Package ‘YAPSA’

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Type Package

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Depends R (>= 4.0.0), GenomicRanges, ggplot2, grid

Description This package provides functions and routines for supervised analyses of mutational signatures (i.e., the signatures have to be known, cf. L. Alexandrov et al., Nature 2013 and L. Alexandrov et al., Bioaxiv 2018). In particular, the family of functions LCD (LCD = linear combination decomposition) can use optimal signature-specific cutoffs which takes care of different detectability of the different signatures. Moreover, the package provides different sets of mutational signatures, including the COSMIC and PCAWG SNV signatures and the PCAWG Indel signatures; the latter infering that with YAPSA, the concept of supervised analysis of mutational signatures is extended to Indel signatures. YAPSA also provides confidence intervals as computed by profile likelihoods and can perform signature analysis on a stratified mutational catalogue (SMC = stratify mutational catalogue) in order to analyze enrichment and depletion patterns for the signatures in different strata.

License GPL-3

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add_annotation

Add information to an annotation data structure

Description

Function to iteratively add information to an annotation data structure as needed for HeatmapAnnotation and especially for annotation_exposures_barplot

Usage

add_annotation(
  in_annotation_col,
  in_annotation_df,
  in_attribution_vector,
  in_colour_vector,
  in_name
)
**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>in_annotation_col</td>
<td>List, every element of which refers to one layer of annotation. List elements are structures corresponding to named colour vectors.</td>
</tr>
<tr>
<td>in_annotation_df</td>
<td>Data frame, every column of which corresponds to a layer of annotation. It has as many rows as there are samples, every entry in a row corresponding to the attribute the samples has for the corresponding layer of annotation. The factor levels of a column of in_annotation_df correspond to the names of the corresponding element in in_annotation_col.</td>
</tr>
<tr>
<td>in_attribution_vector</td>
<td>A vector which is going to be cbinded to in_annotation_df, carrying the annotation information of the new layer to be added.</td>
</tr>
<tr>
<td>in_colour_vector</td>
<td>Named vector of colours to be attributed to the new annotation.</td>
</tr>
<tr>
<td>in_name</td>
<td>Name of the new layer of annotation.</td>
</tr>
</tbody>
</table>

**Value**

A list with entries

- annotation_col: A list as in in_annotation_col but with one additional layer of annotation
- annotation_df: A data frame as in in_annotation_df but with one additional layer of annotation

**Examples**

```r
NULL
```

---

**add_as_first_to_list**  
*Add an element as first entry to a list*

**Description**

Works for all types of lists and inputs

**Usage**

```r
add_as_first_to_list(in_list, in_element)
```

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>in_list</td>
<td>List to which an element is to be added</td>
</tr>
<tr>
<td>in_element</td>
<td>Element to be added</td>
</tr>
</tbody>
</table>
aggregate_exposures_by_category

Aggregate exposures by category

Description

If a valid category (i.e. it matches to a category specified in in_sig_ind_df) is supplied, then the exposures are aggregated over this category.

Usage

aggregate_exposures_by_category(in_exposures_df, in_sig_ind_df, in_category)

Arguments

in_exposures_df
Input data frame of exposures.

in_sig_ind_df
Input data frame of meta information on the signatures. Has to match the signatures in in_exposures_df

in_category
Category to be aggregated over

Value

A list with entries:

• exposures: The exposures H, a numeric data frame with l rows and m columns, l being the number of aggregated signatures and m being the number of samples
• norm_exposures: The normalized exposures H, a numeric data frame with l rows and m columns, l being the number of aggregated signatures and m being the number of samples
• out_sig_ind_df: Data frame of the type signature_indices_df, i.e. indicating name, function and meta-information of the aggregated signatures.

See Also

LCD_complex_cutoff

Examples

NULL
annotate_intermut_dist_cohort

Annotate the intermutation distance of variants cohort-wide

Description

The function annotates the intermutational distance to a cohort wide data frame by applying `annotate_intermut_dist_PID` to every PID-specific subtraction of the cohort wide data. Note that `annotate_intermut_dist_PID` calls `rainfallTransform`. If the PID information is missing, `annotate_intermut_dist_PID` is called directly for the whole input.

Usage

```r
annotate_intermut_dist_cohort(
  in_dat,
  in_CHROM.field = "CHROM",
  in_POS.field = "POS",
  in_PID.field = NULL,
  in_mode = "min",
  in_verbose = FALSE
)
```

Arguments

- `in_dat` VRanges object, VRangesList, data frame or list of data frames which carries (at least) one column for the chromosome and one column for the position. Optionally, a column to specify the PID can be provided.
- `in_CHROM.field` String indicating which column of `in_df` carries the chromosome information
- `in_POS.field` String indicating which column of `in_df` carries the position information
- `in_PID.field` String indicating which column of `in_df` carries the PID information
- `in_mode` String passed through `annotate_intermut_dist_PID` to `rainfallTransform` indicating which method to choose for the computation of the intermutational distance.
- `in_verbose` Whether verbose or not.

Value

VRanges object, VRangesList, data frame or list of data frames identical to `in_df` (reordered by `in_PID.field`), but with the intermutation distance annotated as an additional column on the right named `dist`.

See Also

- `annotate_intermut_dist_PID`
- `rainfallTransform`
annotate_intermut_dist_PID

Annotate the intermutation distance of variants per PID

Description

The function annotates the intermutational distance to a PID wide data frame by applying `rainfallTransform` to every chromosome-specific subfraction of the PID wide data.

Usage

```r
annotate_intermut_dist_PID(
    in_dat,
    in_CHROM.field = "CHROM",
    in_POS.field = "POS",
    in_mode = "min",
    in_verbose = FALSE
)
```

Arguments

- `in_dat` VRanges object or data frame which carries (at least) one column for the chromosome and one column for the position.
- `in_CHROM.field` String indicating which column of `in_dat` carries the chromosome information if dealing with data frames.
- `in_POS.field` String indicating which column of `in_dat` carries the position information if dealing with data frames.

Examples

```r
test_df <- data.frame(CHROM=c(1,1,1,2,2,2,3,3,3,4,4,4,5,5),
                       POS=c(1,2,4,6,9,14,8,10,20,40,100,200),
                       REF=c("C","C","C","T","T","T","A",
                             "A","A","G","G","G","N","A"),
                       ALT=c("A","G","T","A","C","G","C",
                             "G","T","A","C","T","A","N"),
                       PID=c(1,1,1,2,2,2,1,1,2,2,2,1,1,2))

test_df <- test_df[order(test_df$PID,test_df$CHROM,test_df$POS),]

min_dist_df <-
    annotate_intermut_dist_cohort(test_df,in_CHROM.field="CHROM",
                                in_POS.field="POS", in_PID.field="PID",
                                in_mode="min")

max_dist_df <-
    annotate_intermut_dist_cohort(test_df,in_CHROM.field="CHROM",
                                in_POS.field="POS", in_PID.field="PID",
                                in_mode="max")

min_dist_df
max_dist_df
```
in_mode: String passed to `rainfallTransform` indicating which method to choose for the computation of the intermutational distance.

inVerbose: Whether verbose or not.

Value

`VRanges` object or data frame identical to `in_dat`, but with the intermutation distance annotated as an additional column on the right named `dist`.

See Also

- `annotate_intermut_dist_cohort`
- `rainfallTransform`

Examples

```r
test_df <- data.frame(
  CHROM=c(1,1,2,2,3,3,4,4,4,5,5),
  POS=c(1,2,4,4,6,9,1,4,8,10,20,40,100,200),
  REF=c("C","C","C","T","T","A","A","G","G","N","A"),
  ALT=c("A","G","T","A","C","G","C","G","T","A","C","T","A","N"))
min_dist_df <- annotate_intermut_dist_PID(test_df, in_CHROM.field="CHROM",
                                         in_POS.field="POS",
                                         in_mode="min")
max_dist_df <- annotate_intermut_dist_PID(test_df, in_CHROM.field="CHROM",
                                         in_POS.field="POS",
                                         in_mode="max")
min_dist_df
max_dist_df
```

---

**annotation_exposures_barplot**

*Plot the exposures of a cohort with different layers of annotation*

Description

The exposures $H$, determined by NMF or by LCD, are displayed as a stacked barplot by calling `Heatmap`. The x-axis displays the PIDs (patient identifier or sample), the y-axis the counts attributed to the different signatures with their respective colours per PID. It is analogous to `plot_exposures`. As many layers of information as desired can be added via an annotation data frame. The annotation data is handled in a way similar to `annotation_heatmap_exposures`. This function calls:

- `rowAnnotation`,
- `HeatmapAnnotation` and
- `Heatmap`
Usage

```
annotation_exposures_barplot(
    in_exposures_df,
    in_signatures_ind_df,
    in_subgroups_df,
    in_annotation_df = NULL,
    in_annotation_col = NULL,
    ylab = NULL,
    title = "",
    in_labels = FALSE,
    in_barplot_borders = TRUE,
    in_column_annoBorders = FALSE,
    in_annotation_legend_side = "right",
    in_padding = unit(c(2, 20, 2, 2), "mm"),
    in_annotation = NULL
)
```

Arguments

- `in_exposures_df`: Numerical data frame encoding the exposures H, i.e. which signature contributes how much to which PID (patient identifier or sample).
- `in_signatures_ind_df`: A data frame containing meta information about the signatures.
- `in_subgroups_df`: A data frame indicating which PID (patient or sample identifier) belongs to which subgroup.
- `in_annotation_df`: A data frame indicating which PID (patient or sample identifier) belongs to which subgroup for all layers of annotation.
- `in_annotation_col`: A list indicating colour attributions for all layers of annotation.
- `ylab`: String indicating the column name in `in_subgroups_df` to take the subgroup information from.
- `title`: Title for the plot to be created.
- `in_labels`: Whether or not to show the names of the samples.
- `in_barplot_borders`: Whether or not to show border lines in barplot.
- `in_column_annoBorders`: Whether or not to draw separating lines between the fields in the annotation.
- `in_annotation_legend_side`: Where to put the legends of the annotation df, default is right.
- `in_padding`: Parameter passed on to function `draw`.
- `in_annotation`: A full annotation object may also be provided by the educated user.
Details

It might be necessary to install the newest version of the development branch of the packages circlize and ComplexHeatmap by Zuguang Gu:

```r
devtools::install_github("jokergoo/circlize")
devtools::install_github("jokergoo/ComplexHeatmap")
```

It might be necessary to install the newest version of the development branch of the packages circlize and ComplexHeatmap by Zuguang Gu: devtools::install_github("jokergoo/circlize") and devtools::install_github("jokergoo/ComplexHeatmap")

Value

The function doesn’t return any value.

See Also

HeatmapAnnotation
Heatmap
decorate_heatmap_body
annotation_heatmap_exposures
plot_exposures

Examples

`NULL`

---

**annotation_exposures_list_barplot**

*Plot the exposures of a cohort with different layers of annotation for SNV and INDEL signatures*

---

Description

The exposures $H$, determined by NMF or by LCD, are displayed as a stacked barplot by calling Heatmap. The x-axis displays the PIDs (patient identifier or sample), the y-axis the counts attributed to the different signatures with their respective colours per PID. It is analogous to plot_exposures. As many layers of information as desired can be added via an annotation data frame. The annotation data is handled in a way similar to annotation_heatmap_exposures. In comparison to annotation_exposures_barplot allows this function to deal with a list of different signature and mutation types. This function calls:

- rowAnnotation,
- HeatmapAnnotation and
- Heatmap
annotation_exposures_list_barplot

Usage

annotation_exposures_list_barplot(
  in_exposures_list,
  in_signatures_ind_list,
  in_subgroups_list,
  in_annotation_list,
  ylab = NULL,
  title = "",
  in_indel_sigs = FALSE,
  in_labels = FALSE,
  in_barplot_borders = TRUE,
  in_column_anno_borders = FALSE,
  in_annotation_legend_side = "right",
  in_padding = unit(c(2, 20, 2, 2), "mm"),
  in_annotation = NULL
)

Arguments

in_exposures_list
  A list of numerical data frame encoding the exposures \( H \) of different signature
types, i.e. which signature contributes how much to which PID (patient identifier
or sample).
in_signatures_ind_list
  A list of data frame containing meta information about the each signature type
individually
in_subgroups_list
  A list of data frame indicating of each siganture type which PID (patient or
sample identifier) belongs to which subgroup
in_annotation_list
  A list data frame indicating which PID (patient or sample identifier) belongs to which subgroup
for all layers of annotation and a list indicating colour attribu-
tions for all layers of annotation for each signature type individually
ylab
  String indicating the column name in \( \text{in}_{\text{subgroups}} \_\text{df} \) to take the subgroup
information from.
title
  Title for the plot to be created.
in_indel_sigs
  Tag which is default FALSE when whole genome data are analysed the tag will
be TRUE
in_labels
  Whether or not to show the names of the samples.
in_barplot_borders
  Whether or not to show border lines in barplot
in_column_anno_borders
  Whether or not to draw separating lines between the fields in the annotation
in_annotation_legend_side
  Where to put the legends of the annotation df, default is right.
in_padding
  Parameter passed on to function \text{draw}
in_annotation
  A full annotation object may also be provided by the educated user.
Details

It might be necessary to install the newest version of the development branch of the packages circlize and ComplexHeatmap by Zuguang Gu:

```r
devtools::install_github("jokergoo/circlize")
and devtools::install_github("jokergoo/ComplexHeatmap")
```

It might be necessary to install the newest version of the development branch of the packages circlize and ComplexHeatmap by Zuguang Gu:

```r
devtools::install_github("jokergoo/circlize")
and devtools::install_github("jokergoo/ComplexHeatmap")
```

Value

The function doesn’t return any value.

See Also

HeatmapAnnotation
Heatmap
decorate_heatmap_body
annotation_heatmap_exposures
plot_exposures

Examples

NULL

**Description**

The PIDs are clustered according to their signature exposures. The procedure is analogous to complex_heatmap_exposures, but enabling more than one annotation row for the PIDs. This function calls:

- rowAnnotation,
- HeatmapAnnotation and
- Heatmap
annotation_heatmap_exposures

Usage

annotation_heatmap_exposures(
  in_exposures_df,
  in_annotation_df,
  in_annotation_col,
  in_signatures_ind_df,
  in_data_type = "norm exposures",
  in_method = "manhattan",
  in_palette = colorRamp2(c(0, 0.2, 0.4, 0.6), c("white", "yellow", "orange", "red")),
  in_cutoff = 0,
  in_filename = NULL,
  in_column_anno_borders = FALSE,
  in_row_anno_borders = FALSE,
  in_show_PIDs = TRUE,
  in_annotation_legend_side = "right"
)

Arguments

in_exposures_df
  Numerical data frame encoding the exposures H, i.e. which signature contributes
  how much to which PID (patient identifier or sample).

in_annotation_df
  A data frame indicating which PID (patient or sample identifier) belongs to
  which subgroup for all layers of annotation

in_annotation_col
  A list indicating colour attributions for all layers of annotation

in_signatures_ind_df
  A data frame containing meta information about the signatures, especially the
  asserted colour

in_data_type
  Title in the figure

in_method
  Method of the clustering to be supplied to dist. Can be either of: euclidean,
  maximum, manhattan, canberra, binary or minkowski

in_palette
  Palette with colours or colour codes for the heatmap. Default is colorRamp2(c(0,
  0.2, 0.4, 0.6),c(‘white’,‘yellow’,‘orange’,‘red’))

in_cutoff
  A numeric value less than 1. Signatures from within W with an overall exposure
  less than in_cutoff will be discarded for the clustering.

in_filename
  A path to save the heatmap. If none is specified, the figure will be plotted to the
  running environment.

in_column_anno_borders
  Whether or not to draw separating lines between the fields in the annotation

in_row_anno_borders
  Whether or not to draw separating lines between the fields in the annotation

in_show_PIDs
  Whether or not to show the PIDs on the x-axis

in_annotation_legend_side
  Where to put the legends of the annotation df, default is right.
attribute_nucleotide_exchanges

Details

One additional parameter, in_show_legend_bool_vector, indicating which legends to display, is planned but deactivated in this version of the package. In order to use this features, it will be necessary to install the newest version of the packages circlize and ComplexHeatmap by Zuguang Gu:
devtools::install_github("jokergoo/circlize") and devtools::install_github("jokergoo/ComplexHeatmap")

Value

The function doesn’t return any value.

See Also

Heatmap

complex_heatmap_exposures

Examples

NULL

attribute_nucleotide_exchanges

Attribute the nucleotide exchange for an SNV

Description

SNVs are grouped into 6 different categories (12/2 as reverse complements are summed over). This function defines the attribution.

Usage

attribute_nucleotide_exchanges(
  in_dat,
  in_REF.field = "REF",
  in_ALT.field = "ALT",
  in_verbose = FALSE
)

Arguments

in_dat VRanges object or data frame which carries one column for the reference base and one column for the variant base
in_REF.field String indicating which column of in_dat carries the reference base if dealing with data frames
in_ALT.field String indicating which column of in_dat carries the variant base if dealing with data frames
in_verbose Whether verbose or not.
Value

A character vector with as many rows as there are in in_dat which can be annotated (i.e. appended) to the input data frame.

Examples

test_df <- data.frame(
    CHROM=c(1,1,2,2,3,3,4,4,4,5,5),
    POS=c(1,2,3,4,5,6,1,2,3,4,5,6,7,8),
    REF=c("C","C","C","T","T","T","A","A","A","G","G","G","N","A"),

```r
test_df$change <- attribute_nucleotide_exchanges(
    test_df,in_REF.field = "REF",in_ALT.field = "ALT")
```

```r
test_df
```

attribute_sequence_contex_indel

*Attribution of sequence context and size for an INDEL*

Description

The function is a wrapper and uses `getSequenceContext` to annotate the sequence context.

Usage

```r
attribute_sequence_contex_indel(
    in_dat,
    in_REF.field = "REF",
    in_ALT.field = "ALT",
    in_verbose = FALSE,
    in_offsetL = 10,
    in_offsetR = 50
)
```

Arguments

- **in_dat**: VRanges object or data frame which carries one column for the reference base and one column for the variant base.
- **in_REF.field**: String indicating which column of in_dat carries the reference base if dealing with data frames.
- **in_ALT.field**: String indicating which column of in_dat carries the variant base if dealing with data frames.
- **in_verbose**: Verbose if in_verbose=1.
- **in_offsetL**: Number of nucleotides which should be annotated downstream of the variant. Per default 10 bps are annotated.
- **in_offsetR**: Number of nucleotides which should be annotated upstream of the variant. Per default 50 bps are annotated.
**Value**

VRanges object or data frame with the same number rows and additional columns containing the type of INDEL (Ins = insertion and Del = deletion), the annotated sequence context of the defined length, the absolute number of exchanged nucleotides and the nucleotide exchange between `in_REF.field` and `in_ALT.field`.

