only be added if they lie beyond a circle of radius min_d from the origin. If multiple genes are mapped to the range, these will be separated by the string provided in the label_sep argument.

A regression line and correlation co-efficient are added to the plot by default, but can be hidden easily if preferred

Value
A ggside or ggplot2 object

Examples

```r
theme_set(theme_bw())
set.seed(100)
gr1 <- GRanges(paste0("chr1:", seq(10, 150, by = 10)))
width(gr1) <- 5
gr1$logFC <- rnorm(length(gr1))
gr1$FDR <- rbeta(length(gr1), 1, 8)
gr2 <- GRanges(paste0("chr1:", seq(51, 250, by = 15)))
width(gr2) <- 4
```
gr2$logFC <- rnorm(length(gr2))
gr2$FDR <- rbeta(length(gr2), 1, 8)
gr1 <- GRangesList(TF1 = gr1, TF2 = gr2)
gr1 <- addDiffStatus(gr1)

# Using the defaults
plotPairwise(gr1, var = "logFC")

# Density plots on the side panels
plotPairwise(gr1, var = "logFC", xside = "density", yside = "density")
+ scale_fill_viridis_d(alpha = 0.7)

# Turning off side panels, regression line and correlations
plotPairwise(gr1, var = "logFC", xside = "none", yside = "none",
             rho_geom = "none", smooth = FALSE)

# Add colours using the status column
plotPairwise(gr1, var = "logFC", colour = "status")
+ scale_fill_manual(values = rep_len(c("blue", "red", "white", "grey"), 8))
+ guides(fill = "none")

---

**plotPie**

*Draw Pie Graphs based on one or more columns*

**Description**

Draw Pie Graphs based on one or more data.frame columns

**Usage**

plotPie(object, ...)

## S4 method for signature 'GRanges'
plotPie(object, scale_by = c("n", "width"), ...)

## S4 method for signature 'DataFrame'
plotPie(object, ...)

## S4 method for signature 'data.frame'
plotPie(
    object,
    fill,
    x,
    y,
scale_by,
scale_factor = 1000,
width = 0.8,
total_geom = c("label", "text", "none"),
total_glue = "{comma(N})",
total_colour = "black",
total_fill = "white",
total_alpha = 1,
total_size = 3,
min_p = 0.01,
max_p = 1,
cat_geom = c("label", "text", "none"),
cat_glue = ".data[[fill]]\n{comma(n, 1)}\n{percent(p, 0.1)}",
cat_colour = "black",
cat_fill = "white",
cat_size = 3,
cat_alpha = 1,
cat_adj = 0,
hole_width = 0,
...
)

Arguments

object An object (data.frame)
... Not used
scale_by Scale the counts by this column. In this case of a GRanges object this defaults
to the count (scale_by = "n") but can also be specified as being width of each
range (scale_by = "width"). If choosing width, width will be displayed in Kb
fill The category/column used to fill the slices of the pie charts
x The second (optional) category/column to place along the x-axis
y The final (optional) category/column to place along the y-axis
scale_factor When scaling by another column, such as width, totals will be divided by this
value, with 1000 being the default to provide output in kb.
width Scale the width of all pies
total_geom The geom_* to use for the totals at the centre of each pie. Setting this to 'none'
will disable totals
total_glue glue syntax to use for the totals in the centre of each pie. The column 'N' will
produce the totals and any other values or formatting may be added here.
total_colour, total_fill, total_alpha, total_size
Colour, fill, alpha and size for the main totals in the centre of each pie chart
min_p The minimum proportion of the total required for adding labels. Effectively
removes labels from pie charts with few members. Alternatively when only one
column is specified, categories below this will not be shown around the edge of
the plot
max_p only display labels for segments representing less than this proportion of the total.
cat_geom The geom_* to use for category labels corresponding to each slice of the pie. Setting this to 'none' will disable category labels
cat_glue glue syntax to use for the category labels corresponding to each slice of the pie charts. The columns ‘n’ and ‘p’ can be used to print totals and proportions for each slice.
cat_colour, cat_fill, cat_size, cat_alpha Colour, fill, size and alpha for category labels
cat_adj Adjust category labels
hole_width Add a hole in the middle to turn the plot into a donut. Values between zero and 1 work best. Only implemented for pie charts using one value (i.e. fill)

Details
Using a data.frame as input, this function will draw pie graphs based on one or more columns, by simply counting the values in combination across these columns. One column must be selected for the fill as a bare minimum, with up to three being possible. Additional columns can be set for the x-axis to draw a series of pie-graphs in a row, with a further column able to be added to layout a series of pie graphs in a grid.

