

# Package ‘comapr’

October 9, 2024

**Title** Crossover analysis and genetic map construction

**Version** 1.8.0

**Description** comapr detects crossover intervals for single gametes from their haplotype states sequences and stores the crossovers in GRanges object. The genetic distances can then be calculated via the mapping functions using estimated crossover rates for maker intervals. Visualisation functions for plotting interval-based genetic map or cumulative genetic distances are implemented, which help reveal the variation of crossovers landscapes across the genome and across individuals.

**biocViews** Software, SingleCell, Visualization, Genetics

**Depends** R (>= 4.1.0)

**Imports** methods, ggplot2, reshape2, dplyr, gridExtra, plotly, circlize, rlang, GenomicRanges, IRanges, foreach, BiocParallel, GenomeInfoDb, scales, RColorBrewer, tidyr, S4Vectors, utils, Matrix, grid, stats, SummarizedExperiment, plyr, Gviz

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**Encoding** UTF-8

**LazyData** false

**RoxygenNote** 7.2.3

**VignetteBuilder** knitr

**Suggests** BiocStyle, knitr, rmarkdown, testthat (>= 2.1.0), statmod

**git\_url** <https://git.bioconductor.org/packages/comapr>

**git\_branch** RELEASE\_3\_19

**git\_last\_commit** 553ee62

**git\_last\_commit\_date** 2024-04-30

**Repository** Bioconductor 3.19

**Date/Publication** 2024-10-09

**Author** Ruqian Lyu [aut, cre] (<<https://orcid.org/0000-0002-7736-6612>>)

**Maintainer** Ruqian Lyu <[xaoru.best@gmail.com](mailto:xaoru.best@gmail.com)>

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.change_missing	<i>change SNPs with genotype 'Fail' to NA</i>
-----------------	---

---

## Description

change SNPs with genotype 'Fail' to NA

**Usage**

```
.change_missing(s_gt, missing = "Fail")
```

**Arguments**

s\_gt            a column of labelled genotypes  
missing        the string used for encoding missing values default to Fail

**Details**

calculation.

**Value**

a vector of genotypes with Fail substituted by 'NA'

**Author(s)**

Ruqian Lyu

---

*.filterCOsExtra*            *Filter out doublet cells and uninformative SNPs*

---

**Description**

This function filter out cells that have been called too many crossovers due to diploid cell contamination or doublets. It also only keeps SNPs (rows) that ever contribute to a crossover interval. This function should be run for individual chromosomes and is called internally by 'readHapState'

**Usage**

```
.filterCOsExtra(  
  se,  
  minSNP = 30,  
  minlogllRatio = 200,  
  minCellsNP = 200,  
  bpDist = 100,  
  maxRawCO = 10,  
  biasTol = 0.45,  
  nmad = 1.5  
)
```

**Arguments**

se	the SummarizedExperiment object that contains the called haplotype state matrix in the assay field and haplotype segment information in the metadata field.
minSNP	the crossover(s) will be filtered out if introduced by a segment that has fewer than 'minSNP' SNPs to support.
minlogllRatio	the crossover(s) will be filtered out if introduced by a segment that has lower than 'minlogllRatio' to its reversed state.
minCellSNP	the minimum number of SNPs detected for a cell to be kept, used with 'nmds'
bpDist	the crossover(s) will be filtered out if introduced by a segment that is shorter than 'bpDist' basepairs.
maxRawCO	if a cell has more than 'maxRawCO' number of raw crossovers called across a chromosome, the cell is filtered out
biasTol	the SNP's haplotype ratio across all cells is assumed to be 1:1. This argument can be used for removing SNPs that have a biased haplotype. i.e. almost always inferred to be haplotype state 1. It specifies a bias tolerance value, SNPs with haplotype ratios deviating from 0.5 smaller than this value are kept. Only effective when number of cells are larger than 10
nmd	how many mean absolute deviations lower than the median number of SNPs per cell for a cell to be considered as low coverage cell and filtered Only effective when number of cells are larger than 10. When effective, this or 'minCellSNP', whichever is larger, is applied

**Details**

The 'logllRatio' value is returned by 'sgccaller' for each haplotype segment formed by consecutive SNPs that are called to have a same state. It is calculated by taking log of ratio (likelihood of SNPs with inferred states) and (likelihood of SNPs with reversed states)

**Value**

A 'RangedSummarizedExperiment' object that have different dims with input. the colnames are the cell barcodes, rowRanges specify the location of SNPs that contribute to crossovers.

**Author(s)**

Ruqian Lyu

---

.label\_gt

*'label\_gt' for changing genotypes in alleles format to labels*

---

**Description**

It turns a vector of Genotypes to a vector of Labels consist of 'Homo\_ref', 'Homo\_alt', and 'Het' given the known genotypes for reference and alternative strains.

**Usage**

```
.label_gt(s_gt, ref, alt, failed = "Fail")
```

**Arguments**

s_gt	s_gt, a vector of genotypes for one sample across markers
ref	ref, a vector of genotypes for reference strain across markers
alt	alt, a vector of genotypes for alternative strain across markers
failed	what was used for encoding failed genotype calling such as "Fail" in example

**Details**

This function takes the a sample's genotype across each SNP marker in alleles and compare with genotypes of in-bred reference and alternative strains to. If the sample's genotype for a particular SNP marker is the same with the reference strain, it is labelled as Homo\_ref homogeneous reference for a particular SNP marker; if the sample's genotype is the same with the alternative strain it is labelled as Homo\_alt homogeneous alternative for a particular SNP marker; if the sample's genotype is heterozygous then it is labeled as Het heterozygous for this particular genotypes. If it does not fall in any of the three cases, it is labelled as the string specified by the argument 'missing'.

Note that the wrong/failed genotype is labelled as the string in 'missing' after this function. If there is a different label for failed genotype, provide the label using the 'missing' argument.

