

Package ‘Voyager’

February 7, 2023

Type Package

Title From geospatial to spatial omics

Version 1.0.9

Description SpatialFeatureExperiment (SFE) is a new S4 class for working with spatial single-cell genomics data. The voyager package implements basic exploratory spatial data analysis (ESDA) methods for SFE. This first version supports univariate global spatial ESDA methods such as Moran's I, permutation testing for Moran's I, and correlograms. The Voyager package also implements plotting functions to plot SFE data and ESDA results. Multivariate ESDA and univariate local metrics will be added in later versions.

Imports BiocParallel, bluster, ggnewscale, ggplot2, Matrix, methods, patchwork, rlang, S4Vectors, scales, scico, sf, SingleCellExperiment, SpatialExperiment, SpatialFeatureExperiment, spdep, stats, SummarizedExperiment

Suggests BiocSingular, BiocStyle, cowplot, dbscan, ExperimentHub, hexbin, knitr, rmarkdown, scater, scattermore, scan, SFEData, sparseMatrixStats, testthat (>= 3.0.0), vdiffr

License Artistic-2.0

Encoding UTF-8

RoxygenNote 7.2.3

Config/testthat/edition 3

Depends R (>= 4.2.0)

biocViews GeneExpression, Spatial, Transcriptomics, Visualization

VignetteBuilder knitr

URL <https://github.com/pachterlab/voyager>

BugReports <https://github.com/pachterlab/voyager/issues>

git_url <https://git.bioconductor.org/packages/Voyager>

git_branch RELEASE_3_16

git_last_commit 3802c86

git_last_commit_date 2023-02-04

Date/Publication 2023-02-06

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calculateUnivariate	<i>Univariate spatial statistics</i>
---------------------	--------------------------------------

Description

These functions compute univariate spatial statistics, both global and local, on matrices, data frames, and SFE objects. For SFE objects, the statistics can be computed for numeric columns of `colData`, `colGeometries`, and `annotGeometries`, and the results are stored within the SFE object. `calculateMoransI` and `runMoransI` are convenience wrappers for `calculateUnivariate` and `runUnivariate` respectively.

Usage

```
## S4 method for signature 'ANY'
calculateUnivariate(
  x,
  listw,
```

```

type = c("moran", "geary", "moran.mc", "geary.mc", "moran.test", "geary.test",
"globalG.test", "sp.correlogram", "moran.plot", "localmoran", "localmoran_perm",
"localC", "localC_perm", "localG", "localG_perm", "LOSH", "LOSH.mc", "LOSH.cs",
"gwss"),
BPPARAM = SerialParam(),
zero.policy = NULL,
returnDF = TRUE,
p.adjust.method = "BH",
...
)

## S4 method for signature 'SpatialFeatureExperiment'
calculateUnivariate(
  x,
  type,
  features = NULL,
  colGraphName = 1L,
  sample_id = NULL,
  exprs_values = "logcounts",
  BPPARAM = SerialParam(),
  zero.policy = NULL,
  returnDF = TRUE,
  include_self = FALSE,
  p.adjust.method = "BH",
  ...
)

## S4 method for signature 'ANY'
calculateMoransI(x, ..., BPPARAM = SerialParam(), zero.policy = NULL)

## S4 method for signature 'SpatialFeatureExperiment'
calculateMoransI(
  x,
  features = NULL,
  colGraphName = 1L,
  sample_id = NULL,
  exprs_values = "logcounts",
  BPPARAM = SerialParam(),
  zero.policy = NULL,
  returnDF = TRUE,
  include_self = FALSE,
  p.adjust.method = "BH",
  ...
)

colDataUnivariate(
  x,
  type,