**Examples**

```r
data(GenomeOfNl_raw)
GenomeOfNl_context <- attribute_sequence_contex_indel(
  in_dat = head(GenomeOfNl_raw),
  in_REF.field = "REF",
  in_ALT.field = "ALT",
  in_verbose = FALSE,
  in_offsetL = 10, in_offsetR = 50)
GenomeOfNl_context
```

**Description**

Each variant is categorized into one of the 83 INDEL categories. The classification likewise to Alexandrov et al., 2018 (https://www.synapse.org/#!Synapse:syn11726616). The number of 83 features are classified as follows:

1. Deletion of 1 bp C/(G) or T/(A) in a repetitive context. The context is classified into 1, 2, 3, 4, 5 or larger or equal to 6 times the same nucleotide(s).
2. Insertion of 1 bp C/(G) or T/(A) in a repetitive context. The context is classified into 0, 1, 2, 3, 4, or larger or equal to 5 times the same nucleotide(s).
3. Deletions of 2bps, 3bps, 4bps or more or equal to 5bps in a repetitive context. Each deletion is classified in a context of 1, 2, 3, 4, 5 or larger or equal to 6 times the same motif.
4. Insertion of 2 bps, 3 bps, 4 bps or more or equal to 5 bps in a repetitive context. Each deletion is classified in a context of 0, 1, 2, 3, 4 or larger or equal to 5 times the same motif.
5. Microhomology deletion of 2bps, 3bps, 4bps or more or equal to 5 bps in a partly repetitive context. The partly repetitive context is defined by motif length of minus 1 bp, 2 bps, 3 bps, 4 bps or more or equal to 5bps, which is located before and after the break-point junction of the deletion.

**Usage**

```r
attribution_of_indels(in_dat_return = in_dat_return)
```
**Arguments**

in_dat_return  Data frame constructed from a vcf-like file of a whole cohort or a single-sample. The first columns are those of a standard vcf file, followed by an arbitrary number of custom or defined columns. One of these can carry a PID (patient or sample identifier) and subgroup information. Furthermore, the columns containing the sequence context and the absolute length of the INDEL as well as the INDEL type of the variant can be annotated to the vcf-like df with `attribute_sequence_contex_indel`. These columns are required to enable the construction of a mutational catalog.

**Value**

Data frame with the same dimension as the input data frame plus an additional column with the INDEL classification number corresponding to Alexandrov et al. 2018.

**Examples**

```r
data(GenomeOfNl_raw)
GenomeOfNl_context <- attribute_sequence_contex_indel(in_dat = head(GenomeOfNl_raw))
GenomeOfNl_classified <- attribution_of_indels(GenomeOfNl_context)
GenomeOfNl_classified
```

---

**build_gene_list_for_pathway**

*Build a gene list for a given pathway name*

**Description**

Build a gene list for a given pathway name

**Usage**

```r
build_gene_list_for_pathway(in_string, in_organism)
```

**Arguments**

in_string  Name or description of the pathway

in_organism  Name of the taxon to be searched in

**Value**

A character vector of gene names
classify_indels

INDEL function V1 - not compatible with AlexandrovSignatures

Description

INDEL function V1 - not compatible with AlexandrovSignatures

Usage

classify_indels(
  in_df,
  in_ALT.field = "ALT",
  in_REF.field = "REF",
  in_breaks = c(-Inf, -10, -3, 0, 2, 9, Inf),
  in_labels = c("del3", "del2", "del1", "in1", "in2", "in3")
)

Arguments

  in_df       Input data frame containing the variances in a vcf-like format
  in_ALT.field Column number for alternative field
  in_REF.field Column number for reference field
  in_breaks   Handed over to function cut
  in_labels   Handed over to function cut
compare_exposures

Value
classVector, a factor vector of indel sizes

Examples
NULL

compare_exposures  Compares alternative exposures

Description
Compares exposures computed by two alternative approaches for the same cohort

Usage
compare_exposures(in_exposures1_df, in_exposures2_df, deselect_flag = TRUE)

Arguments
in_exposures1_df  Numeric data frame with exposures, ideally the smaller exposure data is supplied first.
in_exposures2_df  Numeric data frame with exposures, ideally the bigger exposure data is supplied second.
deselect_flag     Whether signatures absent in both exposure data frames should be removed.

Value
A list with entries merge_df, all_cor.coeff, all_p.value, cor.coeff_vector, p.value_vector, all_cor.test, and cor.test_list.
- merge_df: Merged molten input exposure data frames
- all_cor.coeff: Pearson correlation coefficient for all data points, i.e. taken all signatures together
- all_p.value: P-value of the Pearson test for all data points, i.e. taken all signatures together
- cor.coeff_vector: A vector of Pearson correlation coefficients evaluated for every signature independently
- p.value_vector: A vector of p-values of the Pearson tests evaluated for every signature independently
- all_cor.test: A data structure as returned by cor.test for all data points, i.e. taken all signatures together
- cor.test_list: A list of data structures as returned by cor.test, but evaluated for every signature independently
**compare_expousre_sets**  

**Examples**  

NULL

---

**compare_expousre_sets**  
*Compare two sets of exposures by cosine distance*

**Description**

Compare two sets of exposures, stored in numerical data frames \( H_1 \) and \( H_2 \), by computing the row-wise cosine distance

**Usage**

```r
compare_expousre_sets(in_df_small, in_df_big, in_distance = cosineDist)
```

**Arguments**

- `in_df_small`, `in_df_big`
  
  Numerical data frames \( H_1 \) and \( H_2 \), ideally the bigger one first, both with \( l \) rows and \( m_1 \) and \( m_2 \) columns, \( l \) being the number of signatures and \( m_1 \) and \( m_2 \) being the respective numbers of samples or patient identifier of \( H_1 \) and \( H_2 \)

- `in_distance`
  
  A function which computes the distance measure, default is `cosineDist`

**Value**

A list with entries `distance`, `hierarchy_small` and `hierarchy_big`.

- **distance**: A numerical data frame with the cosine distances between the columns of \( H_1 \), indexing the rows, and \( H_2 \), indexing the columns
- **hierarchy_small**: A data frame carrying the information of ranked similarity between the signatures in \( H_2 \) with the signatures in \( H_1 \)
- **hierarchy_big**: A data frame carrying the information of ranked similarity between the signatures in \( H_1 \) with the signatures in \( H_2 \)

**See Also**

- `cosineDist`

**Examples**

```r
sig_1_df <- data.frame(matrix(c(1,0,0,0,1,0,0,0,0,1,0),ncol=3))
names(sig_1_df) <- paste0("B",seq_len(dim(sig_1_df)[2]))
sig_2_df <- data.frame(matrix(c(1,1,0,0,0,0,1,1),ncol=2))
compare_expousre_sets(sig_1_df,sig_2_df)
```
compare_sets

Compare two sets of signatures by cosine distance

Description

Compare two sets of signatures, stored in numerical data frames \(W_1\) and \(W_2\), by computing the column-wise cosine distance.

Usage

\[
\text{compare_sets}(\text{in\_df\_small}, \text{in\_df\_big}, \text{in\_distance} = \text{cosineDist})
\]

Arguments

\text{in\_df\_small}, \text{in\_df\_big}

Numerical data frames \(W_1\) and \(W_2\), ideally the bigger one first, both with \(n\) rows and \(l_1\) and \(l_2\) columns, \(n\) being the number of features and \(l_1\) and \(l_2\) being the respective numbers of signatures of \(W_1\) and \(W_2\).

\text{in\_distance}

A function which computes the distance measure, default is \text{cosineDist}.

Value

A list with entries distance, hierarchy_small and hierarchy_big.

- distance: A numerical data frame with the cosine distances between the columns of \(W_1\), indexing the rows, and \(W_2\), indexing the columns.
- hierarchy_small: A data frame carrying the information of ranked similarity between the signatures in \(W_2\) with the signatures in \(W_1\).
- hierarchy_big: A data frame carrying the information of ranked similarity between the signatures in \(W_1\) with the signatures in \(W_2\).

See Also

\text{cosineDist}

Examples

\[
\begin{align*}
\text{sig\_1\_df} & \leftarrow \text{data.frame(matrix(c(1,0,0,0,1,0,0,0,1,0),ncol=3))} \\
\text{names(sig\_1\_df)} & \leftarrow \text{paste0("B",seq_len(dim(six\_of\_df)[2]))} \\
\text{sig\_2\_df} & \leftarrow \text{data.frame(matrix(c(1,1,0,0,0,1,1),ncol=2))} \\
\text{compare\_sets(six\_of\_df,six\_of\_df)}
\end{align*}
\]
compare_SMCS

Compare all strata from different stratifications

Description

Compare all strata from different orthogonal stratification axes, i.e. orthogonal SMCs by cosine similarity of signature exposures. First calls

- `make_strata_df`, then
- `plot_strata` and finally
- `make_comparison_matrix`

Usage

```r
compare_SMCS(
  in_stratification_lists_list,
  in_signatures_ind_df,
  output_path,
  in_nrect = 5,
  in_attribute = ""
)
```

Arguments

- `in_stratification_lists_list` List of lists with entries from different (orthogonal) stratification axes or SMCs
- `in_signatures_ind_df` A data frame containing meta information about the signatures
- `output_path` Path to directory where the results, especially the figure produced by `corrplot` is going to be stored.
- `in_nrect` Number of clusters in the clustering procedure provided by `corrplot`
- `in_attribute` Additional string for the file name where the figure produced by `corrplot` is going to be stored.

Value

The comparison matrix of cosine similarities.

See Also

- `plot_strata`
- `make_comparison_matrix`

Examples

NULL
**compare_to_catalogues**  
*Compare one mutational catalogue to reference mutational catalogues*

**Description**

Compare one mutational catalogue (e.g. of one index patient) to a list of reference mutational catalogues (e.g. from the initial Alexandrov publication) by cosine similarities

**Usage**

`compare_to_catalogues(in_index_df, in_comparison_list)`

**Arguments**

- `in_index_df` Data frame containing the mutational catalogue of interest
- `in_comparison_list` List of data frames (ideally named) containing the reference mutational catalogues

**Value**

A similarity dataframe

**Examples**

NULL

**complex_heatmap_exposures**  
*Heatmap to cluster the PIDs on their signature exposures (Complex-Heatmap)*

**Description**

The PIDs are clustered according to their signature exposures. uses package ComplexHeatmap by Zuguang Gu. This function calls:

- `rowAnnotation`,
- `HeatmapAnnotation` and
- `Heatmap`
Usage

complex_heatmap_exposures(
  in_exposures_df,
  in_subgroups_df,
  in_signatures_ind_df,
  in_data_type = "norm exposures",
  in_method = "manhattan",
  in_subgroup_column = "subgroup",
  in_subgroup_colour_column = NULL,
  in_palette = colorRamp2(c(0, 0.2, 0.4, 0.6), c("white", "yellow", "orange", "red")),
  in_cutoff = 0,
  in_filename = NULL,
  in_column_anno_borders = FALSE,
  in_row_anno_borders = FALSE
)

Arguments

in_exposures_df
  Numerical data frame encoding the exposures H, i.e. which signature contributes how much to which PID (patient identifier or sample).

in_subgroups_df
  A data frame indicating which PID (patient or sample identifier) belongs to which subgroup

in_signatures_ind_df
  A data frame containing meta information about the signatures, especially the asserted colour

in_data_type
  Title in the figure

in_method
  Method of the clustering to be supplied to dist. Can be either of: euclidean, maximum, manhattan, canberra, binary or minkowski

in_subgroup_column
  Indicates the name of the column in which the subgroup information is encoded in in_subgroups_df

in_subgroup_colour_column
  Indicates the name of the column in which the colour information for subgroups is encoded in in_subgroups_df. If NULL, a rainbow palette is used instead.

in_palette
  Palette with colours for the heatmap. Default is colorRamp2(c(0, 0.2, 0.4, 0.6), c('white','yellow','orange','red'))

in_cutoff
  A numeric value less than 1. Signatures from within W with an overall exposure less than in_cutoff will be discarded for the clustering.

in_filename
  A path to save the heatmap. If none is specified, the figure will be plotted to the running environment.

in_column_anno_borders
  Whether or not to draw separating lines between the fields in the annotation

in_row_anno_borders
  Whether or not to draw separating lines between the fields in the annotation
Details

It might be necessary to install the newest version of the development branch of the packages \texttt{circlize} and \texttt{ComplexHeatmap} by Zuguang Gu: \texttt{devtools::install_github("jokergoo/circlize")} and \texttt{devtools::install_github("jokergoo/ComplexHeatmap")}

Value

The function doesn’t return any value.

See Also

\texttt{Heatmap}

Examples

\begin{verbatim}
data(lymphoma_cohort_LCD_results) complex_heatmap_exposures( rel_lymphoma_Nature2013_COSMIC_cutoff_exposures_df, COSMIC_subgroups_df, chosen_signatures_indices_df, in_data_type="norm exposures", in_subgroup_colour_column="col", in_method="manhattan", in_subgroup_column="subgroup")
\end{verbatim}

\begin{verbatim}
computeLogLik  Compute the loglikelihood
\end{verbatim}

Description

Compute the loglikelihood

Usage

\texttt{computeLogLik(in_vector, in_pdf = NULL, verbose = FALSE)}

Arguments

- \texttt{in_vector} Numeric vector of input values of which the loglikelihood is computed.
- \texttt{in_pdf} Probability distribution function, if NULL a normal distribution is used.
- \texttt{verbose} Verbose if \texttt{in_verbose}=1

Value

A numeric value (sum of the logarithms of the likelihoods of the input vector)
compute_comparison_stat_df

Extract statistical measures for entity comparison

Description

Compare one mutational catalogue (e.g. of one index patient) to a list of reference mutational catalogues (e.g. from the initial Alexandrov publication) by cosine similarities

Usage

compute_comparison_stat_df(in_sim_df)

Arguments

in_sim_df A similarity data frame as extracted by compare_to_catalogues

Value

A dataframe containing statistical measures, prepared for bar plot

Examples

NULL

compute_comparison_stat_df

confidence_indel_calulation

Wrapper to compute confidence intervals for SNV and INDEL signatures of a cohort or single-sample

Description

Wrapper function around confIntExp, which applies to every signature or sample pair in a cohort. The extracted lower bound of the confidence intervals are added to the input data which is reordered and melted in order to prepare for visualization with ggplot2. The calculates of confidence intervals is based on a profiling likelihood algorithm and the wrapper calculates the data for the exposure contubution identefied with SNV and INDEL signature decompositions and application of the following cutoffs:

1. CosmicValid_absCutoffVector
2. CosmicValid_normCutoffVector
3. CosmicArtif_absCutoffVector
4. CosmicArtif_normCutoffVector
5. PCAWGValidSNV_absCutoffVector
6. PCAWGValidID_absCutoffVector

The function makes use of different YAPSA functions. For each of the above stated cutoff vectors a per PID decomposition of the SNV and INDEL catalog is calculated respectively using `LCD_complex_cutoff_perPID`. In a next step, `variateExp` which is a wrapper around `confIntExp` to compute confidence intervals for a cohort is used. A dataframe is returned with the upper and lower bounds of the confidence intervals. In a last step `plotExposuresConfidence_indel` to plot the exposures to extracted signatures including confidence intervals computed with e.g. by `variateExp`.

**Usage**

```r
confidence_indel_calulation(in_current_indel_df, in_current_snv_df)
```

**Arguments**

- `in_current_indel_df`
  A INDEL mutational catalog. Mutational catalog can be constructed with `create_indel_mutation_catalogue_from_df`
- `in_current_snv_df`
  A SNV mutational catalog. Mutational catalog can be constructed with `create_mutation_catalogue_from_df`

**Value**

A list is returned containing 12 objects. For each cutoff data frame two corresponding object are present. First, the `p gtabelle` object which can be used for graphically visualisation, and second a dataframe containing the corresponding upper and lower bounds of the confidence intervals.

**Examples**

```r
data("GenomeOfNl_MutCat")
```

---

**confidence_indel_only_calulation**

*Wrapper to compute confidence intervals for only INDEL signatures.*

**Description**

Wrapper function around `confIntExp`, which is applies to every signature or sample pair in a cohort. The extracted lower bound of the confidence intervals are added to the input data which is reordered and melted in order to prepare for visualization with `ggplot2`. The calculates of confidence intervals is based on a profiling likelihood algorithm and the wrapper calculates the data for the exposure contribution identified with INDEL signature decomposition and the usage of `PCAWGValidID_absCutoffVector` data frame.
confIntExp

Usage

    confidence_indel_only_calculation(in_current_indel_df)

Arguments

    in_current_indel_df
    A INDEL mutational catalog. Mutational catalog can be constructed with `create_indel_mutation_catalogue_from_df`.

Details

    The function makes use of different YAPSA functions. For each of the above stated cutoff vectors a per PID decomposition of the SNV and INDEL catalog is calculated respectively using `LCD_complex_cutoff_perPID`. In a next step, `variateExp` which is a wrapper around `confIntExp` to compute confidence intervals for a cohort is used. A dataframe is returned with the upper and lower bounds of the confidence intervals. In a last step `plotExposuresConfidence_indel` to plot the exposures to extracted signatures including confidence intervals computed with e.g. `variateExp`.

Value

    A list is returned containing two objects. First, the `pgtable` object which can be used for graphically visualization, and second a dataframe containing the corresponding upper and lower bounds of the confidence intervals.

Examples

    data("GenomeOfNl_MutCat")
    temp_list <- confidence_indel_only_calculation(
        in_current_indel_df=MutCat_indel_df)
    plot(temp_list$p_complete_PCAWG_ID)
    head(temp_list$complete_PCAWG_ID)

---

confIntExp  
*Compute confidence intervals*

Description

    Compute confidence intervals using the (log-)likelihood ratio test, primarily for one input sample.

Usage

    confIntExp(
        in_ind = 1,
        in_sigLevel = 0.05,
        in_delta = 1,
        in_exposure_vector = NULL,
        in_verbose = FALSE,
        ...
    )
Arguments

in_ind    Index of the input signature to be variated.
in_sigLevel    Significance leve (one-sided)
in_delta    Inflation parameter for the alternative model.
in_exposure_vector    Exposure vector computed for the input sample.
in_verbose    Whether to run verbose (TRUE) or not (FALSE)
...

Input parameters passed on to variateExpSingle.