**Value**

a vector of labels Homo\_ref, Homo\_alt, Het indicating the progeny's genotypes across markers

**Author(s)**

Ruqian Lyu

---

bootstrapDist

*bootstrapDist*

---

**Description**

Generating distribution of sample genetic distances

**Usage**

```
bootstrapDist(co_gr, B = 1000, mapping_fun = "k", group_by)
```

**Arguments**

co_gr	GRanges or RangedSummarizedExperiment object that contains the crossover counts for each marker interval across all samples. Returned by countCOs
B	integer the number of sampling times
mapping_fun	character default to "k" (kosambi mapping function). It can be one of the mapping functions: "k","h"
group_by	the prefix for each group that we need to generate distributions for(only when co_gr is a GRanges object). Or the column name for 'colData(co_gr)' that contains the group factor (only when co_gr is a RangedSummarizedExperiment object)

**Details**

It takes the crossover counts for samples in multiple groups that is returned by 'countCO'. It then draws samples from a group with replacement and calculate the distribution of relevant statistics.

**Value**

lists of numeric genetic distances for multiple samples

**Author(s)**

Ruqian Lyu

**Examples**

```
data(coCount)

bootsDiff <- bootstrapDist(coCount, group_by = "sampleGroup",B=10)
```

---

calGeneticDist	<i>calGeneticDist</i>
----------------	-----------------------

---

**Description**

Calculate genetic distances of marker intervals or binned-chromosome Given whether crossover happens in each marker interval, calculate the recombination fraction in samples and then derive the Haldane or Kosambi genetic distances via mapping functions

**Usage**

```
calGeneticDist(
  co_count,
  bin_size = NULL,
  mapping_fun = "k",
  ref_genome = "mm10",
  group_by = NULL,
```

```
    chrom_info = NULL
  )

## S4 method for signature 'GRanges,missing,ANY,ANY,missing'
calGeneticDist(
  co_count,
  bin_size = NULL,
  mapping_fun = "k",
  ref_genome = "mm10",
  group_by = NULL,
  chrom_info = NULL
)

## S4 method for signature 'GRanges,numeric,ANY,ANY,missing'
calGeneticDist(
  co_count,
  bin_size = NULL,
  mapping_fun = "k",
  ref_genome = "mm10",
  group_by = NULL,
  chrom_info = NULL
)

## S4 method for signature 'GRanges,missing,ANY,ANY,character'
calGeneticDist(
  co_count,
  bin_size = NULL,
  mapping_fun = "k",
  ref_genome = "mm10",
  group_by = NULL,
  chrom_info = NULL
)

## S4 method for signature 'GRanges,numeric,ANY,ANY,character'
calGeneticDist(
  co_count,
  bin_size = NULL,
  mapping_fun = "k",
  ref_genome = "mm10",
  group_by = NULL,
  chrom_info = NULL
)

## S4 method for signature 'RangedSummarizedExperiment,missing,ANY,ANY,missing'
calGeneticDist(
  co_count,
  bin_size = NULL,
  mapping_fun = "k",
```

```

    ref_genome = "mm10",
    group_by = NULL,
    chrom_info = NULL
)

## S4 method for signature
## 'RangedSummarizedExperiment,missing,ANY,ANY,character'
calGeneticDist(
  co_count,
  bin_size = NULL,
  mapping_fun = "k",
  ref_genome = "mm10",
  group_by = NULL,
  chrom_info = NULL
)

## S4 method for signature
## 'RangedSummarizedExperiment,numeric,ANY,ANY,character'
calGeneticDist(
  co_count,
  bin_size = NULL,
  mapping_fun = "k",
  ref_genome = "mm10",
  group_by = NULL,
  chrom_info = NULL
)

## S4 method for signature 'RangedSummarizedExperiment,numeric,ANY,ANY,missing'
calGeneticDist(
  co_count,
  bin_size = NULL,
  mapping_fun = "k",
  ref_genome = "mm10",
  group_by = NULL,
  chrom_info = NULL
)

```

### Arguments

co_count	GRange or RangedSummarizedExperiment object, returned by countCO
bin_size	The binning size for grouping marker intervals into bins. If not supplied, the original marker intervals are returned with converted genetic distances based on recombination rate
mapping_fun	The mapping function to use, can be one of "k" or "h" (kosambi or haldane)
ref_genome	The reference genome name. It is used to fetch the chromosome size information from UCSC database.
group_by	character vector contains the unique prefix of sample names that are used for defining different sample groups. Or the column name in colData(co_count)



that specify the group factor. If missing all samples are assumed to be from one group

chrom\_info A user supplied data.frame containing two columns with column names chrom and size, describing the chromosome names and lengths if not using ref\_genome from UCSC. If supplied, the 'ref\_genome' is ignored.

**Value**

GRanges object GRanges for marker intervals or binned intervals with Haldane or Kosambi centi-Morgans

**Examples**

```
data(coCount)
dist_se <- calGeneticDist(coCount)
# dist_se <- calGeneticDist(coCount,group_by="sampleGroup")
```

---

coCount	<i>RangedSummarizedExperiment object containing the crossover counts across samples for the list of SNP marker intervals</i>
---------	--

---

**Description**

RangedSummarizedExperiment object containing the crossover counts across samples for the list of SNP marker intervals

**Usage**

```
data(coCount)
```

**Format**

An object of class RangedSummarizedExperiment with 3 rows and 10 columns.

---

comapr	comapr <i>package</i>
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---

**Description**

crossover inference package

**Details**

See the README on [GitLab](#)

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combineHapState	<i>combineHapState</i>
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---

### Description

combine two 'RangedSummarizedExperiment' objects, each contains the haplotype state for a list of SNPs across a set of cells. The combined result will have cells from two individuals and merged list of SNPs from the two.