If only one column/category is chosen, category labels will be added around the edge of the plot.
If show_total = TRUE the overall counts for each pie graph will be added in the centre using geom_label. Parameters for these labels are customisable.

Value
A ggplot2 object able to be customised with colour scales and themes. Also note that the $data element of the returned object will contain the data.frame used for plotting. The additional column label_radians represents the mid-point of each pie slice and can be used for manually adding labels to each pie. Only applies when plotting across the x or y axes.

Examples
```r
set.seed(200)
df <- data.frame(
  feature = sample(c("Promoter", "Enhancer", "Intergenic"), 200, replace = TRUE),
  TF1 = sample(c("Up", "Down", "Unchanged"), 200, replace = TRUE),
  TF2 = sample(c("Up", "Down", "Unchanged"), 200, replace = TRUE),
  w = rexp(200)
)
plotPie(df, fill = "feature", total_glue = "N = \{comma(N)\}"
plotPie(df, fill = "feature", scale_by = "w", total_geom = "none",
cat_glue = "\{percent(p)\}", cat_size = 5
) plotPie(df, fill = "feature", x = "TF1"
)```
plotPie(
  df, fill = "feature", x = "TF1", y = "TF2", min_p = 0.02,
  total_geom = "none", cat_glue = "{} / {}"
) +
  scale_fill_viridis_d() +
  theme_bw()

## And using a GRanges object
data("ex_prom")
gr <- ex_prom
mcols(gr) <- df[seq_along(gr),]
## Show values by counts
plotPie(gr, fill = "feature", total_size = 5)
## Show values scaled by width of each range as a donut plot
plotPie(
gr, fill = "feature", scale_by = "width", total_glue = "(round(N, 1))kb",
cat_glue = "(percent(p, 0.1))", cat_size = 4, total_size = 5, hole_width = 0.2
)

---

plotProfileHeatmap  
*Draw a coverage Profile Heatmap*

**Description**

Plot a coverage Profile Heatmap across multiple ranges

**Usage**

plotProfileHeatmap(object, ...)

## S4 method for signature 'GenomicRangesList'
plotProfileHeatmap(
  object,
  profileCol = "profile_data",
  xValue = "bp",
  fillValue = "score",
  facetX = NULL,
  facetY = NULL,
  colour = facetY,
  linetype = NULL,
  summariseBy = c("mean", "median", "min", "max", "none"),
  xLab = xValue,
  yLab = NULL,
  fillLab = fillValue,
  relHeight = 0.3,
  sortFilter = NULL,
  maxDist = 100,

plotProfileHeatmap

... }

## S4 method for signature 'GenomicRanges'
plotProfileHeatmap(
  object,
  profileCol = "profile_data",
  xValue = "bp",
  fillValue = "score",
  facetX = NULL,
  facetY = NULL,
  colour = facetY,
  linetype = NULL,
  summariseBy = c("mean", "median", "min", "max", "none"),
  xLab = xValue,
  yLab = NULL,
  fillLab = fillValue,
  relHeight = 0.3,
  summaryLabelSide = "left",
  respectLevels = FALSE,
  sortFilter = NULL,
  maxDist = 100,
  ...
)

Arguments

object A GRanges or GRangesList object

... Passed to facet_grid internally. Can be utilised for switching panel strips or passing a labeller function

profileCol Column name specifying where to find the profile DataFrames

xValue, fillValue Columns within the profile DataFrames for heatmaps

facetX, facetY Columns used for faceting across the x- or y-axis respectively

colour Column used for colouring lines in the summary panel. Defaults to any column used for facetY

linetype Column used for linetypes in the summary panel

summariseBy Function for creating the summary plot in the top panel. If set to 'none', no summary plot will be drawn. Otherwise the top panel will contain a line-plot representing this summary value for each x-axis bin

xLab, yLab, fillLab Labels for plotting aesthetics. Can be overwritten using labs() on any returned object

relHeight The relative height of the top summary panel. Represents the fraction of the plotting area taken up by the summary panel.
sortFilter

If calling on a GRangesList, a method for subsetting the original object (e.g., 1:2). If calling on a GRanges object should be and expression able to be parsed as a filtering expression using eval_tidy. This is applied when sorting the range order down the heatmap such that ranges can be sorted by one or specific samples, or all. Ranges will always be sorted such that those with the strongest signal are at the top of the plot.

maxDist

Maximum distance from the centre to find the strongest signal when arranging the ranges.

summaryLabelSide

Side to place y-axis for the summary plot in the top panel.

respectLevels

logical(1) If FALSE, facets along the y-axis will be arranged in descending order of signal, otherwise any original factor levels will be retained.

Details

Convenience function for plotting coverage heatmaps across a common set of ranges, shared between one or more samples. These are most commonly the coverage values from merged samples within a treatment group. The input data structure is based on that obtained from getProfileData, and can be provided either as a GRanges object (generally for one sample) or as a GRangesList.

A ‘profile DataFrame’ here refers to a data.frame (or tibble, or DataFrame) with a coverage value in one column that corresponds to a genomic bin of a fixed size denoted in another, as generated by getProfileData. Given that multiple ranges are most likely to be drawn, each profile data frame must be the same size in terms of the number of bins, each of which represent a fixed number of nucleotides. At a minimum this is a two column data frame although getProfileData will provide three columns for each specified genomic region.

If using a GRangesList, each list element will be drawn as a separate panel by default. These panels will appear in the same order as the list elements of the GRangesList, although this can easily be overwritten by passing a column name to the facetX argument. The default approach will add the original element names as the column "name" which can be seen in the $data element of any resultant ggplot object produced by this function.

Value

A ggplot2 object, able to be customised using standard ggplot2 syntax.

Examples