```

```
    features,  
    colGraphName = 1L,  
    sample_id = NULL,  
    BPPARAM = SerialParam(),  
    zero.policy = NULL,  
    include_self = FALSE,  
    p.adjust.method = "BH",  
    ...  
  )  
  
colDataMoransI(  
  x,  
  features,  
  colGraphName = 1L,  
  sample_id = NULL,  
  BPPARAM = SerialParam(),  
  zero.policy = NULL,  
  include_self = FALSE,  
  p.adjust.method = "BH",  
  ...  
)  
  
colGeometryUnivariate(  
  x,  
  type,  
  features,  
  colGeometryName = 1L,  
  colGraphName = 1L,  
  sample_id = NULL,  
  BPPARAM = SerialParam(),  
  zero.policy = NULL,  
  include_self = FALSE,  
  p.adjust.method = "BH",  
  ...  
)  
  
colGeometryMoransI(  
  x,  
  features,  
  colGeometryName = 1L,  
  colGraphName = 1L,  
  sample_id = NULL,  
  BPPARAM = SerialParam(),  
  zero.policy = NULL,  
  include_self = FALSE,  
  p.adjust.method = "BH",  
  ...  
)
```

```
annotGeometryUnivariate(  
  x,  
  type,  
  features,  
  annotGeometryName = 1L,  
  annotGraphName = 1L,  
  sample_id = NULL,  
  BPPARAM = SerialParam(),  
  zero.policy = NULL,  
  include_self = FALSE,  
  p.adjust.method = "BH",  
  ...  
)  
  
annotGeometryMoransI(  
  x,  
  features,  
  annotGeometryName = 1L,  
  annotGraphName = 1L,  
  sample_id = NULL,  
  BPPARAM = SerialParam(),  
  zero.policy = NULL,  
  include_self = FALSE,  
  p.adjust.method = "BH",  
  ...  
)  
  
runUnivariate(  
  x,  
  type,  
  features = NULL,  
  colGraphName = 1L,  
  sample_id = NULL,  
  exprs_values = "logcounts",  
  BPPARAM = SerialParam(),  
  zero.policy = NULL,  
  include_self = FALSE,  
  p.adjust.method = "BH",  
  ...  
)  
  
runMoransI(  
  x,  
  features = NULL,  
  colGraphName = 1L,  
  sample_id = NULL,  
  exprs_values = "logcounts",
```

```

BPPARAM = SerialParam(),
zero.policy = NULL,
include_self = FALSE,
p.adjust.method = "BH",
...
)

```

Arguments

x	A numeric matrix whose rows are features/genes, or a <code>SpatialFeatureExperiment</code> (SFE) object with such a matrix in an assay.
listw	Weighted neighborhood graph as a <code>spdep listw</code> object.
type	An integer specifying the index or string specifying the name of the <code>*Geometry</code> to query or replace. If missing, then the first item in the <code>*Geometries</code> will be returned or replaced.
BPPARAM	A <code>BiocParallelParam</code> object specifying whether and how computing the metric for numerous genes shall be parallelized.
zero.policy	default <code>NULL</code> , use global option value; if <code>TRUE</code> assign zero to the lagged value of zones without neighbours, if <code>FALSE</code> assign <code>NA</code>
returnDF	Logical, when the results are not added to a SFE object, whether the results should be formatted as a <code>DataFrame</code> .
p.adjust.method	Method to correct for multiple testing, passed to <code>p.adjustSP</code> . Methods allowed are in <code>p.adjust.methods</code> .
...	Other arguments passed to S4 method (for convenience wrappers like <code>calculateMoransI</code>) or method used to compute metrics as specified by the argument <code>type</code> (as in more general functions like <code>calculateUnivariate</code>). See documentation in the <code>spdep</code> package for the latter.
features	Genes (<code>calculate*</code> SFE method and <code>run*</code>) or numeric columns of <code>colData(x)</code> (<code>colData*</code>) or any <code>colGeometry</code> (<code>colGeometry*</code>) or <code>annotGeometry</code> (<code>annotGeometry*</code>) for which the univariate metric is to be computed. Default to <code>NULL</code> . When <code>NULL</code> , then the metric is computed for all genes with the values in the assay specified in the argument <code>exprs_values</code> . This can be parallelized with the argument <code>BPPARAM</code> . For genes, if the column "symbol" is present in <code>rowData</code> and the row names of the SFE object are Ensembl IDs, then the gene symbol can be used and converted to IDs behind the scene. However, if one symbol matches multiple IDs, a warning will be given and the first match will be used. Internally, the results are always stored by the Ensembl ID rather than symbol.
colGraphName	Name of the <code>listw</code> graph in the SFE object that corresponds to entities represented by columns of the gene count matrix. Use <code>colGraphNames</code> to look up names of the available graphs for cells/spots. Note that for multiple <code>sample_ids</code> , it is assumed that all of them have a graph of this same name.
sample_id	Sample(s) in the SFE object whose cells/spots to use. Can be "all" to compute metric for all samples; the metric is computed separately for each sample.
exprs_values	Integer scalar or string indicating which assay of <code>x</code> contains the expression values.