### Usage

```
combineHapState(rse1, rse2, groupName = c("Sample1", "Sample2"))
```

### Arguments

rse1	the first 'RangedSummarizedExperiment'
rse2	the second 'RangedSummarizedExperiment'
groupName	a character vector of length 2 that contains the first and the second group's names

### Value

A 'RangedSummarizedExperiment' that contains the cells and SNPs in both 'rse'

### Author(s)

Ruqian Lyu

### Examples

```
BiocParallel::register(BiocParallel::SnowParam(workers = 1))
demo_path <- paste0(system.file("extdata", package = "comapr"), "/")
s1_rse_state <- readHapState("s1", chroms=c("chr1"),
                           path=demo_path, barcodeFile=NULL, minSNP = 0,
                           minlogllRatio = 50,
                           bpDist = 100, maxRawCO=10,
                           minCellSNP = 1)

s2_rse_state <- readHapState("s2", chroms=c("chr1"),
                           path=demo_path,
                           barcodeFile=paste0(demo_path, "s2_barcode.txt"),
                           minSNP = 0,
                           minlogllRatio = 50,
                           bpDist = 100, maxRawCO=10,
                           minCellSNP = 1)

sb <- combineHapState(s1_rse_state, s2_rse_state)
```

---

correctGT	<i>correctGT</i>
-----------	------------------

---

**Description**

function for formatting and correction genotypes of markers

**Usage**

```
correctGT(gt_matrix, ref, alt, failed = "Fail", wrong_label = "Homo_ref")
```

**Arguments**

gt_matrix	the input genotype matrix of markers by samples with rownames as marker IDs and column names as sample IDs
ref	a vector of genotypes of the inbred reference strain
alt	a vector of genotypes of the inbred alternative strain
failed	what was used for encoding failed genotype calling such as "Fail" in example data snp_geno
wrong_label	what would be considered a wrong genotype label for example Homo_ref which should not be in the possible genotypes of BC1F1 samples

**Details**

This function changes genotype in alleles to genotype labels, change Homo\_ref to Hets/Fail, infer Failed genotype, and change "Failed" to NA for counting crossover later

This function changes genotype in alleles to labels by calling internal functions lable\_gt, and changes the wrong genotype Homo\_ref to Fail by calling .change\_missing.

**Value**

a genotype data.frame of sample genotypes with dimension as the input 'gt\_matrix' with genotypes converted to labels and failed calls are changed to NA.

**Author(s)**

Ruqian Lyu

**Examples**

```
data(snp_geno_gr)
data(parents_geno)
snp_gt_crt <- correctGT(gt_matrix = GenomicRanges::mcols(snp_geno_gr),
                        ref = parents_geno$ref,
                        alt = parents_geno$alt,
                        fail = "Fail",
                        wrong_label = "Homo_ref")
```

---

countBinState	<i>countBinState</i>
---------------	----------------------

---

**Description**

Bins the chromosome into supplied number of bins and find the state of the chromosome bins across all gamete cells

**Usage**

```
countBinState(chr, snpAnno, viState, genomeRange, ntile = 5)
```

**Arguments**

chr	character, the chromosome to check
snpAnno	data.frame, the SNP annotation for the supplied chromosome
viState	dgTMatrix/Matrix, the viterbi state matrix, output from ‘sgcocaller’
genomeRange	GRanges object with seqlengths information for the genome
ntile	integer, how many tiles the chromosome is binned into

**Details**

This function is used for checking whether chromosome segregation pattern obeys the expected ratio.

**Value**

a data.frame that contains chromosome bin segregation ratio

**Author(s)**

Ruqian Lyu

**Examples**

```
library(IRanges)
library(S4Vectors)

chrom_info <- GenomeInfoDb::getChromInfoFromUCSC("mm10")
seq_length <- chrom_info$size
names(seq_length) <- chrom_info$chrom

dna_mm10_gr <- GenomicRanges::GRanges(
  seqnames = Rle(names(seq_length)),
  ranges = IRanges(1, end = seq_length, names = names(seq_length)),
  seqlengths = seq_length)

GenomeInfoDb::genome(dna_mm10_gr) <- "mm10"
```

```

demo_path <- system.file("extdata",package = "comapr")
sampleName <- "s1"
chr <- "chr1"
vi_mtx <- Matrix::readMM(file = paste0(demo_path,"/", sampleName, "_",
                                       chr, "_vi mtx"))

snpAnno <- read.table(file = paste0(demo_path,"/", sampleName,
                                    "_", chr, "_snpAnnot.txt"),
                    stringsAsFactors = FALSE,
                    header = TRUE)

countBinState(chr = "chr1",snpAnno = snpAnno,
viState = vi_mtx,genomeRange = dna_mm10_gr, ntile = 1)

```

---

countCOs

*countCOs*


---

### Description

Count number of COs within each marker interval COs identified in the interval overlapping missing markers are distributed according to marker interval base-pair sizes. Genotypes encoded with "0" are treated as missing value.

### Usage

```

countCOs(geno)

## S4 method for signature 'GRanges'
countCOs(geno)

## S4 method for signature 'RangedSummarizedExperiment'
countCOs(geno)

```

### Arguments

**geno** GRanges object or RangedSummarizedExperiment object with genotype matrix that has SNP positions in the rows and cells/samples in the columns

### Value

GRanges object or RangedSummarizedExperiment with markers-intervals as rows and samples in columns, values as the number of COs estimated for each marker interval

### Author(s)

Ruqian Lyu

**Examples**

```
data(twoSamples)
cocount <- countCOs(twoSamples)
```

---

countGT

*countGT*

---

**Description**

count how many samples have genotypes calls across markers and count how many markers that each individual has called genotypes for. This function helps identify poor samples or poor markers for filtering. It can also generate plots that help identify outlier samples/markers

**Usage**

```
countGT(geno, plot = TRUE, interactive = FALSE)
```

**Arguments**

geno	the genotype data.frame of markers by samples from output of function correctGT
plot	it determines whether a plot will be generated, defaults to TRUE
interactive	it determines whether an interactive plot will be generated

**Value**

A list of two elements including `n_markers` and `n_samples`

**Author(s)**

Ruqian Lyu

**Examples**

```
data(snp_geno_gr)
genotype_counts <- countGT(GenomicRanges::mcols(snp_geno_gr))
```

---

fill_fail	<i>Infer the genotype of failed SNPs If we have a Fail in the genotype data and the Fail in a block of either Home_alt, or Het, we fill in the Fails using values of the ones adjacent to it, otherwise they remain as "Fail" to indicate missing values.</i>
-----------	---

---

### Description

Infer the genotype of failed SNPs If we have a Fail in the genotype data and the Fail in a block of either Home\_alt, or Het, we fill in the Fails using values of the ones adjacent to it, otherwise they remain as "Fail" to indicate missing values.