Value

A list with entries

- upper: Upper bound of the confidence interval
- lower: Lower bound of the confidence interval

Examples

library(BSgenome.Hsapiens.UCSC.hg19)
data(lymphoma_test)
data(lymphoma_cohort_LCD_results)
data(sigs)
word_length <- 3
temp_list <- create_mutation_catalogue_from_df(
  lymphoma_test_df,
  this_seqnames.field = "CHROM",
  this_start.field = "POS",
  this_end.field = "POS",
  this_PID.field = "PID",
  this_subgroup.field = "SUBGROUP",
  this_refGenome = BSgenome.Hsapiens.UCSC.hg19,
  this_wordLength = word_length)
lymphoma_catalogue_df <- temp_list$matrix
lymphoma_PIDs <- colnames(lymphoma_catalogue_df)
data("lymphoma_cohort_LCD_results")
lymphoma_exposures_df <-
  lymphoma_Nature2013_COSMIC_cutoff_exposures_df[, lymphoma_PIDs]
lymphoma_sigs <- rownames(lymphoma_exposures_df)
lymphoma_sig_df <- AlexCosmicValid_sig_df[, lymphoma_sigs]
confIntExp(in_ind = 1, in_sigLevel = 0.05, in_delta = 0.4,
in_exposure_vector = lymphoma_exposures_df[, 1],
in_catalogue_vector = lymphoma_catalogue_df[, 1],
in_sig_df = lymphoma_sig_df)
correct_rounded

Readjust the vector to its original norm after rounding

Description

After use of the function `round_precision` the norm of the input vector may have been altered by the rounding procedure. This function restores the norm by altering only the largest entry in the rounded vector (in order to create the least possible relative error).

Usage

```r
correct_rounded(x, in_interval = c(0, 1))
```

Arguments

- `x`: vector to be rounded
- `in_interval`: Interval

Value

The adapted form of the input vector `x`.

Examples

```r
NULL
```

cosineDist

Compute the cosine distance of two vectors

Description

Compute the cosine distance of two vectors

Usage

```r
cosineDist(a, b)
```

Arguments

- `a`, `b`: Numerical vectors of same length

Value

The scalar product of the two input vectors divided by the product of the norms of the two input vectors
Examples

```r
## 1. Orthogonal vectors:
cosineMatchDist(c(1,0),c(0,1))
## 2. Non-orthogonal vectors:
  cosineDist(c(1,0),c(1,1))
## Compare trigonometry:
  1-cos(pi/4)
```

---

**cosineMatchDist**

*Compute an altered cosine distance of two vectors*

Description

This is an altered cosine distance: it first reduced the dimension of the two input vectors to only those coordinates where both have non-zero entries. The cosine similarity is then computed on these reduced vectors, i.e. on a sub-vector space.

Usage

```r
cosineMatchDist(a, b)
```

Arguments

- `a, b` Numerical vectors of same length

Value

The scalar product of the reduced input vectors divided by the product of the norms of the two reduced input vectors

Examples

```r
## 1. Orthogonal vectors:
cosineMatchDist(c(1,0),c(0,1))
## 2. Non-orthogonal vectors:
cosineMatchDist(c(1,0),c(1,1))
```
**create_indel_mutation_catalogue_from_df**

Wrapper to create an INDEL mutational catalog from a vlf-like data frame

**Description**

From data frame constructed from a vcf-file file the function `create_indel_mutation_catalogue_from_df` creates a mutational catalog V by sequentially applying the `attribute_sequence_contex_indel`, `attribute_sequence_contex_indel` and then `attribute_of_indels`. The runtime of the function is about 1 sec per 6 variants as sequence context as well as INDEL classification are timeconsuming to compute (optimization ongoing)

**Usage**

```r
create_indel_mutation_catalogue_from_df(
  in_dat, 
  in_signature_df, 
  in_REF.field = "REF", 
  in_ALT.field = "ALT", 
  in_verbose = FALSE
)
```

**Arguments**

- **in_dat**
  A data frame constructed from a vcf-like file of a whole cohort or single-sample. The first columns are those of a standard vcf file (CHROM, POS, REF and ALT), followed by an arbitrary number of custom or used defined columns. One of these can carry a PID (patient or sample identifyier) and one can carry subgroup information.

- **in_signature_df**
  A numeric data frame W with n rows and l columns, n being the number of features and l being the number od signatures. Data frame containing INDEL signatures which should be used to create the mutational cataloge V.

- **in_REF.field**
  String indicating which column of in_dat carries the reference base if dealing with data frames

- **in_ALT.field**
  String indicating which column of in_dat carries the variant base if dealing with data frames

- **in_verbose**
  Verbose if in_verbose=1

**Value**

A dataframe in the format of a mutational catalog V, which can be used for LCD analysis
create_indel_mut_cat_from_df

Create a Mutational catalog from a data frame

Description

This function creates a mutational catalog from a data frame. It requires the returned data frame obtained with attribution_of_indels.

Usage

create_indel_mut_cat_from_df(in_df, in_signatures_df)

Arguments

in_df A data frame constructed from a vcf-like file of a whole cohort or single-sample. The first columns are those of a standard vcf file, followed by an arbitrary number of custom or used defined columns. One of these can carry a PID (patient or sample identifier) and the subgroup information. Additionaly to construct the mutational catalog each variant needs to be characterized into one of the 83 INDEL feature classes, which can be performed with attribution_of_indels

in_signatures_df A numeric data frame with \( n \) rows and \( l \) columns, \( n \) being the number of features and \( l \) being the number of signatures. Data frame containing INDEL signatures which should be used to create the mutational catalog \( V \).

Value

A count dataframe, the mutational catalog \( V \) with rownames indicating the INDELS and colnames having the PIDs.
create_mutation_catalogue_from_df

Create a Mutational Catalogue from a data frame

Description
This function creates a mutational catalogue from a data frame. It is a wrapper function for create_mutation_catalogue_from_VR: it first creates a VRanges object from the data frame by makeVRangesFromDataFrame and then passes this object on to the above mentioned custom function.

Usage
create_mutation_catalogue_from_df(this_df,
   this_refGenome_Seqinfo = NULL,
   this_seqnames.field = "X.CHROM",
   this_start.field = "POS",
   this_end.field = "POS",
   this_PID.field = "PID",
   this_subgroup.field = "subgroup",
   this_refGenome,
   this_wordLength,
   this_verbose = 1,
   this_rownames = c(),
   this_adapt_rownames = 1)

Arguments
this_df A data frame constructed from a vcf-like file of a whole cohort. The first columns are those of a standard vcf file, followed by an arbitrary number of custom or used defined columns. One of these can carry a PID (patient or sample identifier) and one can carry subgroup information.
this_refGenome_Seqinfo A seqInfo object, referring to the reference genome used. Argument passed on to makeGRangesFromDataFrame and thus indirectly to makeGRangesFromDataFrame.

Examples
data(GenomeOfNl_raw)
data(sigs_pcawg)
GenomeOfNl_context <- attribute_sequence_contex_indel(in_dat = head(GenomeOfNl_raw))
GenomeOfNl_classified <- attribution_of_indels(GenomeOfNl_context)
GenomeOfNl_mut_cat <- create_indel_mut_cat_from_df(GenomeOfNl_classified,
in_signatures_df=PCAWG_SP_ID_sigs_df)
create_mutation_catalogue_from_df

this_seqnames.field
Indicates the name of the column in which the chromosome is encoded

this_start.field
Indicates the name of the column in which the start coordinate is encoded

this_end.field
Indicates the name of the column in which the end coordinate is encoded

this_PID.field
Indicates the name of the column in which the PID (patient or sample identifier) is encoded

this_subgroup.field
Indicates the name of the column in which the subgroup information is encoded

this_refGenome
The reference genome handed over to create_mutation_catalogue_from_VR and indirectly to mutationContext and used to extract the motif context of the variants in in_vr.

this_wordLength
The size of the motifs to be extracted by mutationContext

this_verbose
Verbose if this.verbose=1

this_rownames
Optional parameter to specify rownames of the mutational catalogue V i.e. the names of the features.

this_adapt_rownames
Rownames of the output matrix will be adapted if this.adapt_rownames=1

Value
A list with entries matrix and frame obtained from create_mutation_catalogue_from_VR:

- matrix: The mutational catalogue V
- frame: Additional and meta information on rownames (features), colnames (PIDs) and subgroup attribution.

See Also
makeVRangesFromDataFrame
create_mutation_catalogue_from_VR

Examples
library(BSgenome.Hsapiens.UCSC.hg19)
data(lymphoma_test)
word_length <- 3
temp_list <- create_mutation_catalogue_from_df(  lymphoma_test_df,this_seqnames.field = "CHROM",  this_start.field = "POS",this_end.field = "POS",  this_PID.field = "PID",this_subgroup.field = "SUBGROUP",  this_refGenome = BSgenome.Hsapiens.UCSC.hg19,  this_wordLength = word_length)
dim(temp_list$matrix)
head(temp_list$matrix)
create_mutation_catalogue_from_VR

Create a Mutational Catalogue from a VRanges Object

Description

This function creates a mutational catalogue from a VRanges Object by first calling `mutationContext` to establish the motif context of the variants in the input VRanges and then calling `motifMatrix` to build the mutational catalogue V.

Usage

```r
create_mutation_catalogue_from_VR(
  in_vr, 
  in_refGenome, 
  in_wordLength, 
  in_PID.field = "PID", 
  in_verbose = 0, 
  in_rownames = c(), 
  adapt_rownames = 1
)
```

Arguments

- **in_vr**: A VRanges object constructed from a vcf-like file of a whole cohort. The first columns are those of a standard vcf file, followed by an arbitrary number of custom or used defined columns. One of these can carry a PID (patient or sample identifier) and one can carry subgroup information.
- **in_refGenome**: The reference genome handed over to `mutationContext` and used to extract the motif context of the variants in `in_vr`.
- **in_wordLength**: The size of the motifs to be extracted by `mutationContext`.
- **in_PID.field**: Indicates the name of the column in which the PID (patient or sample identifier) is encoded.
- **in_verbose**: Verbose if `in_verbose=1`.
- **in_rownames**: Optional parameter to specify rownames of the mutational catalogue V i.e. the names of the features.
- **adapt_rownames**: Rownames of the output matrix will be adapted if `adapt_rownames=1`.

Value

A list with entries `matrix`, `frame`,

- **matrix**: The mutational catalogue V
- **frame**: Additional and meta information on rownames (features), colnames (PIDs) and subgroup attribution.
cutoffs

See Also

mutationContext
motifMatrix

Examples

```r
library(BSgenome.Hsapiens.UCSC.hg19)
data(lymphoma_test)
data(sigs)
word_length <- 3
temp_vr <- makeVRangesFromDataFrame(
  lymphoma_test_df,in_seqnames.field="CHROM",
  in_subgroup.field="SUBGROUP",verbose_flag=1)
temp_list <- create_mutation_catalogue_from_VR(
  temp_vr,in_refGenome=BSgenome.Hsapiens.UCSC.hg19,
  in_wordLength=word_length,in_PID.field="PID",
  in_verbose=1)
dim(temp_list$matrix)
head(temp_list$matrix)
test_list <- split(lymphoma_test_df,f=lymphoma_test_df$PID)
other_list <- list()
for(i in seq_len(length(test_list))){
  other_list[[i]] <- test_list[[i]][c(1:80),]
}
other_df <- do.call(rbind,other_list)
other_vr <- makeVRangesFromDataFrame(
  other_df,in_seqnames.field="CHROM",
  in_subgroup.field="SUBGROUP",verbose_flag=1)
other_list <- create_mutation_catalogue_from_VR(
  other_vr,in_refGenome=BSgenome.Hsapiens.UCSC.hg19,
  in_wordLength=word_length,in_PID.field="PID",
  in_verbose=1,in_rownames=rownames(AlexCosmicValid_sig_df))
dim(other_list$matrix)
head(other_list$matrix)
```

cutoffs

Cutoffs for a supervised analysis of mutational signatures.

Description

Series of data frames with signature-specific cutoffs. All values represent optimal cutoffs. The optimal cutoffs were determined for different choices of parameters in the cost function of the optimization. The row index is equivalent to the ratio between costs for false negative attribution and false positive attribution. The columns correspond to the different signatures. To be used with `LCD_complex_cutoff`. There are two different sets of cutoffs one for the signatures described by Alexandrov et al.(Natue 2013) and one for the signatures dokumented in Alexandriv et al. (biorxiv 2018). The calculation of the PCAWG signature specific cutoffs was perfomed in a single-sample resolution which are both valid for whole genome and whole exome sequencing data analysis.
cutoffCosmicValid_rel_df: Optimal cutoffs for AlexCosmicValid_sig_df, i.e. COSMIC signatures, only validated, trained on relative exposures.
cutoffCosmicArtif_rel_df: Optimal cutoffs for AlexCosmicArtif_sig_df, i.e. COSMIC signatures, including artifact signatures, trained on relative exposures.
cutoffCosmicValid_abs_df: Optimal cutoffs for AlexCosmicValid_sig_df, i.e. COSMIC signatures, only validated, trained on absolute exposures.
cutoffCosmicArtif_abs_df: Optimal cutoffs for AlexCosmicArtif_sig_df, i.e. COSMIC signatures, including artifact signatures, trained on absolute exposures.
cutoffInitialValid_rel_df: Optimal cutoffs for AlexInitialValid_sig_df, i.e. initially published signatures, only validated signatures, trained on relative exposures.
cutoffInitialArtif_rel_df: Optimal cutoffs for AlexInitialArtif_sig_df, i.e. initially published signatures, including artifact signatures, trained on relative exposures.
cutoffInitialValid_abs_df: Optimal cutoffs for AlexInitialValid_sig_df, i.e. initially published signatures, only validated signatures, trained on absolute exposures.
cutoffInitialArtif_abs_df: Optimal cutoffs for AlexInitialArtif_sig_df, i.e. initially published signatures, including artifact signatures, trained on absolute exposures.

Usage

data(cutoffs)

Author(s)

Daniel Huebschmann <huebschmann.daniel@googlemail.com>

---

cutoffs_pcawg

| Opt. cutoffs, PCAWG SNV signatures, including artifacts |

Description

cutoffPCAWG_SBS_WGSWES_artifPid_df: Optimal cutoffs for PCAWG_SP_SBS_sigs_Artif_df, i.e. initially published signatures, including artifact signatures, trained in a single-sample resolution.
cutoffPCAWG_SBS_WGSWES_realPid_df: Optimal cutoffs for PCAWG_SP_SBS_sigs_Real_df, i.e. initially published signatures, only validated signatures, trained in a single-sample resolution.
cutoffPCAWG_ID_WGS_Pid_df: Optimal cutoffs for PCAWG_SP_ID_sigs_df, i.e. initially published signatures, signatures, trained in a single-sample resolution.

Usage

data(cutoffs_pcawg)

Author(s)

Lea Jopp-Saile <huebschmann.daniel@googlemail.com>
cut_breaks_as_intervals

*Wrapper for cut*

**Description**

In this wrapper function for the known `cut` function, the breaks vector need not be supplied directly, instead, for every break, an interval is supplied and the function optimizes the choice of the breakpoint by choosing a local minimum of the distribution.

**Usage**

```r
cut_breaks_as_intervals(
  in_vector,
  in_outlier_cutoffs = c(0, 3000),
  in_cutoff_ranges_list = list(c(60, 69), c(25, 32)),
  in_labels = c("late", "intermediate", "early"),
  in_name = "",
  output_path = NULL
)
```

**Arguments**

- `in_vector` Vector of numerical continuously distributed input
- `in_outlier_cutoffs` Interval specifying the upper and lower bounds of the range to be considered
- `in_cutoff_ranges_list` List of intervals in which the cutoffs for `cut` have to be optimized.
- `in_labels` Labels assigned to the strata or factors returned
- `in_name` String specifying the name of the quantity analyzed (and plotted on the x-axis of the figure to be created).
- `output_path` Path where the figure produced by the density function should be stored if non-NULL.

**Value**

A list with entries `category_vector`, `density_plot` and `cutoffs`

- `category_vector`: Factor vector of the categories or strata, of the same length as `in_vector`
- `density_plot`: Density plot produced by the density function and indication of the chosen cutoffs.
- `cutoffs`: Vector of the computed optimal cutoffs

**See Also**

- `cut`
- `density`
deriveSigInd_df

Examples

```r
data(lymphoma_test)
lymphoma_test_df$random_norm <- rnorm(dim(lymphoma_test_df)[1])
temp_list <- cut_breaks_as_intervals(
  lymphoma_test_df$random_norm,
  in_outlier_cutoffs=c(-4,4),
  in_cutoff_ranges_list=list(c(-2.5,-1.5),c(0.5,1.5)),
  in_labels=c("small","intermediate","big"))
temp_list$density_plot
```

---

**deriveSigInd_df**

**Derive a signature_indices_df object**

### Description

Derive a data frame of type signature_indices_df (additional information for a set of signatures) from a set of given signatures for a set of new signatures.

### Usage

```r
deriveSigInd_df(querySigs, subjectSigs, querySigInd = NULL, in_sort = FALSE)
```

### Arguments

- **querySigs**: The signatures to compare to (given signatures).
- **subjectSigs**: The signatures to be compared (new signatures). Alternatively this may be a complex object of type list and contain data from different deconvolutions, each of which having a set of signature to be compared.
- **querySigInd**: The object of type signature_indices_df (additional information for a set of signatures) belonging to the set of known signatures.
- **in_sort**: Whether to sort or not

### Value

An object of type signature_indices_df (additional information for a set of signatures) belonging to the set of new signatures.

### See Also

relateSigs

### Examples

```r
NULL
```
disambiguateVector  Disambiguate a vector

Description
Add numbered suffixes to redundant entries in a vector

Usage
disambiguateVector(in_vector)

Arguments
in_vector Input vector

Value
The disambiguated vector.

Examples
NULL

enrichSigs  Compare to background distribution

Description
Compare exposures from an analysis of mutational signatures in a cohort of interest to exposures computed in a background (e.g. the set of WES and WGS samples from Alexandrov 2013).

Usage
enrichSigs(in_cohort_exposures_df, in_background_exposures_df, in_sig_df)

Arguments
in_cohort_exposures_df Numerical data frame of the exposures of the cohort of interest.
in_background_exposures_df Numerical data frame of the exposures of the background.
in_sig_df Numerical data frame encoding the mutational signatures.

Value
A data frame with counts and p-values from Fisher tests.
ExampleINDEL_YAPSA

Data structures used in examples, Indel tests and the Indel signature vignette of the YAPSA package.

Description

Data structures used in examples, Indel tests and the Indel signature vignette of the YAPSA package.