```r
library(rtracklayer)
fl <- system.file(
  "extdata", "bigwig", c("ex1.bw", "ex2.bw"), package = "extraChIPs"
)
bwf1 <- BigWigFileList(fl)
names(bwf1) <- c("ex1", "ex2")
gr <- GRanges(
  c(
    "chr10:103880281-103880460", "chr10:103892581-103892760",
    "chr10:103877281-103877460"
  )
)
```

plotSplitDonut <- function(object, ...) {
  ## S4 method for signature 'GRanges'
  plotSplitDonut(object, scale_by = c("n", "width"), ...) 

  ## S4 method for signature 'DataFrame'
  plotSplitDonut(object, ...) 

  ## S4 method for signature 'data.frame'
  plotSplitDonut(
    object,
    inner,
    outer,
    scale_by,
    scale_factor = 1000,
    r_centre = 0.5,
    r_inner = 1,
    r_outer = 1,
    total_glue = "{comma(N)}",
    total_size = 5,
    total_colour = "black",
    inner_glue = "{inner} {.data[[inner]]}\n{percent(p,0.1)}",
    outer_glue = "{outer} {.data[[outer]]}\n{percent(p,0.1)}",
    total_label = c("label", "text", "none"),
    inner_label = c("label", "text", "none"),
    outer_label = c("label", "text", "none"),
    label_alpha = 1,
    inner_label_alpha = NULL,
    outer_label_alpha = NULL,
    label_size = 3,
    inner_label_size = NULL,
  )
}

pd <- getProfileData(bwfl, gr)
plotProfileHeatmap(pd, "profile_data") +
  scale_fill_viridis_c(option = "inferno", direction = -1) +
  labs(fill = "Coverage")

---

**plotSplitDonut**  
*Draw Two-Level Donut Charts*

**Description**

Create Donut charts based on one or two columns in a data frame

**Usage**

```r
plotSplitDonut(object, ...)
```

## S4 method for signature 'GRanges'
```r
plotSplitDonut(object, scale_by = c("n", "width"), ...) 
```

## S4 method for signature 'DataFrame'
```r
plotSplitDonut(object, ...) 
```

## S4 method for signature 'data.frame'
```r
plotSplitDonut(
  object,
  inner,
  outer,
  scale_by,
  scale_factor = 1000,
  r_centre = 0.5,
  r_inner = 1,
  r_outer = 1,
  total_glue = "{comma(N)}",
  total_size = 5,
  total_colour = "black",
  inner_glue = "{inner} {.data[[inner]]}\n{percent(p,0.1)}",
  outer_glue = "{outer} {.data[[outer]]}\n{percent(p,0.1)}",
  total_label = c("label", "text", "none"),
  inner_label = c("label", "text", "none"),
  outer_label = c("label", "text", "none"),
  label_alpha = 1,
  inner_label_alpha = NULL,
  outer_label_alpha = NULL,
  label_size = 3,
  inner_label_size = NULL,
)```