### Usage

```
fill_fail(s_gt, fail = "Fail", chr = NULL)
```

### Arguments

s_gt	a column of labelled genotypes
fail	the string that is used for encoding failed genotype results, default to Fail
chr	the factor vector indicating which chromosomes the markers are on, default to NULL which means the input markers are all on the same chromosome.

### Value

a vector of genotypes with Failed genotype imputed or changed to 'NA' if not imputable

### Author(s)

Ruqian Lyu

---

filterGT	<i>filterGT</i>
----------	-----------------

---

### Description

Filter markers or samples that have too many missing values

### Usage

```
filterGT(geno, min_markers = 5, min_samples = 3)

## S4 method for signature 'matrix,numeric,numeric'
filterGT(geno, min_markers = 5, min_samples = 3)

## S4 method for signature 'GRanges,numeric,numeric'
filterGT(geno, min_markers = 5, min_samples = 3)
```

**Arguments**

**geno**                the genotype data.frame of markers by samples from output of function correctGT  
**min\_markers**        the minimum number of markers for a sample to be kept  
**min\_samples**        the minimum number of samples for a marker to be kept

**Details**

This function takes the geno data.frame and filter the data.frame by the provided cut-offs.

**Value**

The filtered genotype matrix

**Author(s)**

Ruqian Lyu

**Examples**

```
data(snp_geno_gr)
corrected_geno <- filterGT(snp_geno_gr, min_markers = 30, min_samples = 2)
```

---

findDupSamples        *findDupSamples*

---

**Description**

Find the duplicated samples by look at the number of matching genotypes in all pair-wise samples

**Usage**

```
findDupSamples(geno, threshold = 0.99, in_text = FALSE)
```

**Arguments**

**geno**                the genotype data.frame of markers by samples from output of function correctGT  
**threshold**           the frequency cut-off of number of matching genotypes out of all genotypes for determining whether the pair of samples are duplicated, defaults to 0.99. NAs are regarded as same genotypes for two samples if they both have NA for a marker.  
**in\_text**             whether text of frequencies should be displayed in the heatmap cells

**Value**

The paris of duplicated samples.



**Author(s)**

Ruqian Lyu

**Examples**

```

data(snp_genotype)
or_genotype <- snp_genotype[,grep("X",colnames(snp_genotype))]
rownames(or_genotype) <- paste0(snp_genotype$CHR,"_",snp_genotype$POS)
or_genotype[,1] <- or_genotype[,5]
cr_genotype <- correctGT(or_genotype,ref = snp_genotype$C57BL.6J,
                        alt = snp_genotype$FVB.NJ..i.)
dups <- findDupSamples(cr_genotype)

```

getAFTracks

*getAFTracks***Description**

Generate the raw alternative allele frequencies tracks for all cells in the columns of provided 'co\_count'

**Usage**

```

getAFTracks(
  chrom = "chr1",
  path_loc = "./output/firstBatch/WC_522/",
  sampleName = "WC_522",
  nwindow = 80,
  barcodeFile,
  co_count,
  snp_track = NULL
)

```

**Arguments**

chrom	the chromosome
path_loc	the path prefix to the output files from sscocaller including "*_totalCount.mtx" and "_altCount.mtx"
sampleName	the sample name, which is the prefix of sscocaller's output files
nwindow	the number of windows for binning the chromosome
barcodeFile	the barcode file containing the list of cell barcodes used as the input file for sscocaller
co_count	'GRange' or 'RangedSummarizedExperiment' object, returned by countCO that contains the crossover intervals and the number of crossovers in each cell.
snp_track	the SNP position track which is used for obtaining the SNP chromosome locations. It could be omitted and the SNP positions will be acquired from the "*_snpAnnot.txt" file.

**Value**

a list object, in which each element is a list of two items with the cell's alternative allele frequency DataTrack and the called crossover ranges.

**Author(s)**

Ruqian Lyu

**Examples**

```
demo_path <- system.file("extdata",package = "comapr")
s1_rse_state <- readHapState("s1",chroms=c("chr1"),
                           path=demo_path,barcodeFile=NULL,minSNP = 0,
                           minlogllRatio = 50,
                           bpDist = 100,maxRawCO=10,
                           minCellSNP = 0)
s1_counts <- countCOs(s1_rse_state)

af_co_tracks <- getAFTracks(chrom ="chr1",
                           path_loc = demo_path,
                           sampleName = "s1",
                           barcodeFile = file.path(demo_path,
                                                    "s1_barcodes.txt"),
                           co_count = s1_counts)
```

---

getCellAFTrack	<i>getCellAFTrack Generates the DataTracks for plotting AF and crossover regions</i>
----------------	--

---

**Description**

It plots the raw alternative allele frequencies and highlight the crossover regions for the selected cell.

It plots the raw alternative allele frequencies and highlight the crossover regions for the selected cell.

**Usage**

```
getCellAFTrack(
  chrom = "chr1",
  path_loc = "./output/firstBatch/WC_522/",
  sampleName = "WC_522",
  nwindow = 80,
  barcodeFile,
  cellBarcode,
  co_count,
  snp_track = NULL,
```

```

    chunk = 1000L
  )

  getCellAFTrack(
    chrom = "chr1",
    path_loc = "./output/firstBatch/WC_522/",
    sampleName = "WC_522",
    nwindow = 80,
    barcodeFile,
    cellBarcode,
    co_count,
    snp_track = NULL,
    chunk = 1000L
  )

```

### Arguments

chrom	the chromosome
path_loc	the path prefix to the output files from sscocaller including <code>"*_totalCount.mtx"</code> and <code>"_altCount.mtx"</code>
sampleName	the sample name, which is the prefix of sscocaller's output files
nwindow	the number of windows for binning the chromosome
barcodeFile	the barcode file containing the list of cell barcodes used as the input file for sscocaller
cellBarcode	the selected cell barcode
co_count	'GRange' or 'RangedSummarizedExperiment' object, returned by countCO that contains the crossover intervals and the number of crossovers in each cell.
snp_track	the SNP position track which is used for obtaining the SNP chromosome locations. It could be omitted and the SNP positions will be acquired from the <code>"*_snpAnnot.txt"</code> file.
chunk	An integer scalar indicating the chunk size to use, i.e., number of rows to read at any one time.