Author(s)

Daniel Huebschmann <huebschmann.daniel@googlemail.com>

References


ExampleYAPSA

Test and example data

Description

Data structures used in examples, SNV tests and the SNV signature vignette of the YAPSA package.

lymphoma_PID_df: A data frame carrying subgroup information for a subcohort of samples used in the vignette. Data in the vignette is downloaded from ftp://ftp.sanger.ac.uk/pub/cancer/AlexandrovEtAl/somatic_mutation_data/Lymphoma%20B-cell/Lymphoma%20B-cell_clean_somatic_mutation_calls_for_signature_analysis.txt. In the file available under that link somatic point mutation calls from several samples are listed in a vcf-like format. One column encodes the sample the variant was found in. In the vignette we want to restrict the analysis to only a fraction of these involved samples. The data frame lymphoma_PID_df carries the sample identifiers (PID) as rownames and the attributed subgroup in a column called subgroup.

lymphoma_test_df: A data frame carrying point mutation calls. It represents a subset of the data stored in ftp://ftp.sanger.ac.uk/pub/cancer/AlexandrovEtAl/somatic_mutation_data/Lymphoma%20B-cell/Lymphoma%20B-cell_clean_somatic_mutation_calls_for_signature_analysis.txt. In the file available under that link somatic point mutation calls from several samples are listed in a vcf-like format. One column encodes the sample the variant was found in. The data frame lymphoma_test_df has only the variants occuring in the sample identifiers (PIDs) 4112512, 4194218 and 4121361.

In the file available under that link somatic point mutation calls from several samples are listed in a vcf-like format. One column encodes the sample the variant was found in.


`rel_lymphoma_Nature2013_COSMIC_cutoff_exposures_df`: Data frame with normalized or relative exposures for testing the plot functions. Data taken from ftp://ftp.sanger.ac.uk/pub/cancer/AlexandrovEtAl/somatic_mutation_data/LymphomaB-cell/LymphomaB-cell_clean_somatic_mutations_for_signature_analysis.txt.


`chosen_AlexInitialArtif_sigInd_df`: Signature information for the data stored in `lymphoma_Nature2013_COSMIC_cutoff_exposures_df` and `rel_lymphoma_Nature2013_COSMIC_cutoff_exposures_df`.


### Usage

```r
data(lymphoma_PID)
data(lymphoma_test)
data(lymphoma_Nature2013_raw)
data(lymphoma_cohort_LCD_results)
data(lymphoma_cohort_LCD_results)
data(lymphoma_cohort_LCD_results)
data(lymphoma_cohort_LCD_results)
data(lymphoma_cohort_LCD_results)
data(lymphoma_cohort_LCD_results)
data(lymphoma_cohort_LCD_results)
data(lymphoma_cohort_LCD_results)
```

### Author(s)

Daniel Huebschmann <huebschmann.daniel@googlemail.com>

### References


### Examples

```r
data(lymphoma_test)
head(lymphoma_test_df)
dim(lymphoma_test_df)
```
exchange_colour_vector

```r
table(lymphoma_test_df$PID)
data(lymphoma_Nature2013_raw)
head(lymphoma_Nature2013_raw_df)
dim(lymphoma_Nature2013_raw_df)
```

---

### Description

Vector attributing colours to nucleotide exchanges used when displaying SNV information, e.g. in a rainfall plot.

### Usage

```r
data(exchange_colour_vector)
```

### Value

A named character vector

### Author(s)

Daniel Huebschmann <huebschmann.daniel@googlemail.com>

---

### exome_mutCatRaw_df

**Example mutational catalog for the exome vignette**

**Description**

exome_mutCatRaw_df: A data frame in the format of a SNV mutation catalog. The mutational catalog contains SNV variants from a cohort of small-cell lung cancer published by Rudin et al. (Nature Genetics 2012) which was later used in the de novo discovery analysis of mutational signatures in human cancer by Alexandrov et al. (Nature 2013).

**Usage**

```r
data(smallCellLungCancerMutCat_NatureGenetics2012)
```

**Value**

A data frame in the layout of a SNV mutational catalog
References

https://www.nature.com/articles/ng.2405

Examples

data(smallCellLungCancerMutCat_NatureGenetics2012)
head(exome_mutCatRaw_df)
dim(exome_mutCatRaw_df)

exposures_barplot Wrapper for enhanced_barplot

Description

Wrapper for enhanced_barplot

Usage

exposures_barplot(
  in_exposures_df,
  in_signatures_ind_df = NULL,
  in_subgroups_df = NULL,
  in_sum_ind = NULL,
  in_subgroups.field = "subgroup",
  in_title = "",
  in_labels = TRUE,
  in_show_subgroups = TRUE,
  ylab = NULL,
  in_barplot_borders = TRUE,
  in_column_anno_borders = FALSE
)

Arguments

in_exposures_df
  Numerical data frame encoding the exposures H, i.e. which signature contributes how much to which PID (patient identifier or sample).

in_signatures_ind_df
  A data frame containing meta information about the signatures. If NULL, the colour information for the signatures is taken from a rainbow palette.

in_subgroups_df
  A data frame indicating which PID (patient or sample identifier) belongs to which subgroup. If NULL, it is assumed that all PIDs belong to one common subgroup. The colour coding for the default subgroup is red.

in_sum_ind
  Index vector influencing the order in which the PIDs are going to be displayed


extract_names_from_gene_list

extract_names_from_gene_list

Return gene names from gene lists

Description

Return gene names from gene lists

Usage

extract_names_from_gene_list(in_KEGG_gene_list, l)

Arguments

in_KEGG_gene_list
  Gene list to extract names from

l
  Index of the gene to be extracted

Value

The gene name.
find_affected_PIDs

See Also

   keggGet
   build_gene_list_for_pathway

Examples

   NULL

find_affected_PIDs  Find samples affected

Description

   Find samples affected by SNVs in a certain pathway

Usage

   find_affected_PIDs(in_gene_list, in_gene_vector, in_PID_vector)

Arguments

   in_gene_list    List of genes in the pathway of interest.
   in_gene_vector Character vector for genes annotated to SNVs as in vcf_like_df.
   in_PID_vector  Character vector for sample names annotated to SNVs as in vcf_like_df.

Value

   A character vector of the names of the affected samples

Examples

   NULL
**GenomeOfNl_raw**

**Example data for the Indel vignette**

**Description**

GenomeOfNl_raw: A data frame contains the germline variants of the Dutch population, carrying point mutation calls. It represents a subset of the data stored in ftp://ftp.sanger.ac.uk/pub/cancer/AlexandrovEtAl/somatic_mutation_data/Lymphoma%20B-cell/Lymphoma%20B-cell_clean_somatic_mutations_for_signature_analysis.txt. In the file available under that link, somatic point mutation calls from several samples are listed in a vcf-like format. One column encodes the sample the variant was found in.

**Usage**

data(GenomeOfNl_raw)

**Value**

A data frame in a vcf-like format

**References**

release version 5 https://www.nlgenome.nl/menu/main/app-go-nl/?page_id=9

**Examples**

data(GenomeOfNl_raw)
head(GenomeOfNl_raw)
dim(GenomeOfNl_raw)

---

**getSequenceContext**

Extracts the sequence context up and downstream of a nucleotide position

**Description**

Extracts the sequence context up and downstream of a nucleotide position

**Usage**

getSequenceContext(position, chr, offsetL = 10, offsetR = 50)
get_extreme_PIDs

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>position</td>
<td>Start position of the considered INDEL</td>
</tr>
<tr>
<td>chr</td>
<td>Chromosome of the considered INDEL</td>
</tr>
<tr>
<td>offsetL</td>
<td>Number of nucleotides downstream of position</td>
</tr>
<tr>
<td>offsetR</td>
<td>Number of nucleotides upstream of position</td>
</tr>
</tbody>
</table>

Value

Returns a character string containing the defined sequence context

Examples

```r
library(Biostrings)
library(BSgenome.Hsapiens.UCSC.hg19)

sequence_context <- getSequenceContext(position = 123456789, chr = "chr12",
                                        offsetL = 10, offsetR = 50)
sequence_context
```

get_extreme_PIDs

Return those PIDs which have an extreme pattern for signature exposure

Description

For all signatures found in a project, this function returns the sample identifiers (PIDs) with extremely high or extremely low exposures of the respective signatures.

Usage

```r
get_extreme_PIDs(in_exposures_df, in_quantile = 0.03)
```

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>in_exposures_df</td>
<td>Data frame with the signature exposures</td>
</tr>
<tr>
<td>in_quantile</td>
<td>Quantile for the amount of extreme PIDs to be selected.</td>
</tr>
</tbody>
</table>

Value

A data frame with 4 rows per signature (high PIDs, high exposures, low PIDs, low exposures); the number of columns depends on the quantile chosen.

Examples

```r
data(lymphoma_cohort_LCD_results)
get_extreme_PIDs(lymphoma_Nature2013_COSMIC_cutoff_exposures_df, 0.05)
```
hclust_exposures  

Cluster the PIDs according to their signature exposures

Description

The PIDs are clustered according to their signature exposures by calling first creating a distance matrix:

- `dist`, then
- `hclust` and then
- `labels_colors` to colour the labels (the text) of the leaves in the dendrogram.

Typically one colour per subgroup.

Usage

```r
hclust_exposures(
  in_exposures_df,
  in_subgroups_df,
  in_method = "manhattan",
  in_subgroup_column = "subgroup",
  in_palette = NULL,
  in_cutoff = 0,
  in_filename = NULL,
  in_shift_factor = 0.3,
  in_cex = 0.2,
  in_title = "",
  in_plot_flag = FALSE
)
```

Arguments

- `in_exposures_df`  
  Numerical data frame encoding the exposures H, i.e. which signature contributes how much to which PID (patient identifier or sample).

- `in_subgroups_df`  
  A data frame indicating which PID (patient or sample identifyier) belongs to which subgroup

- `in_method`  
  Method of the clustering to be supplied to `dist`. Can be either of: euclidean, maximum, manhattan, canberra, binary or minkowski

- `in_subgroup_column`  
  Indicates the name of the column in which the subgroup information is encoded in `in_subgroups_df`

- `in_palette`  
  Palette with colours or colour codes for the labels (the text) of the leaves in the dendrogram. Typically one colour per subgroup. If none is specified, a rainbow palette of the length of the number of subgroups will be used as default.
LCD

**Description**

LCD performs a mutational signatures decomposition of a given mutational catalogue \( V \) with known signatures \( W \) by solving the minimization problem \( \min(||W \ast H - V||) \) with additional constraints of non-negativity on \( H \) where \( W \) and \( V \) are known.

**Usage**

```r
LCD(in_mutation_catalogue_df, in_signatures_df, in_per_sample_cutoff = 0)
```

---

**in_cutoff**

A numeric value less than 1. Signatures from within \( W \) with an overall exposure less than \( \text{in\_cutoff} \) will be discarded for the clustering.

**in_filename**

A path to save the dendrogram. If none is specified, the figure will be plotted to the running environment.

**in_shift_factor**

Graphical parameter to adjust figure to be created

**in_cex**

Graphical parameter to adjust figure to be created

**in_title**

Title in the figure to be created under \( \text{in\_filename} \)

**in_plot_flag**

Whether or not to display the dendrogram

**Value**

A list with entries \texttt{hclust} and \texttt{dendrogram}.

- \texttt{hclust}: The object created by \texttt{hclust}
- \texttt{dendrogram}: The above object wrapped in \texttt{as.dendrogram}

**See Also**

- \texttt{hclust}
- \texttt{dist}
- \texttt{labels\_colors}

**Examples**

```r
data(lymphoma_cohort_LCD_results)
hclust_exposures(rel_lymphoma_Nature2013_COSMIC_cutoff_exposures_df, COSMIC_subgroups_df,
  in_method="manhattan",
  in_subgroup_column="subgroup")
```
Arguments

  in_mutation_catalogue_df
  A numeric data frame V with n rows and m columns, n being the number of
  features and m being the number of samples

  in_signatures_df
  A numeric data frame W with n rows and l columns, n being the number of
  features and l being the number of signatures

  in_per_sample_cutoff
  A numeric value less than 1. Signatures from within W with an exposure per
  sample less than in_cutoff will be discarded.

Value

  The exposures H, a numeric data frame with l rows and m columns, l being the number of signatures
  and m being the number of samples

See Also

lsei

Examples

## define raw data
W_prim <- matrix(c(1,2,3,4,5,6),ncol=2)
W_prim_df <- as.data.frame(W_prim)
W_df <- YAPSA:::normalize_df_per_dim(W_prim_df,2) # corresponds to the sigs
W <- as.matrix(W_df)
## 1. Simple case: non-negativity already in raw data
H <- matrix(c(2,5,3,6,1,9,1,2),ncol=4)
H_df <- as.data.frame(H) # corresponds to the exposures
V <- W %*% H # matrix multiplication
V_df <- as.data.frame(V) # corresponds to the mutational catalogue
exposures_df <- YAPSA:::LCD(V_df,W_df)
## 2. more complicated: raw data already contains negative elements
## define indices where sign is going to be swapped
sign_ind <- c(5,7)
## now compute the indices of the other fields in the columns affected
## by the sign change
row_ind <- sign_ind %% dim(H)[1]
temp_ind <- 2*row_ind -1
other_ind <- sign_ind + temp_ind
## alter the matrix H to yield a new mutational catalogue
H_compl <- H
H_compl[sign_ind] <- (-1)*H[sign_ind]
H_compl_df <- as.data.frame(H_compl) # corresponds to the exposures
V_compl <- W %*% H_compl # matrix multiplication
V_compl_df <- as.data.frame(V_compl) # corresponds to the mutational catalogue
exposures_df <- YAPSA:::LCD(V_compl_df,W_df)
exposures <- as.matrix(exposures_df)
**LCD_complex_cutoff**

**Description**

LCD_cutoff performs a mutational signatures decomposition by Linear Combination Decomposition (LCD) of a given mutational catalogue \( V \) with known signatures \( W \) by solving the minimization problem \( \min(||W \ast H - V||) \) with additional constraints of non-negativity on \( H \) where \( W \) and \( V \) are known, but excludes signatures with an overall contribution less than a given signature-specific cutoff (and thereby accounting for a background model) over the whole cohort.

**LCD_complex_cutoff_perPID** is a wrapper for **LCD_complex_cutoff** and runs individually for every PID.

**LCD_extractCohort_callPerPID** runs **LCD_complex_cutoff** and takes the identified signatures as input for **LCD_complex_cutoff_perPID**.

**LCD_complex_cutoff_consensus** calls **LCD_complex_cutoff_combined** AND **LCD_complex_cutoff_perPID** and makes a consensus signature call set.

**LCD_complex_cutoff_combined** is a wrapper for **LCD_complex_cutoff**, **LCD_complex_cutoff_perPID**, **LCD_complex_cutoff_consensus** AND **LCD_extractCohort_callPerPID**.

**Usage**

```r
LCD_complex_cutoff(
  in_mutation_catalogue_df,
  in_signatures_df,
  in_cutoff_vector = NULL,
  in_filename = NULL,
  in_method = "abs",
  in_per_sample_cutoff = 0,
  in_rescale = TRUE,
  in_sig_ind_df = NULL,
  in_cat_list = NULL
)

LCD_complex_cutoff_perPID(
  in_mutation_catalogue_df,
  in_signatures_df,
  in_cutoff_vector = NULL,
  in_filename = NULL,
  in_method = "abs",
  in_rescale = TRUE,
  in_sig_ind_df = NULL,
  in_cat_list = NULL,
  minimumNumberOfAlterations = 25
)
```
LCD_extractCohort_callPerPID(
    in_mutation_catalogue_df,
    in_signatures_df,
    in_cutoff_vector = NULL,
    in_filename = NULL,
    in_method = "abs",
    in_rescale = TRUE,
    in_sig_ind_df = NULL,
    in_cat_list = NULL,
    in_verbose = FALSE,
    minimumNumberOfAlterations = 25,
    cutoff_type = "adaptive"
)

LCD_complex_cutoff_consensus(
    in_mutation_catalogue_df = NULL,
    in_signatures_df = NULL,
    in_cutoff_vector = NULL,
    in_filename = NULL,
    in_method = "abs",
    in_rescale = TRUE,
    in_sig_ind_df = NULL,
    in_cat_list = NULL,
    in_cohort_LCDlist = NULL,
    in_perPID_LCDlist = NULL,
    addSigs_cohort_cutoff = 0.25,
    addSigs_perPID_cutoff = 0.25,
    addSigs_relAbs_cutoff = 0.01,
    keep.unassigned = FALSE,
    keep.all.cohort.sigs = TRUE,
    in_verbose = FALSE,
    minimumNumberOfAlterations = 25
)

LCD_complex_cutoff_combined(
    in_mutation_catalogue_df = NULL,
    in_signatures_df = NULL,
    in_cutoff_vector = NULL,
    in_filename = NULL,
    in_method = "abs",
    in_rescale = TRUE,
    in_sig_ind_df = NULL,
    in_cat_list = NULL,
    addSigs_cohort_cutoff = 0.25,
    addSigs_perPID_cutoff = 0.25,
    addSigs_relAbs_cutoff = 0.01,
    keep.all.cohort.sigs = TRUE,
    in_verbose = FALSE,
minimumNumberOfAlterations = 25,
cutoff_type = "adaptive"
)

Arguments

in_mutation_catalogue_df
A numeric data frame \(V\) with \(n\) rows and \(m\) columns, \(n\) being the number of features and \(m\) being the number of samples

in_signatures_df
A numeric data frame \(W\) with \(n\) rows and 1 columns, \(n\) being the number of features and 1 being the number of signatures

in_cutoff_vector
A numeric vector of values less than 1. Signatures from within \(W\) with an overall exposure less than the respective value in \(\text{in\_cutoff\_vector}\) will be discarded.

in_filename
A path to generate a histogram of the signature exposures if non-NULL

in_method
Indicate to which data the cutoff shall be applied: absolute exposures, relative exposures

in_per_sample_cutoff
A numeric value less than 1. Signatures from within \(W\) with an exposure per sample less than \(\text{in\_cutoff}\) will be discarded.

in_rescale
Boolean, if TRUE (default) the exposures are rescaled such that colSums over exposures match colSums over mutational catalogue

in_sig_ind_df
Data frame of type \text{signature\_indices\_df}, i.e. indicating name, function and meta-information of the signatures. Default is NULL.

in_cat_list
List of categories for aggregation. Have to be among the column names of \text{in\_sig\_ind\_df}. Default is NULL.

minimumNumberOfAlterations
The perPID part of the analysis issues a warning if one sample has less mutations than this minimum cutoff.

in_verbose
Verbose if \text{in\_verbose}=1

cutoff_type
If chosen to be "adaptive", the default, then signature-specific cutoffs are used for the the per-PID analysis in \text{LCD\_extractCohort\_callPerPID}, otherwise, no cutoffs are used.

in_cohort_LCDlist
Optional, if not provided, the cohort-wide exposures are recalculated by calling \text{LCD\_complex\_cutoff}

in_perPID_LCDlist
Optional, if not provided, the per sample exposures are recalculated by calling \text{LCD\_complex\_cutoff\_perPID}

addSigs_cohort_cutoff
Numeric value for a cutoff: signatures which are detected in a fraction of the samples of the cohort greater than this cutoff are kept for the consensus set of signatures
addSigs_perPID_cutoff
   Numeric value for a cutoff: signatures which are detected in one sample with exposure greater than this cutoff are kept for the consensus set of signatures

addSigs_relAbs_cutoff
   Numeric value for a cutoff: signatures which are detected with at least this fraction of all variants cohort wide are kept for the consensus set of signatures

keep.unassigned
   Boolean, if TRUE the exposures from the signatures which don’t fulfill the criteria to be kept will be added and stored in the exposures as "unassigned", otherwise the exposures are rescaled.

keep.all.cohort.sigs
   If TRUE (default), all signatures extracted cohort wide are kept, if FALSE, the function reevaluates whether the signatures extracted cohort wide still fulfill their criteria (i.e. exposures > cutoff) after perPID extraction.