---
outer_label_size = NULL,
label_colour = "black",
inner_label_colour = NULL,
outer_label_colour = NULL,
min_p = 0.05,
inner_min_p = NULL,
outer_min_p = NULL,
max_p = 1,
in1er_max_p = NULL,
outer_max_p = NULL,
inner_pattern = ".",
outer_pattern = ".",
inner_rotate = FALSE,
outer_rotate = FALSE,
explode_inner = NULL,
explode_outer = NULL,
explode_query = c("AND", "OR"),
explode_x = 0,
explode_y = 0,
explode_r = 0,
nudge_r = 0.5,
inner_nudge_r = NULL,
outer_nudge_r = NULL,
expand = 0.1,
inner_pallete = NULL,
outer_pallete = NULL,
inner_legend = TRUE,
outer_legend = TRUE,
outer_p_by = c("all", "inner"),
layout = c(main = area(1, 1, 12, 12), lg1 = area(2, 12), lg2 = area(11, 12)),
...
)

**Arguments**

- **object** A GRanges or data.frame-like object
- **scale_by** Column to scale values by. If provided, values in this column will be summed, instead of simply counting entries. Any label in the centre of the plot will also reflect this difference
- **inner** Column name to create the inner ring
- **outer** Column name to create the outer ring, subset by the inner ring
- **scale_factor** When scaling by another column, such as width, totals will be divided by this value, with 1000 being the default to provide output in kb.
- **r_centre** The radius of the hole in the centre. Setting to zero will create a Pie chart
- **r_inner, router** The radii of the inner/outer rings
total_glue: **glue**-syntax for formatting the total which appears in the centre of the plot. Internally, the value N will be calculated and as such, this value should appear within this argument.

**total_size**: Label size total number of entries in the centre of the plot.

**total_colour**: Label colour for the summary total in the centre.

inner_glue, outer_glue: **glue**-syntax for formatting labels which appear on each inner/outer segment. Internally, the values n and p will be calculated as totals and proportions of the total. As such, these values can appear within this argument, as well as the fields described in the details.

**total_label, inner_label, outer_label**: Can take values 'text', 'label' or 'none'. If setting one the first two values, the labelling function **geom_** will be called, otherwise no label will be drawn.

**label_alpha, inner_label_alpha, outer_label_alpha**: Transparency for labels.

**label_size, inner_label_size, outer_label_size**: Size of all text labels.

**label_colour, inner_label_colour, outer_label_colour**: Takes any colour specification, with the additional option of 'palette'. In this special case, the same palette as is used for each segment will be applied.

**min_p, inner_min_p, outer_min_p**: Only display labels for segments representing greater than this proportion of the total. If inner/outer values are specified, the values in min_p will be ignored for that layer.

**max_p, inner_max_p, outer_max_p**: Only display labels for segments representing less than this proportion of the total. If inner/outer values are specified, the values in max_p will be ignored for that layer.

**inner_pattern, outer_pattern**: Regular expressions which are combined with max_p and min_p values for accurately choosing labels.

**inner_rotate, outer_rotate**: Logical(1). Rotate labels for inner or outer rings. This will be ignored by when setting the geom as "label". See geom_text.

**explode_inner, explode_outer**: Regular expressions from either the inner or outer ring for which segments will be 'exploded'.

**explode_query**: Setting to AND and specifying values for both the inner and outer ring will require matches in both categories.

**explode_x, explode_y**: Numeric values for shifting exploded values.

**explode_r**: Radius expansion for exploded values.

**nudge_r, inner_nudge_r, outer_nudge_r**: Radius expansion for labels.
**plotSplitDonut**

expand Passed to **expansion** for both x and y axes. Can be helpful if labels are clipped by plot limits

inner_palette Colour palette for the inner ring

outer_palette Optional colour palette for the outer ring

inner_legend, outer_legend logical(1). Show legends for either layer

outer_p_by Scale the proportions for outer segments by the complete dataset, or within each inner segment

layout Passed to **plot_layout**

---

**Details**

Using a data.frame or GRanges object, this function enables creation of a Pie/Donut chart with an inner and outer ring. The function itself is extremely flexible allowing for separate colour palettes in the inner and outer rings, as well as highly customisable labels.