### Value

The DataTrack object defined in [DataTrack](#)

The DataTrack object defined in [DataTrack](#)

### Author(s)

Ruqian Lyu

### Examples

```

demo_path <- paste0(system.file("extdata", package = "comapr"), "/")
s1_rse_state <- readHapState("s1", chroms=c("chr1"),
                             path=demo_path, barcodeFile=NULL, minSNP = 0,

```

```

        minlogllRatio = 50,
        bpDist = 100,maxRawCO=10,
        minCellSNP = 0)
s1_counts <- countCOs(s1_rse_state)
af_co_tracks <- getCellAFTrack(chrom = "chr1",
                              path_loc = demo_path,
                              sampleName = "s1",
                              barcodeFile = paste0(demo_path,
                                                    "s1_barcodes.txt"),
                              cellBarcode = "BC1",
                              co_count = s1_counts)

demo_path <-paste0(system.file("extdata",package = "comapr"),"/")
s1_rse_state <- readHapState("s1",chrom=c("chr1"),
                            path=demo_path,barcodeFile=NULL,minSNP = 0,
                            minlogllRatio = 50,
                            bpDist = 100,maxRawCO=10,
                            minCellSNP = 0)
s1_counts <- countCOs(s1_rse_state)
af_co_tracks <- getCellAFTrack(chrom = "chr1",
                              path_loc = demo_path,
                              sampleName = "s1",
                              barcodeFile = paste0(demo_path,
                                                    "s1_barcodes.txt"),
                              cellBarcode = "BC1",
                              co_count = s1_counts)

```

---

getCellCORange

*getCellCORange*


---

### Description

It finds the crossover intervals for a selected cell

### Usage

```
getCellCORange(co_count, cellBarcode)
```

### Arguments

co\_count            ‘GRanges’ or ‘RangedSummarizedExperiment’ object,  
cellBarcode        the selected cell’s barcode

### Value

GRange object containing the crossover intervals for the selected cell

**Author(s)**

Ruqian Lyu

**Examples**

```

demo_path <- paste0(system.file("extdata", package = "comapr"), "/")
s1_rse_state <- readHapState("s1", chroms=c("chr1"),
                           path=demo_path, barcodeFile=NULL, minSNP = 0,
                           minlogllRatio = 50,
                           bpDist = 100, maxRawCO=10,
                           minCellSNP = 0)
s1_counts <- countCOs(s1_rse_state)

co_ranges <- getCellCORange(cellBarcode = "BC1",
                           co_count = s1_counts)

```

---

getCellDPTrack	<i>getCellDPTrack Generates the DataTrack for plotting DP of a selected cell</i>
----------------	--

---

**Description**

It plots the total allele counts for the selected cell.

**Usage**

```

getCellDPTrack(
  chrom = "chr1",
  path_loc = "./output/firstBatch/WC_522/",
  sampleName = "WC_522",
  nwindow = 80,
  barcodeFile,
  cellBarcode,
  snp_track = NULL,
  chunk = 1000L,
  log = TRUE,
  plot_type = "hist"
)

```

**Arguments**

chrom	the chromosome
path_loc	the path prefix to the output files from sscocaller including "*_totalCount.mtx"
sampleName	the sample name, which is the prefix of sscocaller's output files
nwindow	the number of windows for binning the chromosome
barcodeFile	the barcode file containing the list of cell barcodes used as the input file for sscocaller

cellBarcode	the selected cell barcode
snp_track	the SNP position track which is used for obtaining the SNP chromosome locations. It could be omitted and the SNP positions will be acquired from the "*_snpAnnot.txt" file.
chunk	A integer scalar indicating the chunk size to use, i.e., number of rows to read at any one time.
log	whether the histogram of SNP density should be plotted on log scale (log10)
plot_type	the DataTrack plot type, default to be 'hist'

**Value**

The DataTrack object defined in [DataTrack](#)

**Author(s)**

Ruqian Lyu

**Examples**

```
demo_path <- system.file("extdata",package = "comapr")
s1_rse_state <- readHapState("s1",chroms=c("chr1"),
                           path=demo_path,barcodeFile=NULL,minSNP = 0,
                           minlogllRatio = 50,
                           bpDist = 100,maxRawCO=10,
                           minCellSNP = 0)
s1_counts <- countCOs(s1_rse_state)
dp_co_tracks <- getCellDPTrack(chrom ="chr1",
                              path_loc = demo_path,
                              sampleName = "s1",
                              barcodeFile = file.path(demo_path,
                                                       "s1_barcodes.txt"),
                              cellBarcode = "BC1")
```

---

getDistortedMarkers    *getDistortedMarkers*

---

**Description**

Marker segregation distortion detection using chisq-test

**Usage**

```
getDistortedMarkers(geno, p = c(0.5, 0.5), adj.method = "BH")
```

**Arguments**

**geno**                the genotype data.frame of markers by samples from output of function correctGT  
**p**                     the expected genotype ratio in a numeric vector, defaults to c(0.5,0.5)  
**adj.method**        Methods to adjust for multiple comparisons, defaults to "BH"

**Details**

We expect the genotypes to appear with the frequencies of 1:1 homo\_alt:hets. We use chisq.test for finding markers that have genotypes among samples that are significantly different from the 1:1 ratio and report them

**Value**

data.frame with each row representing one SNP marker and columns containing the chisq.test results