Value
   A list with entries:

   - exposures: The exposures $H$, a numeric data frame with $l$ rows and $m$ columns, $l$ being the number of signatures and $m$ being the number of samples
   - norm_exposures: The normalized exposures $H$, a numeric data frame with $l$ rows and $m$ columns, $l$ being the number of signatures and $m$ being the number of samples
   - signatures: The reduced signatures that have exposures bigger than in_cutoff
   - choice: Index vector of the reduced signatures in the input signatures
   - order: Order vector of the signatures by exposure
   - residual_catalogue: Numerical data frame (matrix) of the difference between fit (product of signatures and exposures) and input mutational catalogue
   - rss: Residual sum of squares (i.e. sum of squares of the residual catalogue)
   - cosDist_fit_orig_per_matrix: Cosine distance between the fit (product of signatures and exposures) and input mutational catalogue computed after putting the matrix into vector format (i.e. one scaler product for the whole matrix)
   - cosDist_fit_orig_per_col: Cosine distance between the fit (product of signatures and exposures) and input mutational catalogue computed per column (i.e. per sample, i.e. as many scaler products as there are samples in the cohort)
   - sum_ind: Decreasing order of mutational loads based on the input mutational catalogue
   - out_sig_ind_df: Data frame of the type signature_indices_df, i.e. indicating name, function and meta-information of the signatures. Default is NULL, non-NULL only if in_sig_ind_df is non-NULL.
   - aggregate_exposures_list: List of exposure data frames aggregated over different categories. Default is NULL, non-NULL only if in_sig_ind_df and in_cat_list are non-NULL and if the categories specified in in_cat_list are among the column names of in_sig_ind_df.
See Also

LCD
aggregate_exposures_by_category
lsei

Examples

NULL

## Description

CD stratification analysis

## Usage

```r
LCD_SMC(in_mutation_sub_catalogue_list, in_signatures_df, in_F_df = NULL)
```

## Arguments

- `in_mutation_sub_catalogue_list`: A list of $s$ stratified mutational catalogues $V_i$ (numeric data frames) with $n$ rows and $m$ columns each, $n$ being the number of features and $m$ being the number of samples. This list is naturally provided in `run_SMC`.

- `in_signatures_df`: A numeric data frame $W$ with $n$ rows and $l$ columns, $n$ being the number of features and $l$ being the number of signatures.

- `in_F_df`: Default NULL

## Value

Returns a list with all exposures and the stratified ones.
logLikelihood

**Description**

Compute a likelihood ratio test based on the loglikelihoods of the residuals of two different models of the same data.

**Usage**

```r
logLikelihood(
  in_1,
  in_2,
  df_1 = NULL,
  df_2 = NULL,
  in_pdf = NULL,
  verbose = FALSE
)
```

**Arguments**

- `in_1`: Residuals of model 1 of the input data.
- `in_2`: Residuals of model 2 of the input data.
- `df_1`: Degrees of freedom of the input model 1. If either `df_1` or `df_2` is `NULL`, the difference between the degrees of freedom of the two models is assumed to be 1.
- `df_2`: Degrees of freedom of the input model 2. If either `df_1` or `df_2` is `NULL`, the difference between the degrees of freedom of the two models is assumed to be 1.
- `in_pdf`: Probability distribution function, passed on to `computeLogLik`, if `NULL` a normal distribution is used.
- `verbose`: Verbose if `in_verbose=1`

**Value**

A list with entries

- `statistic`: The test statistic
- `delta_df`: The difference in degrees of freedom between input model 1 and 2
- `p.value`: p value of the statistical test.
Examples

```r
library(BSgenome.Hsapiens.UCSC.hg19)
data(lymphoma_test)
data(sigs)
data(cutoffs)
word_length <- 3
temp_list <- create_mutation_catalogue_from_df(
  lymphoma_test_df, this_seqnames.field = "CHROM",
  this_start.field = "POS", this_end.field = "POS",
  this_PID.field = "PID", this_subgroup.field = "SUBGROUP",
  this_refGenome = BSgenome.Hsapiens.UCSC.hg19,
  this_wordLength = word_length)
lymphoma_catalogue_df <- temp_list$matrix
lymphoma_PIDs <- colnames(lymphoma_catalogue_df)
current_sig_df <- AlexCosmicValid_sig_df
current_sigInd_df <- AlexCosmicValid_sigInd_df
current_cutoff_vector <- cutoffCosmicValid_rel_df[6, ]
iniLCDList <- LCD_complex_cutoff(
  in_mutation_catalogue_df = lymphoma_catalogue_df[, 1, drop = FALSE],
  in_signatures_df = current_sig_df,
  in_cutoff_vector = current_cutoff_vector,
  in_method = "relative", in_rescale = TRUE,
  in_sig_ind_df = current_sigInd_df)
current_sig_df <- AlexCosmicValid_sig_df[, -9]
current_sigInd_df <- AlexCosmicValid_sigInd_df[-9,]
current_cutoff_vector <- cutoffCosmicValid_rel_df[6, -9]
redLCDList <- LCD_complex_cutoff(
  in_mutation_catalogue_df = lymphoma_catalogue_df[, 1, drop = FALSE],
  in_signatures_df = current_sig_df,
  in_cutoff_vector = current_cutoff_vector,
  in_method = "relative", in_rescale = TRUE,
  in_sig_ind_df = current_sigInd_df)
logLikelihood(iniLCDList, redLCDList)
```

#### lymphomaNature2013_mutCat_df

*Example mutational catalog for the SNV vignette*

**Description**

lymphomaNature2013_mutCat_df: A data frame in the format of a SNV mutation catalog. The mutational catalog contains SNV variants from the lymphoma_Nature2013_raw_df data. Mutational catalog was created with create_mutation_catalogue_from_df function.

**Usage**

data(lymphomaNature2013_mutCat_df)
Value

A data frame in the layout of a SNV mutational catalog

References

ftp://ftp.sanger.ac.uk/pub/cancer/AlexandrovEtAl/somatic_mutation_data/Lymphoma%20B-cell/Lymphoma%20B-cell_clean_somatic_mutations_for_signature_analysis.txt

Examples

data(lymphomaNature2013_mutCat_df)
head(lymphomaNature2013_mutCat_df)
dim(lymphomaNature2013_mutCat_df)

makeVRangesFromDataFrame

Construct a VRanges Object from a data frame

Description

In this package, big data frames are generated from cohort wide vcf-like files. This function constructs a VRanges object from such a data frame by using makeGRangesFromDataFrame from the package GenomicRanges

Usage

makeVRangesFromDataFrame(
  in_df,
  in_keep.extra.columns = TRUE,
  in_seqinfo = NULL,
  in_seqnames.field = "X.CHROM",
  in_start.field = "POS",
  in_end.field = "POS",
  in_PID.field = "PID",
  in_subgroup.field = "subgroup",
  in_strand.field = "strand",
  verbose_flag = 1
)

Arguments

in_df A big dataframe constructed from a vcf-like file of a whole cohort. The first columns are those of a standard vcf file, followed by an arbitrary number of custom or user defined columns. One of these can carry a PID (patient or sample identifier) and one can carry subgroup information.
in_keep.extra.columns

in_seqinfo  Argument passed on to makeGRangesFromDataFrame

in_seqinfo  A seqInfo object, referring to the reference genome used. Argument passed on
to makeGRangesFromDataFrame

in_seqnames.field  Indicates the name of the column in which the chromosome is encoded

in_start.field  Indicates the name of the column in which the start coordinate is encoded

in_end.field  Indicates the name of the column in which the end coordinate is encoded

in_PID.field  Indicates the name of the column in which the PID (patient or sample identifier)
is encoded

in_subgroup.field  Indicates the name of the column in which the subgroup information is encoded

in_strand.field  Indicates the name of the column in which the strandedness is encoded

verbose_flag  Verbose if 1

Value

The constructed VRanges object

See Also

makeGRangesFromDataFrame

Examples

data(lymphoma_test)
temp_vr <- makeVRangesFromDataFrame(lymphoma_test_df,
in_seqnames.field="CHROM",
in_subgroup.field="SUBGROUP",
verbose_flag=1)

make_catalogue_strata_df

Group strata from different stratification axes

Description

For a comparison of the strata from different orthogonal stratification axes, i.e. orthogonal SMCs,
the strata have to be grouped and reformatted. This function does this task for the comparison
by cosine similarity of mutational catalogues. Output of this function is the basis for applying
make_comparison_matrix. It is called by the wrapper function run_comparison_catalogues.
Usage

```r
make_catalogue_strata_df(
    in_stratification_lists_list,
    in_additional_stratum = NULL
)
```

Arguments

- `in_stratification_lists_list`: List of lists with entries from different (orthogonal) stratification axes or SMCs
- `in_additional_stratum`: Include an additionally supplied stratum in comparison in non-NULL.

Value

A list with entries `strata_df`, `number_of_SMCs`, `number_of_strata`.
- `strata_df`: Past numerical data frame of all strata (these are going to be compared e.g. by `make_comparison_matrix`).
- `number_of_SMCs`: Number of orthogonal stratifications in `in_stratification_lists_list` and additional ones.
- `number_of_strata`: Cumulative number of strata (sum over the numbers of strata of the different stratifications in `in_stratification_lists_list` and additional ones.

See Also

- `plot_strata`
- `make_comparison_matrix`
- `run_comparison_catalogues`

Examples

```r
NULL
```

make_comparison_matrix

*Compute a similarity matrix for different strata*

Description

Compute and plot a similarity matrix for different strata from different stratification axes together. First, `compare_sets` is called on `in_strata_df` with itself, yielding a distance matrix (a numerical data frame) `dist_df` of the strata. The corresponding similarity matrix `1-dif_df` is then passed to `corrplot`. 
Usage

```r
make_comparison_matrix(
  in_strata_df,
  output_path = NULL,
  in_nrect = 5,
  in_attribute = "",
  in_palette = NULL
)
```

Arguments

- **in_strata_df**: Numerical data frame of all strata to be compared.
- **output_path**: Path to directory where the results, especially the figure produced by `corrplot` is going to be stored.
- **in_nrect**: Number of clusters in the clustering procedure provided by `corrplot`.
- **in_attribute**: Additional string for the file name where the figure produced by `corrplot` is going to be stored.
- **in_palette**: Colour palette for the matrix

Value

The comparison matrix of cosine similarities.

See Also

- `compare_SMCs`

Examples

```r
data(sigs)
make_comparison_matrix(
  AlexCosmicValid_sig_df, in_nrect=9,
  in_palette=colorRampPalette(c("blue","green","red"))(n=100))
```

---

**make_strata_df**  
*Group strata from different stratification axes*

Description

For a comparison of the strata from different orthogonal stratification axes, i.e. orthogonal SMCs, the strata have to be grouped and reformatted. This function does this task for the comparison by cosine similarity of signature exposures. Output of this function is the basis for applying `plot_strata` and `make_comparison_matrix`. It is called by the wrapper functions `compare_SMCs`, `run_plot_strata_general` or `run_comparison_general`. 
make_strata_df

Usage

make_strata_df(
    in_stratification_lists_list,
    in_remove_signature_ind = NULL,
    in_additional_stratum = NULL
)

Arguments

in_stratification_lists_list
    List of lists with entries from different (orthogonal) stratification axes or SMCs
in_remove_signature_ind
    Omit one of the signatures in in_signatures_ind_df for the comparison if non-NULL. The parameter specifies the index of the signature to be removed.
in_additional_stratum
    Include an additionally supplied stratum in comparison in non-NULL.

Value

A list with entries strata_df, number_of_SMCs, number_of_strata.

- strata_df: Pasted numerical data frame of all strata (these are going to be compared e.g. by make_comparison_matrix).
- number_of_SMCs: Number of orthogonal stratifications in in_stratification_lists_list and additional ones.
- number_of_strata: Cumulative number of strata (sum over the numbers of strata of the different stratifications in in_stratification_lists_list) and additional ones.

See Also

plot_strata
make_comparison_matrix
compare_SMCs
run_plot_strata_general
run_comparison_general

Examples

NULL
make_subgroups_df

Make a custom data structure for subgroups

Description

Creates a data frame carrying the subgroup information and the order in which the PIDs have to be displayed. Calls aggregate on in_vcf_like_df.

Usage

make_subgroups_df(
  in_vcf_like_df,
  in_exposures_df = NULL,
  in_palette = NULL,
  in_subgroup.field = "SUBGROUP",
  in_PID.field = "PID",
  in_verbose = FALSE
)

Arguments

in_vcf_like_df  vcf-like data frame with point mutation calls
in_exposures_df Data frame with the signature exposures
in_palette     Palette for colour attribution to the subgroups if non-NULL
in_subgroup.field String indicating which column of in_vcf_like_df carries the subgroup information
in_PID.field   String indicating which column of in_vcf_like_df and of in_exposures_df carries the PID information
in_verbose     Whether verbose or not.

Value

subgroups_df: A data frame carrying the subgroup and rank information.

See Also

aggregate

Examples

data(lymphoma_test)
data(lymphoma_cohort_LCD_results)
choice_ind <- (names(lymphoma_Nature2013_COSMIC_cutoff_exposures_df)
                  %in% unique(lymphoma_test_df$PID))
lymphoma_test_exposures_df <-
melt_exposures

Generically melts exposure data frames

Description
Melt an exposure data frame with signatures as ID variables.

Usage
melt_exposures(in_df)

Arguments
in_df Numeric data frame with exposures.

Value
A data frame with the molten exposures.

Examples
NULL

merge_exposures
Merge exposure data frames

Description
Merges with the special feature of preserving the signatures and signature order.

Usage
merge_exposures(in_exposures_list, in_signatures_df)

Arguments
in_exposures_list List of data frames (carrying information on exposures).

in_signatures_df Data frame \( \mathbb{W} \) in which the columns represent the signatures.
**Value**

A data frame with the merged exposures.

**Examples**

```r
NULL
```

---

**Description**

`MutCat_indel_df`: A data frame in the format of a mutation catalog. The mutational catalog contains Indel variants from the `GenomeOfNL_raw` data. Variants were random sampled for 15 artificial patient for the purpose to have a Indel mutational catalog and have to show the functionality of the package. The results of the mutational catalog should not be interpreted for their biological relevance. Mutational catalog was created with `create_indel_mutation_catalogue_from_df` function.

**Usage**

```r
data(GenomeOfNL_MutCat)
```

**Value**

A data frame in the layout of a Indel mutational catalog

**References**

Mutational catalog created form release version 5 of the Genome of NL [https://www.nlgenome.nl/menu/main/app-go-nl/?page_id=9](https://www.nlgenome.nl/menu/main/app-go-nl/?page_id=9)

**Examples**

```r
data(GenomeOfNL_MutCat)
head(MutCat_indel_df)
dim(MutCat_indel_df)
```
**normalizeMotifs_otherRownames**

*Normalize Somatic Motifs with different rownames*

**Description**

This is a wrapper function to `normalizeMotifs`. The rownames are first transformed to fit the convention of the `SomaticSignatures` package and then passed on to the above mentioned function.

**Usage**

```r
normalizeMotifs_otherRownames(in_matrix, in_norms, adjust_counts = TRUE)
```

**Arguments**

- in_matrix, in_norms
  - Arguments to `normalizeMotifs`
- adjust_counts
  - Whether to rescale the counts after adaption or not. Default is true.

**Value**

The matrix returned by `normalizeMotifs`, but with rownames transformed back to the convention of the input

**Examples**

```
NULL
```

**normalize_df_per_dim**

*Useful functions on data frames*

**Description**

- `normalize_df_per_dim`: Normalization is carried out by dividing by `rowSums` or `colSums`; for rows with `rowSums`=0 or columns with `colSums`=0, the normalization is left out.
- `average_over_present`: If averaging over columns, zero rows (i.e. those with `rowSums`=0) are left out, if averaging over rows, zero columns (i.e. those with `colSums`=0) are left out.
- `sd_over_present`: If computing the standard deviation over columns, zero rows (i.e. those with `rowSums`=0) are left out, if computing the standard deviation over rows, zero columns (i.e. those with `colSums`=0) are left out.
- `stderrmean_over_present`: If computing the standard error of the mean over columns, zero rows (i.e. those with `rowSums`=0) are left out, if computing the standard error of the mean over rows, zero columns (i.e. those with `colSums`=0) are left out. Uses the function `stderrmean`
normalize_df_per_dim

Usage

normalize_df_per_dim(in_df, in_dimension)

average_over_present(in_df, in_dimension)

sd_over_present(in_df, in_dimension)

stderrmean_over_present(in_df, in_dimension)

Arguments

in_df Data frame to be normalized
in_dimension Dimension along which the operation will be carried out

Value

The normalized numerical data frame (normalize_df_per_dim)
A vector of the means (average_over_present)
A vector of the standard deviations (sd_over_present)
A vector of the standard errors of the mean (stderrmean_over_present)

See Also

stderrmean

Examples

test_df <- data.frame(matrix(c(1,2,3,0,5,2,3,4,0,6,0,0,0,0,4,5,6,0,7),
ncol=4))
## 1. Normalize over rows:
normalize_df_per_dim(test_df,1)
## 2. Normalize over columns:
normalize_df_per_dim(test_df,2)

test_df <- data.frame(matrix(c(1,2,3,0,5,2,3,4,0,6,0,0,0,0,4,5,6,0,7),
ncol=4))
## 1. Average over non-zero rows:
average_over_present(test_df,1)
## 2. Average over non-zero columns:
average_over_present(test_df,2)

test_df <- data.frame(matrix(c(1,2,3,0,5,2,3,4,0,6,0,0,0,0,4,5,6,0,7),
ncol=4))
## 1. Compute standard deviation over non-zero rows:
sd_over_present(test_df,1)
## 2. Compute standard deviation over non-zero columns:
sd_over_present(test_df,2)

test_df <- data.frame(matrix(c(1,2,3,0,5,2,3,4,0,6,0,0,0,0,4,5,6,0,7),
ncol=4))
## Example

```r
ncol=4))
## 1. Compute standard deviation over non-zero rows:
stderrmean_over_present(test_df,1)
## 2. Compute standard deviation over non-zero columns:
stderrmean_over_present(test_df,2)
```

---

### plotExchangeSpectra

**Plot the spectra of nucleotide exchanges**

Plots the spectra of nucleotide exchanges in their triplet contexts. If several columns are present in the input data frame, the spectra are plotted for every column separately.