include_self	Logical, whether the spatial neighborhood graph should include edges from each location to itself. This is for Getis-Ord G_i^* as in <code>localG</code> and <code>localG_perm</code> , not to be used for any other method.
colGeometryName	Name of a <code>colGeometry</code> <code>sf</code> data frame whose numeric columns of interest are to be used to compute the metric. Use <code>colGeometryNames</code> to look up names of the <code>sf</code> data frames associated with cells/spots.
annotGeometryName	Name of a <code>annotGeometry</code> <code>sf</code> data frame whose numeric columns of interest are to be used to compute the metric. Use <code>annotGeometryNames</code> to look up names of the <code>sf</code> data frames associated with annotations.
annotGraphName	Name of the <code>listw</code> graph in the SFE object that corresponds to the <code>annotGeometry</code> of interest. Use <code>annotGraphNames</code> to look up names of available annotation graphs.

Details

Most univariate methods in the package `spdep` are supported here. These methods are global, meaning returning one result for all spatial locations in the dataset: `moran`, `geary`, `moran.mc`, `geary.mc`, `moran.test`, `geary.test`, `globalG.test`, `sp.correlogram`.

The following methods are local, meaning each location has its own results: `moran.plot`, `localmoran`, `localmoran_perm`, `localC`, `localC_perm`, `localG`, `localG_perm`, `LOSH`, `LOSH.mc`, `LOSH.cs`. The `GWmodel::gws` method will be supported soon, but is not supported yet.

Global results for genes are stored in `rowData`. For `colGeometry` and `annotGeometry`, the results are added to an attribute of the data frame called `featureData`, which is a `DataFrame` analogous to `rowData` for the gene count matrix. New column names in `featureData` would follow the same rules as in `rowData`. For `colData`, the results can be accessed with the `colFeatureData` function.

Local results are stored in the field `localResults` field of the SFE object, which can be accessed with `localResults` or `localResult`. If the results have p-values, then $-\log_{10} p$ and Benjamin-Hochberg corrected $-\log_{10} p$ are added. Note that in the multiple testing correction, `p.adjustSP` is used.

Value

In `calculateUnivariate`, if `returnDF = TRUE`, then a `DataFrame`, otherwise a list each element of which is the results for each feature. For `run*`, a `SpatialFeatureExperiment` object with the results added. See Details for where the results are stored.

Examples

```
library(SpatialFeatureExperiment)
library(SingleCellExperiment)
library(SFEData)
sfe <- McKellarMuscleData("small")
colGraph(sfe, "visium") <- findVisiumGraph(sfe)
features_use <- rownames(sfe)[1:5]

# Moran's I
```

```

moran_results <- calculateMoransI(sfe,
  features = features_use,
  colGraphName = "visium",
  exprs_values = "counts"
)

# This does not advocate for computing Moran's I on raw counts.
# Just an example for function usage.

sfe <- runMoransI(sfe,
  features = features_use, colGraphName = "visium",
  exprs_values = "counts"
)
# Look at the results
head(rowData(sfe))

# Local Moran's I
sfe <- runUnivariate(sfe,
  type = "localmoran", features = features_use,
  colGraphName = "visium", exprs_values = "counts"
)
head(localResult(sfe, "localmoran", features_use[1]))

# For colData
sfe <- colDataUnivariate(sfe,
  type = "localmoran", features = "nCounts",
  colGraphName = "visium"
)
head(localResult(sfe, "localmoran", "nCounts"))

# For annotGeometries
annotGraph(sfe, "myofiber_tri2nb") <-
  findSpatialNeighbors(sfe,
    type = "myofiber_simplified", MARGIN = 3L,
    method = "tri2nb", dist_type = "idw",
    zero.policy = TRUE
  )
sfe <- annotGeometryUnivariate(sfe,
  type = "localG", features = "area",
  annotGraphName = "myofiber_tri2nb",
  annotGeometryName = "myofiber_simplified",
  zero.policy = TRUE
)
head(localResult(sfe, "localG", "area",
  annotGeometryName = "myofiber_simplified"
))

```


Description

Cluster the correlograms to find patterns in length scales of spatial autocorrelation. All the correlograms clustered must be computed with the same method and have the same number of lags.