**Author(s)**

Ruqian Lyu

**Examples**

```

data(parents_genotype)
data(snp_genotype)
corrected_genotype <- correctGT(gt_matrix = GenomicRanges::mcols(snp_genotype),
  ref = parents_genotype$ref, alt = parents_genotype$alt, fail = "Fail",
  wrong_label = "Homo_ref")
GenomicRanges::mcols(snp_genotype) <- corrected_genotype
corrected_genotype <- filterGT(snp_genotype, min_markers = 30, min_samples = 2)
pvalues <- getDistortedMarkers(GenomicRanges::mcols(corrected_genotype))
  
```

---

getMeanDPTrack

*getMeanDPTrack*

---

**Description**

Generate the mean DP (Depth) DataTrack (from Gviz) for cells

**Usage**

```

getMeanDPTrack(
  chrom = "chr1",
  path_loc,
  nwindow = 80,
  sampleName,
  barcodeFile,
  plot_type = "hist",
  )
  
```





---

getSNPDensityTrack     *getSNPDensityTrack*

---

### Description

Generate the SNP density DataTrack (from 'Gviz') for selected chromosome

### Usage

```
getSNPDensityTrack(  
  chrom = "chr1",  
  sampleName = "s1",  
  path_loc = ".",  
  nwindow = 80,  
  plot_type = "hist",  
  log = TRUE  
)
```

### Arguments

chrom	the chromosome
sampleName	the sample name, which is the prefix of sscocaller's output files
path_loc	the path prefix to the output files from sscocaller including "*_totalCount.mtx" and "_altCount.mtx"
nwindow	the number of windows for binning the chromosome
plot_type	the DataTrack plot type, default to be 'hist'
log	whether the histogram of SNP density should be plotted on log scale (log10)

### Value

DataTrack object plotting the SNP density histogram

### Author(s)

Ruqian Lyu

### Examples

```
demo_path <- system.file("extdata", package = "comapr")  
snp_track <- getSNPDensityTrack(chrom = "chr1",  
                                path_loc = demo_path,  
                                sampleName = "s1")
```

---

parents_genotype	<i>Parents' genotype for F1 samples in 'snp_genotype'</i>
------------------	---

---

### Description

Parents' genotype for F1 samples in 'snp\_genotype'

### Usage

```
data(parents_genotype)
```

### Format

A data.frame:

**C57BL.6J** genotype of reference mouse train across markers

**FVB.NJ.i** genotype of alternative mouse train across markers

---

perCellChrQC	<i>perCellChrQC</i>
--------------	---------------------

---

### Description

A function that parses output ('\_viSegInfo.txt') from 'sgccaller' <https://gitlab.svi.edu.au/biocellgen-public/sgccaller> and generate cell cell (per chr) summary statistics

### Usage

```
perCellChrQC(
  sampleName,
  chroms = c("chr1", "chr7", "chr15"),
  path,
  barcodeFile = NULL,
  doPlot = TRUE
)
```

### Arguments

sampleName	the name of the sample to parse which is used as prefix for finding relevant files for the underlying sample
chroms	the character vectors of chromosomes to parse. Multiple chromosomes' results will be concated together.
path	the path to the files, with name patterns *chrom_vi.mtx, *chrom_viSegInfo.txt, end with slash
barcodeFile	defaults to NULL, it is assumed to be in the same directory as the other files and with name sampleName_barcodes.txt
doPlot	whether a plot should returned, default to TRUE

**Value**

a list object that contains the data.frame with summarised statistics per chr per cell and a plot (if doPlot)

**Author(s)**

Ruqian Lyu

**Examples**

```
demo_path <-system.file("extdata",package = "comapr")
pcQC <- perCellChrQC(sampleName="s1",chroms=c("chr1"),
path=demo_path,
barcodeFile=NULL)
```

---

permutDist	<i>permutDist</i>
------------	-------------------

---

**Description**

Permutation test of two sample groups

**Usage**

```
permutDist(co_gr, B = 100, mapping_fun = "k", group_by)
```

**Arguments**

co_gr	GRanges or RangedSummarizedExperiment object that contains the crossover counts for each marker interval across all samples. Returned by countCOs
B	integer the number of sampling times
mapping_fun	character default to "k" (kosambi mapping function). It can be one of the mapping functions: "k","h"
group_by	the prefix for each group that we need to generate distributions for(only when co_gr is a GRanges object). Or the column name for 'colData(co_gr)' that contains the group factor (only when co_gr is a RangedSummarizedExperiment object)

**Details**

It shuffles the group labels for the samples and calculate a difference between two groups after shuffling.

**Value**

A list of three elements. 'permutes' of length B with numeric differences of permuted group differences, 'observed\_diff' the observed genetic distances of two groups, 'nSample', the number of samples in the first and second group.

**Author(s)**

Ruqian Lyu

**Examples**

```
data(coCount)
perms <- permuteDist(coCount, group_by = "sampleGroup",B=10)
```

---

perSegChrQC

*perSegChrQC*


---

**Description**

Plots the summary statistics of segments that are generated by ‘sgccaller’ <https://gitlab.svi.edu.au/biocellgen-public/sgccaller> which have been detected by finding consecutive viter states along the list of SNP markers.

**Usage**

```
perSegChrQC(
  sampleName,
  chroms = c("chr1", "chr7", "chr15"),
  path,
  barcodeFile = NULL,
  maxRawCO = 10
)
```

**Arguments**

sampleName	the name of the sample to parse which is used as prefix for finding relevant files for the underlying sample
chroms	the vector of chromosomes
path	the path to the files, with name patterns *chrom_vi.mtx, *chrom_viSegInfo.txt, end with slash
barcodeFile	defaults to NULL, it is assumed to be in the same directory as the other files and with name sampleName_barcodes.txt
maxRawCO	if a cell has more than ‘maxRawCO’ number of raw crossovers called across a chromosome, the cell is filtered out##’

**Details**

It provides guidance in filtering out close double crossovers that are not likely biological but due to technical reasons as well as crossovers that are supported by fewer number of SNPs at the ends of the chromosomes.

