Package ‘cellxgenedp’

May 3, 2024

**Title**  Discover and Access Single Cell Data Sets in the CELLxGENE Data Portal

**Version**  1.8.0

**Description**  The cellxgene data portal

(https://cellxgene.cziscience.com/) provides a graphical user interface to collections of single-cell sequence data processed in standard ways to ‘count matrix’ summaries. The cellxgenedp package provides an alternative, R-based interface, allowing data discovery, viewing, and downloading.

**License**  Artistic-2.0

**Encoding**  UTF-8

**Collate**  

**Depends**  dplyr

**Imports**  httr, curl, utils, tools, shiny, DT, rjsoncons

**Suggests**  zellkonverter, SingleCellExperiment, HDF5Array, tidyr, 
BiocStyle, knitr, rmardown, testthat (>= 3.0.0), mockery

**biocViews**  SingleCell, DataImport, ThirdPartyClient

**URL**  https://mtmorgan.github.io/cellxgenedp/, 
https://github.com/mtmorgan/cellxgenedp

**BugReports**  https://github.com/mtmorgan/cellxgenedp/issues

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collections

Query cellxgene collections, datasets, and files

description

files_download() retrieves one or more cellxgene files to a cache on the local system.

links(), authors() and publisher_metadata() are helper functions to extract ‘nested’ information from collections.

usage

collections(cellxgene_db = db())

datasets(cellxgene_db = db())

datasets_visualize(tbl)

files(cellxgene_db = db())

files_download(tbl, dry.run = TRUE, cache.path = .cellxgene_cache_path())

links(cellxgene_db = db())

authors(cellxgene_db = db())

publisher_metadata(cellxgene_db = db())
collections

Arguments

cellxgene_db  an optional `cellxgene_db` object, as returned by `db()`.

tbl  a tibble() typically derived from `datasets(db)` or `files(db)` and containing columns `dataset_id` (for `datasets_visualize()`), or columns `dataset_id`, `file_id`, and `filetype` (for `files_download()`).

dry.run  logical(1) indicating whether the (often large) file(s) in tbl should be downloaded to a local cache. Files are not downloaded when `dry.run = TRUE` (default).

cache.path  character(1) directory in which to cache downloaded files. The directory must already exist. The default is `tools::R_user_dir("cellxgenedp", "cache"), a package-specific path in the user home directory.

Value

Each function returns a tibble describing the corresponding component of the database.

files_download() returns a character() vector of paths to the local files.

links() returns a tibble of external links associated with each collection. Common links include DOI, raw data / data sources, and lab websites.

authors() returns a tibble of authors associated with each collection.

publisher_metadata() returns a tibble of publisher metadata (journal, publicate date, doi) associated with each collection.

Examples

db <- db()

collections(db)

collections(db) |>  
dplyr::glimpse()

datasets(db) |>  
dplyr::glimpse()

if (interactive()) {
  ## visualize the first dataset
  datasets(db) |>  
    dplyr::slice(1) |>  
      datasets_visualize()
}

files(db) |>  
  dplyr::glimpse()

## Not run:
files(db) |>  
  dplyr::slice(1) |>


cxg

Shiny application for discovering, viewing, and downloading cellxgene data

Description
Shiny application for discovering, viewing, and downloading cellxgene data

Usage

cxg(as = c("tibble", "sce"))

Arguments

as character(1) Return value when quitting the shiny application. "tibble" returns a tibble describing selected datasets (including the location on disk of the downloaded file). "sce" returns a list of dataset files imported to R as SingleCellExperiment objects.

Value

cxg() returns either a tibble describing datasets selected in the shiny application, or a list of datasets imported into R as SingleCellExperiment objects.

Examples

if (interactive())
cxg()
**db**

Retrieve updated cellxgene database metadata

**Usage**

\[
db(overwrite = .db_online() \&\& .db_first())
\]

**Arguments**

- `overwrite`: logical(1) indicating whether the database of collections should be updated from the internet (the default, when internet is available and, in an interactive session, the user requests the update), or read from disk (assuming previous successful access to the internet). `overwrite = FALSE` might be useful for reproducibility, testing, or when working in an environment with restricted internet access.

**Details**

The database is retrieved from the cellxgene data portal web site. ’collections’ metadata are retrieved on each call; metadata on each collection is cached locally for re-use.

**Value**

\[db()\] returns an object of class ’cellxgene_db’, summarizing available collections, datasets, and files.

**Examples**

\[db()\]

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**FACETS**

Facets available for querying cellxgene data

**Description**

FACETS is a character vector of common fields used to subset cellxgene data.

facets() is used to query the cellxgene database for current values of one or all facets.

facets_filter() provides a convenient way to filter facets based on label or ontology term.
Usage

FACETS

facets(cellxgene_db = db(), facets = FACETS)

facets_filter(facet, key = c("label", "ontology_term_id"), value, exact = TRUE)

Arguments

cellxgene_db an (optional) cellxgene_db object, as returned by db().

facets a character() vector corresponding to one of the facets in FACETS.

facet the column containing faceted information, e.g., sex in datasets(db).

key character(1) identifying whether value is a label or ontology_term_id.

value character() value of the label or ontology term to filter on. The value may be a vector with length(value) > 0 for exact matches (exact = TRUE, default), or a character(1) regular expression.

exact logical(1) whether values match exactly (default, TRUE) or as a regular expression (FALSE).

Format

FACETS is an object of class character of length 8.

Value

facets() returns a tibble with columns facet, label, ontology_term_id, and n, the number of times the facet label is used in the database.

facets_filter() returns a logical vector with length equal to the length (number of rows) of facet, with TRUE indicating that the value of key is present in the dataset.

Examples

f <- facets()

## levels of each facet
f |> dplyr::count(facet)

## same as facets(), facets = "organism"
f |> dplyr::filter(facet == "organism")

db <- db()
ds <- datasets(db)

## datasets with African American females
ds |> dplyr::filter(
facets_filter(self_reported_ethnicity, "label", "African American"),
facets_filter(sex, "label", "female")
)
## datasets with non-European, known ethnicity
facets(db, "self_reported_ethnicity")
ds |> dplyr::filter(
  !facets_filter(
    self_reported_ethnicity, "label", c("European", "na", "unknown")
  )
)
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