Usage

```
clusterCorrelograms(
  sfe,
  features,
  BLUSPARAM,
  sample_id = NULL,
  method = "I",
  colGeometryName = NULL,
  annotGeometryName = NULL,
  show_symbol = TRUE
)
```

Arguments

sfe	A SpatialFeatureExperiment object with correlograms computed for features of interest.
features	Features whose correlograms to cluster.
BLUSPARAM	A BlusterParam object specifying the algorithm to use.
sample_id	Sample(s) in the SFE object whose cells/spots to use. Can be "all" to compute metric for all samples; the metric is computed separately for each sample.
method	"corr" for correlation, "I" for Moran's I, "C" for Geary's C
colGeometryName	Name of colGeometry from which to look for features.
annotGeometryName	Name of annotGeometry from which to look for features.
show_symbol	Logical, whether to show gene symbol instead when Ensembl ID is supplied.

Value

A DataFrame with 3 columns: feature for the features, cluster a factor for cluster membership of the features within each sample, and sample_id for the sample.

Examples

```
library(SpatialFeatureExperiment)
library(SFEData)
library(bluster)
sfe <- McKellarMuscleData("small")
colGraph(sfe, "visium") <- findVisiumGraph(sfe)
inds <- c(1, 3, 4, 5)
sfe <- runUnivariate(sfe,
  type = "sp.correlogram",
  features = rownames(sfe)[inds],
```

```

    exprs_values = "counts", order = 5
  )
  clust <- clusterCorrelograms(sfe,
    features = rownames(sfe)[inds],
    BLUSPARAM = KmeansParam(2)
  )

```

clusterMoranPlot *Find clusters on the Moran plot*

Description

The Moran plot plots the value at each location on the x axis, and the average of the neighbors of each locations on the y axis. Sometimes clusters can be seen on the Moran plot, indicating different types of neighborhoods.

Usage

```

clusterMoranPlot(
  sfe,
  features,
  BLUSPARAM,
  sample_id = NULL,
  colGeometryName = NULL,
  annotGeometryName = NULL,
  show_symbol = TRUE
)

```

Arguments

sfe	A SpatialFeatureExperiment object with Moran plot computed for the feature of interest. If the Moran plot for that feature has not been computed for that feature in this sample_id, it will be calculated and stored in rowData. See calculateUnivariate .
features	Features whose Moran plot are to be cluster. Features whose Moran plots have not been computed will be skipped, with a warning.
BLUSPARAM	A BlusterParam object specifying the algorithm to use.
sample_id	Sample(s) in the SFE object whose cells/spots to use. Can be "all" to compute metric for all samples; the metric is computed separately for each sample.
colGeometryName	Name of colGeometry from which to look for features.
annotGeometryName	Name of annotGeometry from which to look for features.
show_symbol	Logical, whether to show gene symbol instead when Ensembl ID is supplied.

Value

A DataFrame each column of which is a factor for cluster membership of each feature. The column names are the features.

Examples

```
library(SpatialFeatureExperiment)
library(SingleCellExperiment)
library(SFEData)
library(bluster)
sfe <- McKellarMuscleData("small")
colGraph(sfe, "visium") <- findVisiumGraph(sfe)
# Compute moran plot
sfe <- runUnivariate(sfe,
  type = "moran.plot", features = rownames(sfe)[1],
  exprs_values = "counts"
)
clusts <- clusterMoranPlot(sfe, rownames(sfe)[1],
  BLUSPARAM = KmeansParam(2)
)
```

colFeatureData	<i>Get metadata of colData and rowData</i>
----------------	--

Description

Results of spatial analyses on columns in colData and rowData are stored in int_metadata(sfe), or internal metadata. This function allows the users to access these results.

Usage

```
colFeatureData(sfe)
```

```
rowFeatureData(sfe)
```

Arguments

sfe An SFE object.

Value

A DataFrame.

Examples

```
library(SpatialFeatureExperiment)
library(SingleCellExperiment)
library(SFEData)
sfe <- McKellarMuscleData("small")
colGraph(sfe, "visium") <- findVisiumGraph(sfe)
# Moran's I for colData
sfe <- colDataMoransI(sfe, "nCounts")
colFeatureData(sfe)
```

ditto_colors	<i>Colorblind friendly palette from dittoSeq</i>
--------------	--

Description

Just to get the palette without having to install all those dependencies of dittoSeq.