**Value**

Histogram plots for statistics summarized across all Viterbi state segments

**Author(s)**

Ruqian Lyu

**Examples**

```
demo_path <- system.file("extdata",package = "comapr")
s1_rse_qc <- perSegChrQC(sampleName="s1",
                        chroms=c("chr1"),
                        path=demo_path, maxRawCO=10)
```

---

plotCount

*plotCount*

---

**Description**

Plot the number of COs per sample group or per chromosome

**Usage**

```
plotCount(
  co_count,
  by_chr = FALSE,
  group_by = "sampleGroup",
  plot_type = "error_bar"
)

## S4 method for signature 'RangedSummarizedExperiment,missing,missing'
plotCount(
  co_count,
  by_chr = FALSE,
  group_by = "sampleGroup",
  plot_type = "error_bar"
)

## S4 method for signature 'RangedSummarizedExperiment,missing,character'
plotCount(
  co_count,
  by_chr = FALSE,
  group_by = "sampleGroup",
  plot_type = "error_bar"
)
```

```
## S4 method for signature 'RangedSummarizedExperiment,logical,character'  
plotCount(  
  co_count,  
  by_chr = FALSE,  
  group_by = "sampleGroup",  
  plot_type = "error_bar"  
)  
  
## S4 method for signature 'RangedSummarizedExperiment,logical,missing'  
plotCount(  
  co_count,  
  by_chr = FALSE,  
  group_by = "sampleGroup",  
  plot_type = "error_bar"  
)  
  
## S4 method for signature 'GRanges,logical,missing'  
plotCount(  
  co_count,  
  by_chr = FALSE,  
  group_by = "sampleGroup",  
  plot_type = "error_bar"  
)  
  
## S4 method for signature 'GRanges,missing,missing'  
plotCount(  
  co_count,  
  by_chr = FALSE,  
  group_by = "sampleGroup",  
  plot_type = "error_bar"  
)  
  
## S4 method for signature 'GRanges,missing,character'  
plotCount(  
  co_count,  
  by_chr = FALSE,  
  group_by = "sampleGroup",  
  plot_type = "error_bar"  
)  
  
## S4 method for signature 'GRanges,logical,character'  
plotCount(  
  co_count,  
  by_chr = FALSE,  
  group_by = "sampleGroup",  
  plot_type = "error_bar"  
)
```

**Arguments**

co_count	GRange or RangedSummarizedExperiment object, returned by countCO
by_chr	whether it should plot each chromosome separately
group_by	the column name in 'colData(co_count)' that specify the grouping factor. Or the character vector contains the unique prefix of sample names that are used for defining different sample groups. If missing all samples are assumed to be from one group
plot_type	determines what type the plot will be, choose from "error_bar" or "hist". Only relevant when by_chr=TRUE

**Value**

ggplot object

**Examples**

```
demo_path <- paste0(system.file("extdata", package = "comapr"), "/")
s1_rse_state <- readHapState("s1", chroms=c("chr1"),
                           path=demo_path, barcodeFile=NULL, minSNP = 0,
                           minlogllRatio = 50,
                           bpDist = 100, maxRawCO=10,
                           minCellSNP = 0)
s1_count <- countCOs(s1_rse_state)
plotCount(s1_count)
```

---

plotGeneticDist	<i>plotGeneticDist</i>
-----------------	------------------------

---

**Description**

Plotting the calculated genetic distanced for each bin or marker interval supplied by the GRanges object

**Usage**

```
plotGeneticDist(gr, bin = TRUE, chr = NULL, cumulative = FALSE)
```

**Arguments**

gr	GRanges object with genetic distances calculated for marker intervals
bin	TRUE or FALSE, indicating whether the supplied GRange object is for binned interval
chr	the specific chrs selected to plot
cumulative	TRUE or FALSE, indicating whether it plots the bin-wise genetic distances or the cumulative distances

**Value**

ggplot2 plot

**Author(s)**

Ruqian Lyu

**Examples**

```
data(coCount)
dist_se <- calGeneticDist(coCount)
plotGeneticDist(SummarizedExperiment::rowRanges(dist_se))
```

---

plotGTFreq

*plotGTFreq*

---

**Description**

Function to plot the genotypes for all samples faceted by genotype

**Usage**

```
plotGTFreq(geno)
```

**Arguments**

geno                    the genotype data.frame of markers by samples from output of function correctGT

**Value**

A ggplot object

**Author(s)**

Ruqian Lyu

**Examples**

```
data(snp_geno)
or_geno <- snp_geno[,grep("X",colnames(snp_geno))]
rownames(or_geno) <- paste0(snp_geno$CHR,"_",snp_geno$POS)
or_geno[1,] <- rep("Fail",dim(or_geno)[2])
cr_geno <- correctGT(or_geno,ref = snp_geno$C57BL.6J,
                    alt = snp_geno$FVB.NJ..i.)
ft_gt <- filterGT(cr_geno)
plotGTFreq(ft_gt)
```



---

plotWholeGenome	<i>Plot cumulative genetic distances across the genome</i>
-----------------	--

---

**Description**

This function takes the calculated genetic distances for all marker intervals across all chromosomes provided and plot the cumulative genetic distances

**Usage**

```
plotWholeGenome(gr)
```

**Arguments**

gr                    GRanges object with genetic distances calculated for marker intervals

**Value**

A ggplot object

**Examples**

```
data(coCount)
dist_se <- calGeneticDist(coCount)
plotWholeGenome(SummarizedExperiment::rowRanges(dist_se))
```

---

readColMM	<i>readColMM</i>
-----------	------------------

---

**Description**

Modified the 'Matrix::readMM' function for reading matrices stored in the Harwell-Boeing or MatrixMarket formats but only reads selected column.

**Usage**

```
readColMM(file, which.col, chunk = 1000L)
```

**Arguments**

file                    the name of the file to be read from as a character scalar. Those storing matrices in the MatrixMarket format usually end in ".mtx".

which.col                An integer scalar, the column index

chunk                    An integer scalar indicating the chunk size to use, i.e., number of rows to read at any one time.