Sections can be exploded using a value from the inner ring or outer ring separately, or in combination by setting `explode_query = "AND"`. Exploded sections can be shifted by expanding the radius (`explode_r`), or along the x/y co-ordinates using `explode_x/y`, allowing for detailed placement of sections.

If only the inner palette is specified, segments in the outer ring will be assigned the same colours as the inner segments, but with increased transparency. Only a single legend will be drawn in this scenario. If an outer palette is specified, both colour palettes are completely distinct and two distinct legends will be drawn. The placement of these legends, along with the larger donut plot, can be manually specified by providing a layout as defined in **plot_layout**. Names are not required on this layout, but may be beneficial for code reproducibility.

The inner label denoting the total can also be heavily customised using the **glue** syntax to present the calculated value N along with any additional text, such as ‘kb’ if scaling GenomicRanges by width. The same approach can be taken for the inner and outer labels, where totals are held in the value `n`, proportions are held in the value `p` and the values corresponding to each segment can be accessed using `.data[[inner]]` or `.data[[outer]]`. Column titles can be added using `{inner}`/{outer}. Values from the inner segments can be added to the outer labels using this strategy enabling a wide variety of labelling approaches to be utilised.

**Value**

A patchwork object consisting of both ggplot2 objects and legend grobs

**Examples**

```r
set.seed(200)
df <- data.frame(
  feature = sample(c("Promoter", "Enhancer", "Intergenic"), 200, replace = TRUE),
  TF1 = sample(c("Up", "Down", "Unchanged"), 200, replace = TRUE),
  TF2 = sample(c("Up", "Down", "Unchanged"), 200, replace = TRUE)
)
## The standard plot
```
propOverlap

Find the proportions of an overlapping range

Description

Find the proportion of a query range which overlaps the subject

Usage

propOverlap(x, y, ...)

## S4 method for signature 'GRanges,GRanges'
propOverlap(x, y, ignore.strand = FALSE, ...)

Arguments

x, y           A GenomicRanges object
...            Not used
ignore.strand  If set to TRUE, then the strand of x and y is set to "*" prior to any computation.

Details

This behaves similarly to overlapsAny except the proportion of the query range which overlaps one or more subject ranges is returned instead of a logical vector

Value

Numeric vector the same length as x

Examples

x <- GRanges("chr1:1-10")
y <- GRanges("chr1:1-5")
propOverlap(x, y)
propOverlap(y, x)
reduceMC

Reduce ranges retaining mcols

Description

Reduce ranges retaining mcols

Usage

reduceMC(x, ignore.strand = FALSE, simplify = TRUE, ...)

Arguments

x
A GenomicRanges object

ignore.strand
If set to TRUE, then the strand of x and y is set to "*" prior to any computation.

simplify
logical(1). Attempt to simplify returned columns where possible

...
Passed to reduce

Details

This function extends reduce so that all mcols are returned in the output. Where the reduced ranges
map to multiple ranges in the original range, mcols will be returned as CompressedList columns.

If simplify = TRUE columns will be returned as vectors where possible.

Value

A GRanges object

Examples

x <- GRanges(c("chr1:1-10:+", "chr1:6-12:-"))
x$id <- c("range1", "range2")
reduceMC(x)
reduceMC(x, ignore.strand = TRUE)
Description

Perform set operations retaining all mcols from the query range

Usage

setdiffMC(x, y, ...)

intersectMC(x, y, ...)

unionMC(x, y, ...)

## S4 method for signature 'GRanges,GRanges'
setdiffMC(x, y, ignore.strand = FALSE, simplify = TRUE, ...)

## S4 method for signature 'GRanges,GRanges'
intersectMC(x, y, ignore.strand = FALSE, simplify = TRUE, ...)

## S4 method for signature 'GRanges,GRanges'
unionMC(x, y, ignore.strand = FALSE, simplify = TRUE, ...)

Arguments

x, y     GenomicRanges objects
...
ignore.strand  If set to TRUE, then the strand of x and y is set to "*" prior to any computation.
simplify  logical(1) If TRUE, any List columns will be returned as vectors where possible. This can only occur if single, unique entries are present in all initial elements.

Details

This extends the methods provided by setdiff, intersect and union so that mcols from x will be returned as part of the output.

Where output ranges map back to multiple ranges in x, CompressedList columns will be returned. By default, these will be simplified if possible, however this behaviour can be disabled by setting simplify = FALSE.