#### Usage

```r
plotExchangeSpectra(
  in_catalogue_df,
  in_colour_vector = NULL,
  in_show_triplets = FALSE,
  in_show_axis_title = FALSE,
  in_scales = "free_x",
  in_refLine = NULL,
  in_refAlpha = 0.5,
  in_background = NULL
)
```

#### Arguments

- **in_catalogue_df**
  - Numerical data frame encoding the exchange spectra to be displayed, either a mutational catalogue \( V \) or a signatures matrix \( W \).
- **in_colour_vector**
  - Specifies the colours of the 6 nucleotide exchanges if non-null.
- **in_show_triplets**
  - Whether or not to show the triplets on the x-axis
- **in_show_axis_title**
  - Whether or not to show the name of the y-axis
- **in_scales**
  - Argument passed on to `facet_grid`
- **in_refLine**
  - If non-null, value on the y-axis at which a horizontal line is to be drawn
- **in_refAlpha**
  - Transparency of the horizontal line if it is to be drawn
- **in_background**
  - Option to provide a background theme, e.g. `theme_grey`

#### Value

The generated barplot - a ggplot2 plot
plotExchangeSpectra_indel

Plot the spectra of nucleotide exchanges of INDELs

Description

Plots the spectra of nucleotides in their triplet contexts. If several columns are present in the input data frame, the spectra are plotted for every column separately. The function is only suitable for an INDEL spectra and for SNV representation the function `plotExchangeSpectra` should be used.

Usage

```r
plotExchangeSpectra_indel(
    in_catalogue_df,
    in_colour_vector = NULL,
    in_show_indel = FALSE,
    in_show_axis_title = FALSE,
    in_scales = "free_x",
    in_refLine = NULL,
    in_refAlpha = 0.5,
    in_background = NULL
)
```

Arguments

- `in_catalogue_df`: Numerical data frame encoding the exchange spectra to be displayed, either a mutational catalogue `V` or a signatures matrix `W`
- `in_colour_vector`: Specifies the colours of the INDELs if non-null
- `in_show_indel`: Whether or not to show the INDEL names on the x-axis
- `in_show_axis_title`: Whether or not to show the name of the y-axis
- `in_scales`: Argument passed on to `facet_grid`
- `in_refLine`: If non-null, value on the y-axis at which a horizontal line is to be drawn
- `in_refAlpha`: Transparency of the horizontal line if it is to be drawn
- `in_background`: Option to provide a background theme, e.g. `theme_grey`
plotExposuresConfidence

Value

The generated barplot - a ggplot2 plot

Examples

data(sigs_pcawg)
plotExchangeSpectra_indel(PCAWG_SP_ID_sigs_df[,c(6,8)])

plotExposuresConfidence

Plot exposures including confidence intervals

Description

Plot the exposures to extracted signatures including confidence intervals computed e.g. by variate-Exp.

Usage

plotExposuresConfidence(in_complete_df, in_subgroups_df, in_sigInd_df)

Arguments

in_complete_df Melted numeric input data frame e.g. as computed by variateExp
in_subgroups_df Data frame containing meta information on subgroup attribution of the samples in the cohort of interest.
in_sigInd_df Data frame with meta information on the signatures used in the analysis.

Value

The function doesn’t return any value but plots instead.

Examples

NULL
plotExposuresConfidence_indel

Plot exposures including confidence intervals for exposures of SNVs and INDELS

Description

Plot the exposures to extracted signatures including the confidence intervals computed e.g. by variateExp

Usage

plotExposuresConfidence_indel(in_complete_df, in_subgroups_df, in_sigInd_df)

Arguments

in_complete_df   Melted numeric input data frame e.g. as computed by variateExp
in_subgroups_df  Data frame containing meta information on subgroup attribution of the samples in the cohort of interest.
in_sigInd_df     Data frame with meta information on the signatures used in the analysis.

Value

The function returns a gtable object which can be plotted with plot or grid.draw

Examples

NULL

plot_exposures

Plot the exposures of a cohort

Description

plot_exposures: The exposures \( H \), determined by NMF or by LCD, are displayed as a stacked barplot by calling

- geom_bar and optionally
- geom_text.

The x-axis displays the PIDs (patient identifier or sample), the y-axis the counts attributed to the different signatures with their respective colours per PID. Is called by plot_relative_exposures.

plot_relative_exposures: Plot the relative or normalized exposures of a cohort. This function first normalizes its input and then sends the normalized data to plot_exposures.
plot_exposures

Usage

plot_exposures(
  in_exposures_df,
  in_signatures_ind_df,
  in_subgroups_df = NULL,
  in_sum_ind = NULL,
  in_subgroups.field = "subgroup",
  in_title = "",
  in_labels = TRUE,
  in_show_subgroups = TRUE,
  legend_height = 10
)

plot_relative_exposures(
  in_exposures_df,
  in_signatures_ind_df,
  in_subgroups_df,
  in_sum_ind = NULL,
  in_subgroups.field = "subgroup",
  in_title = "",
  in_labels = TRUE,
  in_show_subgroups = TRUE
)

Arguments

in_exposures_df
  Numerical data frame encoding the exposures H, i.e. which signature contributes
  how much to which PID (patient identifier or sample).

in_signatures_ind_df
  A data frame containing meta information about the signatures

in_subgroups_df
  A data frame indicating which PID (patient or sample identifyier) belongs to
  which subgroup

in_sum_ind
  Index vector influencing the order in which the PIDs are going to be displayed

in_subgroups.field
  String indicating the column name in in_subgroups_df to take the subgroup
  information from.

in_title
  Title for the plot to be created.

in_labels
  Flag, if TRUE the PIDs are displayed on the x-axis

in_show_subgroups
  Flag, if TRUE then PIDs are grouped by subgroups

legend_height
  How many signatures should be displayed in one column together at most.

Value

The generated barplot - a ggplot2 plot
See Also

LCD
geom_bar
geom_text

Examples

data(lymphoma_cohort_LCD_results)
plot_exposures(lymphoma_Nature2013_COSMIC_cutoff_exposures_df, chosen_signatures_indices_df, COSMIC_subgroups_df)

data(lymphoma_cohort_LCD_results)
plot_relative_exposures(lymphoma_Nature2013_COSMIC_cutoff_exposures_df, chosen_signatures_indices_df, COSMIC_subgroups_df)

plot_SMC

Plot results of the Stratification of a Mutational Catalogue

Description

Plot a big composite figure with 3 columns: in the left column the per-PID absolute exposures will be shown, in the middle column the per_PID relative or normalized exposures will be shown, in the right column the cohort-wide exposures are shown (averaged over PIDs).

Usage

plot_SMC(
  number_of_strata, 
  output_path, 
  decomposition_method, 
  number_of_sigs, 
  name_list, 
  exposures_strata_list, 
  this_signatures_ind_df, 
  this_subgroups_df, 
  in_strata_order_ind, 
  exposures_both_rel_df_list, 
  cohort_method_flag, 
  fig_width = 1200, 
  fig_height = 900, 
  fig_type = "png", 
  in_label_orientation = "turn", 
  this_sum_ind = NULL
)
**Arguments**

- `number_of_strata`: Number of strata as deduced from `link{SMC}`
- `output_path`: Path to file where the results are going to be stored. If NULL, the results will be plotted to the running environment.
- `decomposition_method`: String for the filename of the generated barplot.
- `number_of_sigs`: Number of signatures
- `name_list`: Names of the constructed strata.
- `exposures_strata_list`: The list of strata specific exposures $H_i$, all are numerical data frames with $l$ rows and $m$ columns, $l$ being the number of signatures and $m$ being the number of samples
- `this_signatures_ind_df`: A data frame containing meta information about the signatures
- `this_subgroups_df`: A data frame indicating which PID (patient or sample identifyier) belongs to which subgroup
- `in_strata_order_ind`: Index vector defining reordering of the strata
- `exposures_both_rel_df_list`: A list of strata specific cohortwide (i.e. averaged over cohort) normalized exposures
- `cohort_method_flag`: Either or several of c("all_PIDs","cohort","norm_PIDs"). representing alternative ways to average over the cohort.
- `fig_width`: Width of the figure to be plotted
- `fig_height`: Height of the figure to be plotted
- `fig_type`: png or pdf
- `in_label_orientation`: Whether or not to turn the labels on the x-axis.
- `this_sum_ind`: Optional set of indices for reordering the PIDs

**Value**

The function doesn’t return any value.

**Examples**

```r
NULL
```
plot_strata  

Plot all strata from different stratification axes together

Description

Plot the cohort wide signature exposures of all strata from different stratification axes together. Naturally called by compare_SMCs.

Usage

plot_strata(
  in_strata_list,
  in_signatures_ind_df,
  output_path = NULL,
  in_attribute = ""
)

Arguments

in_strata_list  Data structure created by make_strata_df or make_catalogue_strata_df in which the strata from different orthogonal stratification axes are reorganized in a consistent structure.

in_signatures_ind_df  A data frame containing meta information about the signatures

output_path  Path to directory where the results, especially the figure produced, are going to be stored.

in_attribute  Additional string for the file name where the figure output is going to be stored.

Value

The function doesn’t return any value.

See Also

compare_SMCs

Examples

NULL
**read_entry**  

*Read a single vcf-like file into a single data frame*

**Description**

Note: this function uses `read.csv` to read vcf-like files into data frames for single samples. As it uses `read.csv`, the default value for `comment.char` is "" and not "#" as it would have been for `read.table`.

**Usage**

```r
read_entry(
  current_ind,  
  in_list,      
  header = TRUE, 
  in_header = NULL,  
  variant_type = "SNV",  
  delete.char = NULL,  
  ...  
)
```

```r
read_list(in_list, in_parallel = FALSE, header = TRUE, in_header = NULL, ...)
```

**Arguments**

- `current_ind`: Index of the file to read from the list provided below.
- `in_list`: List of paths to vcf-like file to be read. The list may be named.
- `header`: Boolean whether a header information should be read (as in `read.table`)
- `in_header`: Vector of column names to be substituted if non-NULL.
- `variant_type`: Default is "SNV" and provides additional plausibility and checks, omitted if other string
- `delete.char`: Character to be deleted, e.g. in order to discriminate between comment lines and header lines, if non-NULL
- `...`: Parameters passed on to `read.table`
- `in_parallel`: If multicore functionality is provided on a compute cluster, this option may be set to TRUE in order to enhance speed.

**Value**

A vcf-like data frame

A list with entries:

- `vcf_like_df_list`: List of the read data frames
- `readVcf_time`: Object of class proc.time, which stores the time needed for reading in the data
relateSigs

Examples

NULL

NULL

relateSigs Make unique assignments between sets of signatures

Description

Make unique assignments between a set of given signatures and a set of new signatures.

Usage

relateSigs(querySigs, subjectSigs)

Arguments

querySigs The signatures to compare to (given signatures).
subjectSigs The signatures to be compared (new signatures).

Value

A list of comparison vectors

See Also

compare_sets
disambiguateVector

Examples

NULL
**repeat_df**

Create a data frame with default values

**Description**

Create a data frame with default values

**Usage**

repeat_df(in_value, in_rows, in_cols)

**Arguments**

- **in_value**: Default entry to be repeated in the data frame
- **in_rows, in_cols**: Dimensions of the data frame to be created

**Value**

The created data frame

**Examples**

```r
## 1. Initialize with numeric value:
repeat_df(1,2,3)
## 2. Initialize with NA value:
repeat_df(NA,3,2)
## 3. Initialize with character:
repeat_df("a",4,3)
```

---

**round_precision**

Round to a defined precision

**Description**

This function is an extension with regard to the function `round` from base R as it allows not only digits as precision, but can also round to a user-specified precision. The interval in which the rounding operation is to be carried out also can be specified by the user (default is the unit interval). Alternatively, breaks can be provided.

**Usage**

round_precision(x, breaks = NULL, in_precision = 0.05, in_interval = c(0, 1))
Arguments

- `x`: Vector to be rounded
- `breaks`: The breaks used for rounding. Default NULL
- `in_precision`: Precision default 0.05
- `in_interval`: Interval needs to be larger than the precision value

Value

A list with two entries:

- `values`: the rounded vector
- `breaks`: the breaks used for rounding

Examples

`NULL`

Description

Wrapper function to the perl script annotate_vcf.pl which annotates data of a track stored in file_B (may be different formats) to called variants stored in a vcf-like file_A.

Usage

```r
run_annotate_vcf_pl(
  in_data_file,
  in_anno_track_file,
  in_new_column_name,
  out_file,
  in_data_file_type = "custom",
  in_anno_track_file_type = "bed",
  in_data_CHROM.field = "CHROM",
  in_data_POS.field = "POS",
  in_data_END.field = "POS"
)
```
Arguments

in_data_file    Path to the input vcf-like file to be annotated
in_anno_track_file    Path to the input file containing the annotation track
in_new_column_name    String indicating the name of the column to be created for annotation.
out_file    Path where the created files can be stored.
in_data_file_type custom for vcf-like
in_anno_track_file_type Type of the file in_anno_track_file containing the annotation track.
in_data_CHROM.field     String indicating which column of in_data_file contains the chromosome information.
in_data_POS.field     String indicating which column of in_data_file contains the position information.
in_data_END.field     String indicating which column of in_data_file contains the end information if regions are considered.

Value

Return zero if no problems occur.

Examples

NULL

---

**run_comparison_catalogues**

*Compare all strata from different stratifications*

---

**Description**

Compare all strata from different orthogonal stratification axes, i.e. orthogonal SMCs by cosine similarity of mutational catalogues. Function similar to **run_comparison_general**. First calls

- make_catalogue_strata_df, then
- make_comparison_matrix
Usage

\[
\text{run\_comparison\_catalogues(}
\begin{align*}
\text{in\_stratification\_lists\_list}, \\
\text{output\_path = NULL,} \\
\text{in\_nrect = 5,} \\
\text{in\_attribute = ""}
\end{align*}
\)\]

Arguments

- \textit{in\_stratification\_lists\_list} : List of lists with entries from different (orthogonal) stratification axes or SMCs
- \textit{output\_path} : Path to directory where the results, especially the figure produced by \texttt{corrplot} is going to be stored.
- \textit{in\_nrect} : Number of clusters in the clustering procedure provided by \texttt{corrplot}
- \textit{in\_attribute} : Additional string for the file name where the figure produced by

Value

The comparison matrix of cosine similarities.

See Also

- \texttt{make\_comparison\_matrix}
- \texttt{run\_comparison\_general}

Examples

\[
\text{NULL}
\]
**run_comparison_general**

**Usage**

```r
run_comparison_general(
  in_stratification_lists_list,
  output_path = NULL,
  in_nrect = 5,
  in_attribute = "",
  in_remove_signature_ind = NULL,
  in_additional_stratum = NULL
)
```

**Arguments**

- `in_stratification_lists_list`  
  List of lists with entries from different (orthogonal) stratification axes or SMCs
- `output_path`  
  Path to directory where the results, especially the figure produced by `corrplot` is going to be stored.
- `in_nrect`  
  Number of clusters in the clustering procedure provided by `corrplot`
- `in_attribute`  
  Additional string for the file name where the figure produced by `corrplot` is going to be stored.
- `in_remove_signature_ind`  
  Omit one of the signatures in `in_signatures_ind_df` for the comparison if non-NULL. The parameter specifies the index of the signature to be removed.
- `in_additional_stratum`  
  Include an additionally supplied stratum in comparison in non-NULL.

**Value**

The comparison matrix of cosine similarities.

**See Also**

- `make_comparison_matrix`
- `compare_SMCs`
- `run_comparison_catalogues`

**Examples**

```r
NULL
```
run_kmer_frequency_correction

Provide comprehensive correction factors for kmer content

Description

This function is analogous to normalizeMotifs. If an analysis of mutational signatures is performed on e.g. Whole Exome Sequencing (WES) data, the signatures and exposures have to be adapted to the potentially different kmer (trinucleotide) content of the target capture. The present function takes as arguments paths to the used reference genome and target capture file. It extracts the sequence of the target capture by calling bedtools getfasta on the system command prompt. run_kmer_frequency_normalization then calls a custom made perl script kmer_frequencies.pl also included in this package to count the occurrences of the triplets in both the whole reference genome and the created target capture sequence. These counts are used for normalization as in normalizeMotifs. Note that kmerFrequency provides a solution to approximate kmer frequencies by random sampling. As opposed to that approach, the function described here deterministically counts all occurrences of the kmers in the respective genome.

Usage

run_kmer_frequency_correction(
    in_ref_genome_fasta,
    in_target_capture_bed,
    in_word_length,
    project_folder,
    target_capture_fasta = "targetCapture.fa",
    in_verbose = 1
)

Arguments

in_ref_genome_fasta
    Path to the reference genome fasta file used.

in_target_capture_bed
    Path to a bed file containing the information on the used target capture. May also be a compressed bed.

in_word_length
    Integer number defining the length of the features or motifs, e.g. 3 for tripletts or 5 for pentamers

project_folder
    Path where the created files, especially the fasta file with the sequence of the target capture and the count matrices, can be stored.

target_capture_fasta
    Name of the fasta file of the target capture to be created if not yet existent.

in_verbose
    Verbose if in_verbose=1
Value

A list with 2 entries:

- rel_cor: The correction factors after normalization as in `run_kmer_frequency_normalization`
- abs_cor: The correction factors without normalization.

See Also

`normalizeMotifs`

Examples

`NULL`

```r
run_kmer_frequency_normalization

Provide normalized correction factors for kmer content
```

Description

This function is analogous to `normalizeMotifs`. If an analysis of mutational signatures is performed on e.g. Whole Exome Sequencing (WES) data, the signatures and exposures have to be adapted to the potentially different kmer (trinucleotide) content of the target capture. The present function takes as arguments paths to the used reference genome and target capture file. It extracts the sequence of the target capture by calling `bedtools getfasta` on the system command prompt. `run_kmer_frequency_normalization` then calls a custom made perl script `kmer_frequencies.pl` also included in this package to count the occurrences of the triplets in both the whole reference genome and the created target capture sequence. These counts are used for normalization as in `normalizeMotifs`. Note that `kmerFrequency` provides a solution to approximate kmer frequencies by random sampling. As opposed to that approach, the function described here deterministically counts all occurrences of the kmers in the respective genome.