**Details**

See [readMM](#)

**Value**

A sparse matrix object that inherits from the "Matrix" class which the original dimensions. To get the vector of the specified column, one need to subset the matrix to select the column with the same index.

**Author(s)**

Ruqian Lyu

**Examples**

```
demo_path <-paste0(system.file("extdata",package = "comapr"),"/")
readColMM(file = paste0(demo_path,"s1_chr1_vi.mtx"), which.col=2,chunk=2)
```

---

readHapState	<i>readHapState</i>
--------------	---------------------

---

**Description**

A function that parses the viterbi state matrix (in .mtx format), barcode.txt and snpAnno.txt files for each individual.

**Usage**

```
readHapState(
  sampleName,
  chroms = c("chr1"),
  path,
  barcodeFile = NULL,
  minSNP = 30,
  minlogllRatio = 200,
  bpDist = 100,
  maxRawCO = 10,
  nmad = 1.5,
  minCellSNP = 200,
  biasTol = 0.45
)
```

**Arguments**

sampleName	the name of the sample to parse which is used as prefix for finding relevant files for the underlying sample
chroms	the character vectors of chromosomes to parse. Multiple chromosomes' results will be concated together.
path	the path to the files, with name patterns *chrom_vi.mtx, *chrom_viSegInfo.txt, end with slash
barcodeFile	if NULL, it is assumed to be in the same directory as the other files and with name sampleName_barcode.txt
minSNP	the crossover(s) will be filtered out if introduced by a segment that has fewer than 'minSNP' SNPs to support.
minlogllRatio	the crossover(s) will be filtered out if introduced by a segment that has lower than 'minlogllRatio' to its reversed state.
bpDist	the crossover(s) will be filtered out if introduced by a segment that is shorter than 'bpDist' basepairs. It can be a single value or a vector that is the same length and order with 'chroms'.
maxRawCO	if a cell has more than 'maxRawCO' number of raw crossovers called across a chromosome, the cell is filtered out
nmad	how many mean absolute deviations lower than the median number of SNPs per cell for a cell to be considered as low coverage cell and filtered. Only effective when number of cells are larger than 10. When effective, this or 'minCellSNP', whichever is larger, is applied
minCellSNP	the minimum number of SNPs detected for a cell to be kept, used with 'nmads'
biasTol	the SNP's haplotype ratio across all cells is assumed to be 1:1. This argument can be used for removing SNPs that have a biased haplotype. i.e. almost always inferred to be haplotype state 1. It specifies a bias tolerance value, SNPs with haplotype ratios deviating from 0.5 smaller than this value are kept. Only effective when number of cells are larger than 10

**Value**

a RangedSummarizedExperiment with rowRanges as SNP positions that contribute to crossovers in any cells. colData contains cells annotation including barcodes and sampleName.

**Author(s)**

Ruqian Lyu

**Examples**

```
demo_path <- system.file("extdata", package = "comapr")
s1_rse_state <- readHapState(sampleName="s1", chroms=c("chr1"),
  path=paste0(demo_path, "/"),
  barcodeFile=NULL, minSNP = 0, minlogllRatio = 50,
  bpDist = 100, maxRawCO=10, minCellSNP=3)
s1_rse_state
```

---

snp\_genotype

*Markers by genotype results for a group of samples*

---

### Description

Markers by genotype results for a group of samples

### Usage

```
data(snp_genotype)
```

### Format

A data frame with columns:

**C57BL.6J** genotype of reference mouse train across markers  
**FVB.NJ.i.** genotype of alternative mouse train across markers  
**POS** SNP marker base-pair location  
**CHR** SNP marker chromosome location  
**X100** a mouse sample  
**X101** a mouse sample  
**X102** a mouse sample  
**X103** a mouse sample  
**X104** a mouse sample  
**X105** a mouse sample  
**X106** a mouse sample  
**X107** a mouse sample  
**X108** a mouse sample  
**X109** a mouse sample  
**X110** a mouse sample  
**X111** a mouse sample  
**X112** a mouse sample  
**X113** a mouse sample  
**X92** a mouse sample  
**X93** a mouse sample  
**X94** a mouse sample  
**X95** a mouse sample  
**X96** a mouse sample  
**X97** a mouse sample  
**X98** a mouse sample  
**X99** a mouse sample  
**rsID** the SNP ID

**Source**

Statistics Canada. Table 001-0008 - Production and farm value of maple products, annual. <http://www5.statcan.gc.ca/cansim/>

---

snp\_genogr

*Markers by genotype results for a group of samples*

---

**Description**

Markers by genotype results for a group of samples

**Usage**

```
data(snp_genogr)
```

**Format**

A GRanges object:

**X100** a mouse sample  
**X101** a mouse sample  
**X102** a mouse sample  
**X103** a mouse sample  
**X104** a mouse sample  
**X105** a mouse sample  
**X106** a mouse sample  
**X107** a mouse sample  
**X108** a mouse sample  
**X109** a mouse sample  
**X110** a mouse sample  
**X111** a mouse sample  
**X112** a mouse sample  
**X113** a mouse sample  
**X92** a mouse sample  
**X93** a mouse sample  
**X94** a mouse sample  
**X95** a mouse sample  
**X96** a mouse sample  
**X97** a mouse sample  
**X98** a mouse sample  
**X99** a mouse sample  
**rsID** the SNP ID

**Source**

TBD

---

twoSamples	<i>RangedSummarizedExperiment object containing the Viterbi states SNP markers for samples from two groups. 'colData(twoSamples)' contains the sample group factor.</i>
------------	---

---

**Description**

RangedSummarizedExperiment object containing the Viterbi states SNP markers for samples from two groups. 'colData(twoSamples)' contains the sample group factor.

**Usage**

```
data(twoSamples)
```

**Format**

An object of class RangedSummarizedExperiment with 6 rows and 10 columns.

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