All columns will be returned which can also be time-consuming. A wise approach is to only provide columns you require as part of the query ranges x.

If more nuanced approaches are required, the returned columns can be further modified by many functions included in the plyranges package, such as mutate().
Value

A GRanges object with all mcols returned form the original object. If a range obtained by setdiff maps back to two or more ranges in the original set of Ranges, mcols will be returned as CompressedList columns

Examples

```r
x <- GRanges("chr1:1-100:+")
x$id <- "range1"
y <- GRanges(c("chr1:51-60:+", "chr1:21-30:-"))
setdiffMC(x, y)
setdiffMC(x, y, ignore.strand = TRUE)

# The intersection works similarly
intersectMC(x, y)

# Union may contain ranges not initially in x
unionMC(x, y)
unionMC(x, y, ignore.strand = TRUE)
```

stitchRanges

**Stitch Ranges within a given distance**

Description

Stitch together ranges within a given distance, using excluded ranges as barriers that cannot be crossed

Usage

```r
stitchRanges(x, exclude, maxgap = 12500L, ignore.strand = TRUE)
```

Arguments

- `x`: Ranges to be stitched together
- `exclude`: Ranges to exclude
- `maxgap`: The maximum distance between ranges to be stitched
- `ignore.strand`: logical

Details

Stitches together ranges within a given distance, using any ranges provided for exclusion as barriers between stitched ranges. This may be particularly useful if wanting to stitch enhancers whilst excluding promoters.

All inputs and outputs are Genomic Ranges objects
voomWeightsFromCPM

Value

A GRanges object

Examples

```r
x <- GRanges(c("chr1:1-10", "chr1:101-110", "chr1:201-210", "chr2:1-10"))
y <- GRanges("chr1:200:+")
stitchRanges(x, exclude = y, maxgap = 100)
```

---

**Description**

Estimate voom precision weights directly from CPM values

**Usage**

```r
voomWeightsFromCPM(
  cpm,
  design = NULL,
  w0 = NULL,
  lib.size = NULL,
  isLogCPM = TRUE,
  span = 0.5,
  ...
)
```

**Arguments**

- `cpm` Matrix of CPM or logCPM values
- `design` The design matrix for the experiment
- `w0` Initial vector of sample weights. Should be calculated using `arrayWeights`
- `lib.size` Initial library sizes. Must be provided as these are no estimable from CPM values
- `isLogCPM` logical(1). Indicates whether the data is log2 transformed already. Most commonly (e.g. if using the output of cqn) it will be,
- `span` Width of the smoothing window used for the lowess mean-variance trend. Expressed as a proportion between 0 and 1.
- `...` Passed to lmFit internally
Details

This function takes CPM or logCPM values and estimates the precision weights as would be done by providing counts directly to the `voom` function. Using this function enables the use of logCPM values which have been normalised using other methods such as Conditional-Quantile or Smooth-Quantile Normalisation.

The precision weights are returned as part of the `EList` output, and these are automatically passed to the function `lmFit` during model fitting. This will ensure that the mean-variance relationship is appropriate for the linear modelling steps as performed by limma.

Initial sample weights can be passed to the function, and should be calculated using `arrayWeights` called on the normalised logCPM values. The returned sample weights will be different to these, given that the function `voomWithQualityWeights` performs two rounds of estimation. The first is on the initial data, with the inappropriate mean-variance relationship, whilst the second round is after incorporation of the precision weights.

Value

An object of class `EList` as would be output by `voom`. Importantly, there will be no `genes` element, although this can be added later. Similarly, the returned `targets` element will only contain sample names and library sizes. This can be incorporated with any other metadata as required.

Plotting data is always returned, noting the the value `sx` has been offset by the library sizes and will be simple logCPM values. As such, the fitted `Amean` is also returned in this list element.

If initial sample weights were provided, modified weights will also be returned, as the initial function `voomWithQualityWeights` performs two rounds of estimation of sample weights. Here we would simply provide the initial weights a priori, with the second round performed within the function. Importantly, this second round of sample weight estimation uses the precision weights ensuring the correct mean-variance relationship is used for the final estimation of sample weights.

Examples

```r
bamFiles <- system.file("exdata","rep1.bam","rep2.bam"), package="csaw")
wc <- csaw::windowCounts(bamFiles, filter=1)
cpm <- edgeR::cpm(wc, log = TRUE)
el <- voomWeightsFromCPM(cpm, lib.size = wc$totals)
```
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