Usage

```r
run_kmer_frequency_normalization(
  in_ref_genome_fasta,
  in_target_capture_bed,
  in_word_length,
  project_folder,
  inVerbose = 1
)
```
Arguments

in_ref_genome_fasta
Path to the reference genome fasta file used.
in_target_capture_bed
Path to a bed file containing the information on the used target capture. May also be a compressed bed.
in_word_length
Integer number defining the length of the features or motifs, e.g. 3 for tripletts or 5 for pentamers
project_folder
Path where the created files, especially the fasta file with the sequence of the target capture and the count matrices, can be stored.
in_verbose
Verbose if in_verbose=1

Value
A numeric vector with correction factors

See Also
normalizeMotifs

Examples
NULL
run_SMC

Arguments

in_stratification_lists_list

List of lists with entries from different (orthogonal) stratification axes or SMCs

in_signatures_ind_df

A data frame containing meta information about the signatures

output_path

Path to directory where the results, especially the figure produced by plot_strata is going to be stored.

in_attribute

Additional string for the file name where the figure produced by plot_strata is going to be stored.

in_remove_signature_ind

Omit one of the signatures in in_signatures_ind_df for the comparison if non-NULL. The parameter specifies the index of the signature to be removed.

in_additional_stratum

Include an additionally supplied stratum in comparison in non-NULL.

Value

The function doesn’t return any value.

See Also

plot_strata

Examples

NULL

run_SMC

`Wrapper function for the Stratification of a Mutational Catalogue`

Description

`run_SMC` takes as input a big dataframe constructed from a vcf-like file of a whole cohort. This wrapper function calls custom functions to construct a mutational catalogue and stratify it according to categories indicated by a special column in the input dataframe:

- `create_mutation_catalogue_from_df`
- `adjust_number_of_columns_in_list_of_catalogues`

This stratification yields a collection of stratified mutational catalogues, these are reformatted and sent to the custom function `SMC` and thus indirectly to `LCD_SMC` to perform a signature analysis of the stratified mutational catalogues. The result is then handed over to `plot_SMC` for visualization.
Usage

run_SMC(
  my_table,
  this_signatures_df,
  this_signatures_ind_df,
  this_subgroups_df,
  column_name,
  refGenome,
  cohort_method_flag = "all_PIDs",
  in_strata_order_ind = seq_len(length(unique(my_table[, column_name]))),
  wordLength = 3,
  verbose_flag = 1,
  target_dir = NULL,
  strata_dir = NULL,
  output_path = NULL,
  in_all_exposures_df = NULL,
  in_rownames = c(),
  in_norms = NULL,
  in_label_orientation = "turn",
  this_sum_ind = NULL
)

Arguments

my_table A big dataframe constructed from a vcf-like file of a whole cohort. The first columns are those of a standard vcf file, followed by an arbitrary number of custom or user defined columns. One of these must carry a PID (patient or sample identifyier) and one must be the category used for stratification.

this_signatures_df A numeric data frame in with n rows and l columns, n being the number of features and l being the number of signatures

this_signatures_ind_df A data frame containing meta information about the signatures

this_subgroups_df A data frame indicating which PID (patient or sample identifyier) belongs to which subgroup

column_name Name of the column in my_table which is going to be used for stratification

refGenome FaFile of the reference genome to extract the motif context of the variants in my_table

cohort_method_flag Either or several of c("all_PIDs","cohort","norm_PIDs"), representing alternative ways to average over the cohort.

in_strata_order_ind Index vector defining reordering of the strata

wordLength Integer number defining the length of the features or motifs, e.g. 3 for tripletts or 5 for pentamers
**verbose_flag**  Verbose if verbose_flag=1

**target_dir**  Path to directory where the results of the stratification procedure are going to be stored if non-NULL.

**strata_dir**  Path to directory where the mutational catalogues of the different strata are going to be stored if non-NULL.

**output_path**  Path to directory where the results, especially the figures produced by `plot_SMC` are going to be stored.

**in_all_exposures_df**  Optional argument, if specified, H, i.e. the overall exposures without stratification, is set to equal in_all_exposures_df. This is equivalent to forcing the `LCD_SMC` procedure to use e.g. the exposures of a previously performed NMF decomposition.

**in_rownames**  Optional parameter to specify rownames of the mutational catalogue V i.e. the names of the features.

**in_norms**  If specified, vector of the correction factors for every motif due to differing trinucleotide content. If null, no correction is applied.

**in_label_orientation**  Whether or not to turn the labels on the x-axis.

**this_sum_ind**  Optional set of indices for reordering the PIDs

**Value**

A list with entries exposures_list, catalogues_list, cohort and name_list.

- **exposures_list**: The list of strata specific exposures Hi, all are numerical data frames with \( l \) rows and \( m \) columns, \( l \) being the number of signatures and \( m \) being the number of samples
- **catalogues_list**: A list of strata specific cohortwide (i.e. averaged over cohort) normalized exposures
- **cohort**: subgroups_df adjusted for plotting
- **name_list**: Names of the constructed strata.

**See Also**

`create_mutation_catalogue_from_df`

`normalizeMotifs_otherRownames`

`plot_SMC`

**Examples**

```r
library(BSgenome.Hsapiens.UCSC.hg19)
data(sigs)
data(lymphoma_test)
data(lymphoma_cohort_LCD_results)
strata_list <-
cut_breaks_as_intervals(lymphoma_test_df$random_norm, in_outlier_cutoffs=c(-4,4),
```
in_cutoff_ranges_list = list(c(-2.5, -1.5),
  c(0.5, 1.5)),
in_labels = c("small", "intermediate", "big")

lymphoma_test_df$random_cat <- strata_list$category_vector
choice_ind <- (names(lymphoma_Nature2013_COSMIC_cutoff_exposures_df)
  %in% unique(lymphoma_test_df$PID))

lymphoma_test_exposures_df <-
  lymphoma_Nature2013_COSMIC_cutoff_exposures_df[, choice_ind]

temp_subgroups_df <- make_subgroups_df(lymphoma_test_df,
  lymphoma_test_exposures_df)

mut_density_list <- run_SMC(lymphoma_test_df,
  AlexCosmicValid_sig_df,
  AlexCosmicValid_sigInd_df,
  temp_subgroups_df,
  column_name = "random_cat",
  refGenome = BSgenome.Hsapiens.UCSC.hg19,
  cohort_method_flag = "norm_PIDs",
  in_rownames = rownames(AlexCosmicValid_sig_df))

---

**shapiro_if_possible**  
Wrapper for Shapiro test but allow for all identical values

**Description**
Wrapper for Shapiro test but allow for all identical values

**Usage**

```r
shapiro_if_possible(in_vector)
```

**Arguments**

- `in_vector`: Numerical vector the Shapiro-Wilk test is computed on

**Value**

p-value of the Shapiro-Wilk test, zero if all entries in the input vector `in_vector` are identical.

**See Also**

- `shapiro.test`

**Examples**

```r
shapiro_if_possible(runif(100, min=2, max=4))
shapiro_if_possible(rnorm(100, mean=5, sd=3))
shapiro_if_possible(rep(4.3, 100))
shapiro_if_possible(c("Hello", "World"))
```
Data for mutational signatures

Description

The numerical data of the mutational signatures published initially by Alexandrov et al. (Nature 2013) and Alexandrov et al., (Bioaxiv 2018) is stored in data frames with endings _sig_df, the associated meta-information is stored in data frames with endings _sigInd_df. There are several instances of _sig_df and _sigInd_df, corresponding to results and data obtained at different times and with different raw data. There always is a one-to-one correspondence between a _sig_df and a _sigInd_df. The data frames of type _sig_df have as many rows as there are features, i.e. 96 if analyzing mutational signatures of SNVs in a triplet context, and as many columns as there are signatures. Data frames of type _sigInd_df have as many rows as there are signatures in the corresponding _sig_df and several columns:

- sig: signature name
- index: corresponding to the row index of the signature
- colour: colour for visualization in stacked barplots
- process: asserted biological process
- cat.coarse: categorization of the signatures according to the asserted biological processes at low level of detail
- cat.medium: categorization of the signatures according to the asserted biological processes at intermediate level of detail
- cat.high: categorization of the signatures according to the asserted biological processes at high level of detail
- cat.putative: categorization of the signatures according to the asserted biological processes based on clustering and inference

Please note, that categorization columns are only present for the data frames corresponsing to the data from Alexandorv et al. (Nature 2013).

AlexInitialArtif_sig_df: Data frame of the signatures published initially by Alexandrov et al. (Nature 2013). There are 27 signatures which constitute the columns, 22 of which were validated by an orthogonal sequencing technology. These 22 are in the first 22 columns of the data frame. The column names are A pasted to the number of the signature, e.g. A5. The nonvalidated signatures have an additional letter in their naming convention: either AR1 - AR3 or AU1 - AU2. The rownames are the features, i.e. an encoding of the nucleotide exchanges in their trinucleotide context, e.g. C>A ACA. In total there are 96 different features and therefore 96 rows when dealing with a trinucleotide context.

AlexInitialArtif_sigInd_df: Meta-information for AlexInitialArtif_sig_df

AlexInitialValid_sig_df: Data frame of only the validated signatures published initially by Alexandrov et al. (Nature 2013), corresponding to the first 22 columns of AlexInitialArtif_sig_df

AlexInitialValid_sigInd_df: Meta-information for AlexInitialValid_sig_df

AlexCosmicValid_sig_df: Data frame of the updated signatures list maintained by Ludmil Alexandrov at https://cancer.sanger.ac.uk/cosmic/signatures. The column names are AC pasted
to the number of the signature, e.g. AC5. The naming convention for the rows is as described for 
AlexInitialArtif_sig_df.

AlexCosmicValid_sigInd_df: Meta-information for AlexCosmicValid_sig_df

AlexCosmicArtif_sig_df: Data frame of the updated signatures list maintained by Ludmil Alexandrov at https://cancer.sanger.ac.uk/cosmic/signatures and complemented by the artifact signatures from the initial publication, i.e. the last 5 columns of AlexInitialArtif_sig_df. The column names are AC pasted to the number of the signature, e.g. AC5. The naming convention for the rows is as described for AlexInitialArtif_sig_df.

AlexCosmicArtif_sigInd_df: Meta-information for AlexCosmicArtif_sig_df

Usage

data(sigs)

Author(s)

Daniel Huebschmann <huebschmann.daniel@googlemail.com>

Source

AlexInitial: ftp://ftp.sanger.ac.uk/pub/cancer/AlexandrovEtAl/signatures.txt

AlexCosmic: https://cancer.sanger.ac.uk/cancergenome/assets/signatures_probabilities.txt

References

Alexandrov et al. (Nature 2013)

Data for PCAWG SNV signatures (COSMIC v3), including artifacts.

PCAWG_SP_SBS_sigs_Artif_df: Data frame of the signatures published by Alexandrov et al. (Biorxiv 2013) which were decomposed with the method SigProfiler. SNV signatures are labeled with SBS, single base signature. There are 67 signatures which constitute the columns, 47 of which were validated by a bayesian NFM method, SignatureAnalyzer. Validated signatures are SBS1-SBS26, SBS28-SBS42 and SBS44. SBS7 is split up into 7 a/b/c and d. SBS10 ans SBS17 are both split up into a and b. Resulting in a 47 validated signatures. Please note, unlike the paper by Alexandrov et al. (Biorxiv 2018) the data sets do not contain a SBS84 and SBS85 as not all were availiable to perform supervised signature analysis. In total there are 96 different features and therefore 96 rows when dealing with a trinucleotide context.
Description

PCAWG_SP_SBS_sigInd_Artif_df: Meta-information for PCAWG_SP_SBS_sigs_Artif_df

PCAWG_SP_SBS_sigs_Real_df: Data frame of only the validated signatures published by Alexandrov et al. (Biorxiv 2018), corresponding to the column 1-26, 28-42 and 44 of the PCAWG_SP_SBS_sigs_Artif_df data frame

PCAWG_SP_SBS_sigInd_Real_df: Meta-information for PCAWG_SP_SBS_sigs_Real_df

PCAWG_SP_ID_sigs_df: Data frame with Indel signatures published by Alexandrov et al. (Biorxiv 2018) which were decomposed with the method SigProfiler. There are 17 signatures reported but as supervised signatures are only valid for whole genome sequencing data analysis. In whole genome sequencing data the Indel signature ID15 was not described and thus is not part of this data set. In total 83 features are described. The categorisation considers the size of the insertion and deletion, the motif, and the sequence context. Hereby the number of repetition or partial repetition of the motif is determined.

PCAWG_SP_ID_sigInd_df: Meta-information for PCAWG_SP_ID_sigs_df

Usage

data(sigs_pcawg)

Author(s)

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Source

PCAWG_SNV: https://www.synapse.org/#!Synapse:syn11738319
PCAWG_INDEL: https://cancer.sanger.ac.uk/cosmic/signatures/ID

References

Alexandrov et al. (Biorxiv 2018)

SMC

Stratification of a Mutational Catalogue

Description

SMC takes a given collection of stratified mutational catalogues \( V_i \), sends them to perform a mutational signatures decomposition by Linear Combination Decomposition (LCD) with the functions \( \text{LCD}_\text{SMC} \) with known signatures \( W \). It subsequently performs some useful statistics and preparation for plotting with the function \( \text{plot}_\text{SMC} \). SMC is naturally called by \( \text{run}_\text{SMC} \).
Usage

```r
SMC(
    df_list,
    this_signatures_df,
    in_all_exposures_df,
    number_of_strata,
    number_of_sigs,
    name_list,
    this_subgroups_df,
    mutation_catalogue_all_df,
    cohort_method_flag,
    inVerbose = 1
)
```

Arguments

- `df_list` A list of s stratified mutational catalogues $V_i$ (numeric data frames) with $n$ rows and $m$ columns each, $n$ being the number of features and $m$ being the number of samples. This list is naturally provided in `run_SMC`.

- `this_signatures_df` A numeric data frame $W$ in with $n$ rows and $l$ columns, $n$ being the number of features and $l$ being the number of signatures.

- `in_all_exposures_df` The overall exposures $H$ without stratification, a numeric data frame with $l$ rows and $m$ columns, $l$ being the number of signatures and $m$ being the number of samples.

- `number_of_strata` The length of the list `df_list`.

- `number_of_sigs` The number of signatures used in the current decomposition.

- `name_list` A list of names of the different strata.

- `this_subgroups_df` A data frame indicating which PID (patient or sample identifier) belongs to which subgroup.

- `mutation_catalogue_all_df` The overall mutational catalogue $V$ without stratification.

- `cohort_method_flag` Either or several of c("all_PIDs","cohort","norm_PIDs"), representing alternative ways to average over the cohort.

- `inVerbose` Verbose if `inVerbose=1`.

Value

A list with entries `exposures_strata_list`, `exposures_both_rel_df_list`, `this_subgroups_df`, `subgroup_ind` and `decomposition_method`.

- `exposures_strata_list`: The list of s strata specific exposures $H_i$, all are numerical data frames with $l$ rows and $m$ columns, $l$ being the number of signatures and $m$ being the number of samples.
SMC_perPID

- exposures_both_rel_df_list: A list of strata specific cohortwide (i.e. averaged over cohort) normalized exposures
- this_subgroups_df: subgroups_df adjusted for plotting
- subgroup_ind: Index of the subgroups chosen and relevant for plotting.
- decomposition_method: String telling whether LCD or NMF was used, relevant only for handing over to plot_SMC.

See Also
run_SMC
plot_SMC
LCD_SMC

Examples
NULL

SMC_perPID

Run SMC at a per sample level

Description
Run an SMC analysis (stratification of the mutational catalogue) at per sample / per-PID level, corresponding to a divide and conquer strategy. For every single PID, only those signatures actually present in this PID will be provided for the SMC analysis.

Usage
SMC_perPID(
  in_dfList,
  in_LCDlist,
  in_subgroups_df,
  in_save_plot = TRUE,
  in_save_dir = NULL,
  in_save_name = "KataegisSMCs.pdf",
  in_verbose_flag = 0,
  ...
)

Arguments
in_dfList  Named list of vcf-like data frames, one entry per sample/PID of a cohort.
in_LCDlist  Output of an LCD list performed on the above cohort, carrying notably information on the exposures (in_LCDlist$exposures), the present signatures (in_LCDlist$signatures) and meta information about the signatures(in_LCDlist$out_sig_ind_df).
### split_exposures_by_subgroups

**Description**

If a cohort consists of different subgroups, this function enables to split the data frame storing the signature exposures into a list of data frames with signature exposures, one per subgroup. This functionality is needed for `stat_test_subgroups` and `stat_plot_subgroups`.

**Usage**

```r
split_exposures_by_subgroups(
  in_exposures_df,
  in_subgroups_df,
  in_subgroups.field = "subgroup",
  in_PID.field = "PID"
)
```

**Arguments**

- `in_exposures_df`:
  Numerical data frame of the exposures (i.e. contributions of the different signatures to the number of point mutations per PID).

- `in_subgroups_df`:
  Data frame indicating which PID belongs to which subgroup.

---

**in_subgroups_df**
Data frame with subgroup information about the PIDs in the above mentioned cohort.

**in_save_plot**
Boolean flag to indicate whether per-PID plots should be saved.

**in_save_dir**
If per-PID plots are to be saved, this is the path where to save them.

**in_save_name**
Suffix to be appended to the sample name to generate the name of the saved per-PID plots.

**in_verbose_flag**
Whether to run verbose (1) or not (0).

... Data passed on to `run_SMC`.

**Value**

A list of lists. The top level is a named per-PID list, each entry is of type SMClst (cf. `run_SMC`).

**Examples**

```r
NULL
```
**stat_plot_subgroups**

- **in_subgroups.field** Name indicating which column in `in_subgroups_df` contains the subgroup information
- **in_PID.field** Name indicating which column in `in_subgroups_df` contains the PID information

**Value**

List of data frames with the subgroup specific signature exposures.

**See Also**

- `stat_test_subgroups`
- `stat_plot_subgroups`

**Examples**

```r
NULL
```

---

**stat_plot_subgroups**  
*Plot averaged signature exposures per subgroup*

**Description**

Plot one averaged signature exposure pattern per subgroup. Uses `split_exposures_by_subgroups`.

**Usage**

```r
stat_plot_subgroups(
  in_exposures_df, 
  in_subgroups_df, 
  in_signatures_ind_df, 
  in_subgroups.field = "subgroup", 
  in_PID.field = "PID", 
  in_colour_vector = NULL
)
```

**Arguments**

- **in_exposures_df**  
  Numerical data frame of the exposures (i.e. contributions of the different signatures to the number of point mutations per PID)

- **in_subgroups_df**  
  Data frame indicating which PID belongs to which subgroup

- **in_signatures_ind_df**  
  Data frame carrying additional information on the signatures
**stat_test_SMC**

```r
stat_test_SMC(in_strat_list, in_flag = "norm")
```

**Arguments**

- **in_strat_list** A list with entries `exposures_list`, `catalogues_list`, `cohort` and `name_list` as in the output of `run_SMC`:
  - `exposures_list`: The list of s strata specific exposures Hi, all are numerical data frames with l rows and m columns, l being the number of signatures and m being the number of samples
  - `catalogues_list`: A list of s strata specific cohortwide (i.e. averaged over cohort) normalized exposures
  - `cohort`: subgroups_df adjusted for plotting
  - `name_list`: Names of the contracted strata.

- **in_flag** If "norm", all tests are performed on normalized exposures, otherwise the absolute exposures are taken.

**Description**

`stat_test_SMC` tests for enrichment or depletion in the different strata of a stratification of the mutational catalogue for every signature independently by applying Kruskal Wallis tests. For those signatures where the Kruskal Wallis test gives a significant p-value, pairwise posthoc tests are carried out by calling `kwAllPairsNemenyiTest`. Additionally all data is tested for normality by Shapiro Wilk tests, so that the user may apply ANOVA and pairwise posthoc t-test where allowed.

**Usage**

`stat_test_SMC(in_strat_list, in_flag = "norm")`

**Examples**

NULL

**Value**

The function doesn't return any value, it plots instead.

**See Also**

`split_exposures_by_subgroups`

### in_subgroups.field

Name indicating which column in `in_subgroups_df` contains the subgroup information

### in_PID.field

Name indicating which column in `in_subgroups_df` contains the PID information

### in_colour_vector

If non-null, specifies the colours attributed to the subgroups
Value

A list with entries `kruskal_df`, `shapiro_df`, `kruskal_posthoc_list`,

- `kruskal_df`: A data frame containing results (statistic and p values) of the Kruskal Wallis tests (tests for enrichment or depletion in the different strata for every signature independently).
- `shapiro_df`: A data frame containing results (p values) of the Shapiro Wilk tests (tests for normal distribution in the different strata for every signature independently).
- `kruskal_posthoc_list`: A list of results of pairwise posthoc tests carried out for those signatures where the Kruskal Wallis test yielded a significant p-value (carried out by `kwAllPairsNemenyiTest`).

See Also

`run_SMC`
`kwAllPairsNemenyiTest`
`kruskal.test`
`shapiro_if_possible`
`shapiro.test`

Examples

NULL

---

**stat_test_subgroups**  
*Test for differences in average signature exposures between subgroups*

Description

Apply Kruskal-Wallis tests to detect differences in the signature exposures between different subgroups. Uses `split_exposures_by_subgroups`. Algorithm analogous to `stat_test_SMC`.

Usage

```r
stat_test_subgroups(
  in_exposures_df, 
  in_subgroups_df, 
  in_subgroups.field = "subgroup", 
  in_PID.field = "PID"
)
```
Arguments

- `in_exposures_df`: Numerical data frame of the exposures (i.e. contributions of the different signatures to the number of point mutations per PID)
- `in_subgroups_df`: Data frame indicating which PID belongs to which subgroup
- `in_subgroups.field`: Name indicating which column in `in_subgroups_df` contains the subgroup information
- `in_PID.field`: Name indicating which column in `in_subgroups_df` contains the PID information

Value

A list with entries `kruskal_df`, `kruskal_posthoc_list`,

- `kruskal_df`: A data frame containing results (statistic and p values) of the Kruskal Wallis tests (tests for enrichment or depletion in the different strata for every signature independently).
- `kruskal_posthoc_list`: A list of results of pairwise posthoc tests carried out for those signatures where the Kruskal Wallis test yielded a significant p-value (carried out by `kwAllPairsNemenyiTest`).

See Also

- `split_exposures_by_subgroups`
- `stat_test_SMC`
- `kwAllPairsNemenyiTest`
- `kruskal.test`

Examples

```r
NULL
```

---

**stderrmean**

*Compute the standard error of the mean*

Description

This function returns the standard deviation of an input numerical vector divided by the square root of the length of the input vector

Usage

```r
stderrmean(x)
```
Arguments

x  A numerical vector

Value

Standard deviation of an input numerical vector divided by the square root of the length of the input vector

Examples

A <- c(1,2,3)
sd(A)
stderrmean(A)

sum_over_list_of_df  Elementwise sum over a list of (numerical) data frames

Description

Elementwise sum over a list of (numerical) data frames

Usage

sum_over_list_of_df(in_df_list)

Arguments

in_df_list  List of (numerical) data frames

Value

A numerical data frame with the same dimensions as the entries of in_df_list with elementwise sums

Examples

A <- data.frame(matrix(c(1,1,1,2,2,2),ncol=2))
B <- data.frame(matrix(c(3,3,3,4,4,4),ncol=2))
df_list <- list(A=A,B=B)
sum_over_list_of_df(df_list)
targetCapture_cor_factors

Description

List of lists with correction factors for different target capture kits. The elements of the overall list are lists, every one carrying information for one target capture kit (and named after it). The elements of these sublists are 64 dimensional vectors with correction factors for all triplets. They were computed using counts of occurrence of the respective triplets in the target capture and in the reference genome and making ratios (either for the counts themselves as in abs_cor or for the relative occurrences in rel_cor). The information in this data structure may be used as input to normalizeMotifs_otherRownames.

Usage

data(targetCapture_cor_factors)

Value

A list of lists of data frames

Author(s)

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testSigs

Description

Test for significance of alternative models cohort wide

Usage

testSigs(
  in_catalogue_df,  
in_sig_df,  
in_exposures_df,  
in_factor = 0,  
in_pdf = NULL
)
**Arguments**

- `in_catalogue_df`: Input numerical data frame of the mutational catalog of the cohort to be analyzed.
- `in_sig_df`: Numerical data frame of the signatures used for analysis.
- `in_exposures_df`: Input numerical data frame of the exposures computed for the cohort to be analyzed.
- `in_factor`: Deviation factor of the altered alternative model.
- `in_pdf`: Probability distribution function, parameter passed on to `confIntExp` if NULL assumed to be normal distribution.

**Value**

Returns a data frame.

**Examples**

```r
NULL
```

---

**Description**

Test significance of association between a vector of exposures and a selection of samples, e.g. those affected by mutations in a pathway as returned by `find_affected_PIDs`.

**Usage**

```r
test_exposureAffected(
    in_exposure_vector,
    in_affected_PIDs,
    in_mutation_label = NULL,
    in_exposure_label = NULL
)
```

**Arguments**

- `in_exposure_vector`: Named vector of a phenotype (e.g. exposures to a specific signature).
- `in_affected_PIDs`: Character vector of samples affected by some criterion, e.g. mutations in a pathway as returned by `find_affected_PIDs`.
- `in_mutation_label`: If non-NULL, prefix to the mutation status (x-axis label) in the produced boxplot.
- `in_exposure_label`: If non-NULL, prefix to the exposures (y-axis label) in the produced boxplot.
**Value**

A list with entries:

- current_kruskal: Kruskal test object from testing phenotype against affection
- current_boxplot: Boxplot of phenotype against affection

**Examples**

NULL

---

test_gene_list_in_exposures

*Test if mutated PID\*s are enriched in signatures*

---

**Description**

For all signatures found in a project, this function tests whether PID\*s having mutations in a specified list of genes of interest have significantly higher exposures.

**Usage**

```r
test_gene_list_in_exposures(
  in_gene_list,
  in_exposure_df,
  in_mut_table,
  in_gene.field = "GENE_short",
  in_p_cutoff = 0.05
)
```

**Arguments**

- `in_gene_list`: List with genes of interest
- `in_exposure_df`: Data frame with the signature exposures
- `in_mut_table`: Data frame or table of mutations (derived from vcf-format)
- `in_gene.field`: Name of the column in which the gene names are to be looked up
- `in_p_cutoff`: Significance threshold

**Value**

A list with entries `pvals`, `exposure_df`, `number_of_mutated`,

- `pvals`: p-values of the t-tests performed on mutated vs. unmutated PID\*s
- `exposure_df`: Transposed input exposures data frame with additional annotations for mutation status
- `number_of_mutated`: Number of PID\*s carrying a mutation
transform_rownames_R_to_MATLAB

Change rownames from one naming convention to another

Description
Rownames or names of the features used differ between the different contexts a signature analysis is carried out in. The function `transform_rownames_R_to_MATLAB` changes from the convention used in the Y APSA package to the one used by Alexandrov et al. in the MATLAB framework.
The function `transform_rownames_MATLAB_to_R` changes from the convention used in Alexandrov et al. in the MATLAB framework to the one used by the Y APSA package.
The function `transform_rownames_MATLAB_to_R` changes from the convention used in stored mutational catalogues by Alexandrov et al. to the one used by the Y APSA package.
The function `transform_rownames_YAPSA_to_deconstructSigs` changes from the convention used in the Y APSA package to the one used by the deconstructSigs package.
The function `transform_rownames_YAPSA_to_deconstructSigs` changes from the convention used in the deconstructSigs package to the one used by the Y APSA package.

Usage

```r
transform_rownames_R_to_MATLAB(in_rownames, wordLength = 3)
transform_rownames_MATLAB_to_R(in_rownames, wordLength = 3)
transform_rownames_nature_to_R(in_rownames, wordLength = 3)
transform_rownames_YAPSA_to_deconstructSigs(in_rownames, wordLength = 3)
transform_rownames_deconstructSigs_to_YAPSA(in_rownames, wordLength = 3)
```

Arguments

- `in_rownames`: Character vector of input rownames
- `wordLength`: Size of the considered motif context

Value
A character vector of the translated rownames.

Examples

```r
NULL
```
translate_to_hg19 

Translate chromosome names to the hg19 naming convention

Description

translate_to_hg19: In hg19 naming convention, chromosome names start with the prefix chr and the gonosomes are called X and Y. If data analysis is performed e.g. with BSgenome.Hsapiens.UCSC.hg19, this naming convention is needed. The inverse transform is done with translate_to_1kG.

translate_to_1kG: In 1kG, i.e. 1000 genomes naming convention, chromosome names have no prefix chr and the gonosomes are called 23 for X and 24 for Y. If data analysis is performed e.g. with hs37d5.fa, this naming convention is needed. The inverse transform is done with translate_to_hg19.

Usage

translate_to_hg19(in_dat, in_CHROM.field = "CHROM", in_verbose = FALSE)
translate_to_1kG(in_dat, in_CHROM.field = "chr", in_verbose = FALSE)

Arguments

in_dat 
GRanges object, VRanges object or data frame which carries one column with chromosome information to be reformatted.
in_CHROM.field 
String indicating which column of in_dat carries the chromosome information
in_verbose 
Whether verbose or not.

Value

GRanges object, VRanges object or data frame identical to in_dat, but with the names in the chromosome column replaced (if dealing with data frames) or alternatively the seqlevels replaced (if dealing with GRanges or VRanges objects).

Examples

test_df <- data.frame(CHROM=c(1,2,23,24),POS=c(100,120000000,30000,25000),
  dummy=c("a","b","c","d"))
hg19_df <- translate_to_hg19(test_df, in_CHROM.field = "CHROM")
hg19_df

test_df <- data.frame(CHROM=c(1,2,23,24),POS=c(100,120000000,30000,25000),
  dummy=c("a","b","c","d"))
hg19_df <- translate_to_hg19(test_df, in_CHROM.field = "CHROM")
onekG_df <- translate_to_1kG(hg19_df, in_CHROM.field = "CHROM")
onekG_df
Create a rainfall plot in a trellis structure

Description

A trellis is a plot structure which allows space optimized multi-panel multi track plots. This function uses the package \texttt{gtrellis} developed by Zuguang Gu, also available at \url{https://www.bioconductor.org/packages/release/bioc/html/gtrellis.html}. The graphics in the tracks within a gtrellis plot are mostly drawn with functions from the package \texttt{grid}. Note that for technical reasons, the column indicating the chromosome MUST have the name \textit{chr} and be the first column in the data frame supplied to the gtrellis functions. Therefore reformatting is performed in this function before calling gtrellis functions.

Usage

\begin{verbatim}
 trellis_rainfall_plot(
    in_rainfall_dat,
    in_point_size = unit(1, "mm"),
    in_rect_list = NULL,
    in_title = "",
    in_CHROM.field = "CHROM",
    in_POS.field = "POS",
    in_dist.field = "dist",
    in_col.field = "col"
)
\end{verbatim}

Arguments

\begin{itemize}
  \item \textit{in_rainfall_dat} \hspace*{1cm} Data frame which has to contain at least columns for chromosome, position, intermutational distance and colour information
  \item \textit{in_point_size} \hspace*{1cm} size of the points in the rainfall plot to be created has to be provided with appropriate units, e.g. \texttt{in_point_size=unit(0.5,"mm")}
  \item \textit{in_rect_list} \hspace*{1cm} Optional argument, if present, will lead to highlighting of specified regions by coloured but transparent rectangles
  \item \textit{in_title} \hspace*{1cm} Title in the figure to be created.
  \item \textit{in_CHROM.field} \hspace*{1cm} String indicating which column of \texttt{in_rainfall_dat} carries the chromosome information
  \item \textit{in_POS.field} \hspace*{1cm} String indicating which column of \texttt{in_rainfall_dat} carries the position information
  \item \textit{in_dist.field} \hspace*{1cm} String indicating which column of \texttt{in_rainfall_dat} carries the intermutational distance information
  \item \textit{in_col.field} \hspace*{1cm} String indicating which column of \texttt{in_rainfall_dat} carries the colour information encoding the nucleotide exchange
\end{itemize}
Value

The function doesn’t return any value.
The function doesn’t return any value.

See Also

gtrellis_layout
add_track
grid.points

Examples

data(lymphoma_test)
choice_PID <- "4121361"
PID_df <- subset(lymphoma_test,PID==choice_PID)
trellis_rainfall_plot(PID_df,in_point_size=unit(0.5,"mm"))

variateExp

Wrapper to compute confidence intervals for a cohort

Description

Wrapper function around confIntExp, which is applied to every signature/sample pair in a cohort. The extracted upper and lower bounds of the confidence intervals are added to the input data which is reordered and melted in order to prepare for visualization with ggplot2.

Usage

variateExp(
  in_catalogue_df,
  in_sig_df,
  in_exposures_df,
  in_sigLevel = 0.05,
  in_delta = 0.4,
  in_pdf = NULL
)

Arguments

in_catalogue_df Input numerical data frame of the mutational catalog of the cohort to be analyzed.
in_sig_df Numerical data frame of the signatures used for analysis.
variateExp

in_exposures_df
Input numerical data frame of the exposures computed for the cohort to be analyzed.

in_sigLevel
Significance level, parameter passed to confIntExp.

in_delta
Inflation parameter for the alternative model, parameter passed on to confIntExp

in_pdf
Probability distribution function, parameter passed on to confIntExp, if NULL assumed to be normal distribution.

Value
A melted data frame.

Examples

library(BSgenome.Hsapiens.UCSC.hg19)
data(lymphoma_test)
data(lymphoma_cohort_LCD_results)
data(sigs)
word_length <- 3
temp_list <- create_mutation_catalogue_from_df(
  lymphoma_test_df, this_seqnames.field = "CHROM",
  this_start.field = "POS", this_end.field = "POS",
  this_PID.field = "PID", this_subgroup.field = "SUBGROUP",
  this_refGenome = BSgenome.Hsapiens.UCSC.hg19,
  this_wordLength = word_length)
lymphoma_catalogue_df <- temp_list$matrix
lymphoma_PIDs <- colnames(lymphoma_catalogue_df)
data("lymphoma_cohort_LCD_results")
lymphoma_exposures_df <-
  lymphoma_Nature2013_COSMIC_cutoff_exposures_df[, lymphoma_PIDs]
lymphoma_sigs <- rownames(lymphoma_exposures_df)
lymphoma_sig_df <- AlexCosmicValid_sig_df[, lymphoma_sigs]
lymphoma_complete_df <- variateExp(in_catalogue_df = lymphoma_catalogue_df,
  in_sig_df = lymphoma_sig_df,
  in_exposures_df = lymphoma_exposures_df,
  in_sigLevel = 0.025, in_delta = 0.4)
head(lymphoma_complete_df)
lymphoma_complete_df$sample <-
  factor(lymphoma_complete_df$sample,
    levels = colnames(lymphoma_exposures_df)[
    order(colSums(lymphoma_exposures_df), decreasing = TRUE)])
sig_colour_vector <- c("black", AlexCosmicValid_sigInd_df$colour)
names(sig_colour_vector) <-
  c("total", as.character(AlexCosmicValid_sigInd_df$sig))
ggplot(data = lymphoma_complete_df,
  aes(x = sample, y = exposure, fill = sig)) +
geom_bar(stat = "identity") +
geom_errorbar(aes(ymin = lower, ymax = upper), width = 0.2) +
facet_wrap(~sig, nrow = nrow(lymphoma_exposures_df) + 1) +
theme_grey() +
theme(panel.border = element_rect(fill = NA, colour = "black"),
variateExpSingle

Wrapper for the likelihood ratio test

Description

Application of the likelihood ratio test to mutational signatures, primarily for one single sample.

Usage

variateExpSingle(
  in_catalogue_vector,
  in_sig_df,
  in_exposure_vector,
  in_ind,
  in_factor = 1,
  in_pdf = NULL,
  verbose = FALSE
)

Arguments

in_catalogue_vector
  Mutational catalog of the input sample.

in_sig_df
  Data frame encoding the signatures used for the analysis.

in_exposure_vector
  Exposure vector computed for the input sample.

in_ind
  Index specifying which signature among in_sig_df is to be tested.

in_factor
  Deviation factor of the altered alternative model.

in_pdf
  Probability distribution function, parameter passed on to logLikelihood and later to computeLogLik.

verbose
  Verbose if in_verbose=1

Value

Returns a list
Examples

```r
library(BSgenome.Hsapiens.UCSC.hg19)
data(lymphoma_test)
data(lymphoma_cohort_LCD_results)
data(sigs)
word_length <- 3
temp_list <- create_mutation_catalogue_from_df(
  lymphoma_test_df,
  this_seqnames.field = "CHROM",
  this_start.field = "POS", this_end.field = "POS",
  this_PID.field = "PID", this_subgroup.field = "SUBGROUP",
  this_refGenome = BSgenome.Hsapiens.UCSC.hg19,
  this_wordLength = word_length)
lymphoma_catalogue_df <- temp_list$matrix
lymphoma_PIDs <- colnames(lymphoma_catalogue_df)
data("lymphoma_cohort_LCD_results")
lymphoma_exposures_df <-
  lymphoma_Nature2013_COSMIC_cutoff_exposures_df[, lymphoma_PIDs]
lymphoma_sigs <- rownames(lymphoma_exposures_df)
lymphoma_sig_df <- AlexCosmicValid_sig_df[, lymphoma_sigs]
variateExpSingle(
  in_ind = 1,
  in_factor = 1.5,
  in_catalogue_vector = lymphoma_catalogue_df[, 1],
  in_sig_df = lymphoma_sig_df,
  in_exposure_vector = lymphoma_exposures_df[, 1])
```

Description

Yet Another Package for mutational Signature analysis

Details

This package provides functions and routines useful in the analysis of mutational signatures (cf. L. Alexandrov et al., Nature 2013). In particular, functions to perform a signature analysis with known signatures (LCD = linear combination decomposition) and a signature analysis on stratified mutational catalogue (run_SMC = stratify mutational catalogue) are provided.
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