Usage

```
ditto_colors
```

Format

A character vector of hex colors of the palette. There are 40 colors.

Source

The dittoSeq package.

ElbowPlot	<i>Plot the elbow plot or scree plot for PCA</i>
-----------	--

Description

Apparently, there is no apparent way to plot the PC elbow plot other than extracting the variance explained attribute of the dimred slot, because even the OSCA book makes the elbow plot this way, which I find kind of cumbersome compared to Seurat. So I'm writing this function to make the elbow plot with SCE less cumbersome.

Usage

```
ElbowPlot(sce, ndims = 20, reduction = "PCA")
```

Arguments

sce	A SingleCellExperiment object, or anything that inherits from SingleCellExperiment.
ndims	Number of PCs to plot.
reduction	Name of the dimension reduction to use. It must have an attribute called "percentVar". Defaults to "PCA".

Value

A ggplot object. The y axis is percentage of variance explained.

Examples

```
library(SFEData)
library(scater)
sfe <- McKellarMuscleData("small")
sfe <- runPCA(sfe, ncomponents = 10, exprs_values = "counts")
ElbowPlot(sfe, ndims = 10)
```

getDivergeRange	<i>Get beginning and end of palette to center a divergent palette</i>
-----------------	---

Description

This function is no longer used internally as it's unnecessary for scico divergent palettes. But it can be useful when using divergent palettes outside scico where one must specify beginning and end but not midpoint, to override the default palette.

Usage

```
getDivergeRange(values, diverge_center = 0)
```

Arguments

values	Numeric vector to be colored.
diverge_center	Value to center on, defaults to 0.

Value

A numeric vector of length 2, the first element is for beginning, and the second for end. The values are between 0 and 1.

Examples

```
v <- rnorm(10)
getDivergeRange(v, diverge_center = 0)
```

 moranPlot

Use ggplot to plot the moran.plot results

Description

This function uses ggplot2 to plot the Moran plot. The plot would be more aesthetically pleasing than the base R version implemented in spdep. In addition, contours are plotted to show point density on the plot, and the points can be colored by a variable, such as clusters. The contours may also be filled and only influential points plotted. When filled, the viridis E option is used.

Usage

```

moranPlot(
  sfe,
  feature,
  graphName = 1L,
  sample_id = NULL,
  contour_color = "cyan",
  color_by = NULL,
  colGeometryName = NULL,
  annotGeometryName = NULL,
  plot_singletons = TRUE,
  binned = FALSE,
  filled = FALSE,
  divergent = FALSE,
  diverge_center = NULL,
  show_symbol = TRUE,
  bins = 100,
  binwidth = NULL,
  hex = FALSE,
  plot_influential = TRUE,
  ...
)

```

Arguments

sfe	A SpatialFeatureExperiment object.
feature	Name of one variable to show on the plot. It will be converted to sentence case on the x axis and lower case in the y axis appended after "Spatially lagged". One feature at a time since the colors in color_by may be specific to this feature (e.g. from clusterMoranPlot).
graphName	Name of the colGraph or annotGraph, the spatial neighborhood graph used to compute the Moran plot. This is to determine which points are singletons to plot differently on this plot.
sample_id	One sample_id for the sample whose graph to plot.

contour_color	Color of the point density contours, which can be changed so the contours stand out from the points.
color_by	Variable to color the points by. It can be the name of a column in colData, a gene, or the name of a column in the colGeometry specified in colGeometryName. Or it can be a vector of the same length as the number of cells/spots in the sample_id of interest.
colGeometryName	Name of a colGeometry sf data frame whose numeric columns of interest are to be used to compute the metric. Use colGeometryNames to look up names of the sf data frames associated with cells/spots.
annotGeometryName	Name of a annotGeometry of the SFE object, to annotate the gene expression plot.
plot_singletons	Logical, whether to plot items that don't have spatial neighbors.
binned	Logical, whether to plot 2D histograms. This argument has precedence to filled.
filled	Logical, whether to plot filled contours for the non-influential points and only plot influential points as points.
divergent	Logical, whether a divergent palette should be used.
diverge_center	If divergent = TRUE, the center from which the palette should diverge. If NULL, then not centering.
show_symbol	Logical, whether to show human readable gene symbol on the plot instead of Ensembl IDs when the row names are Ensembl IDs. There must be a column in rowData(sfe) called "symbol" for this to work.
bins	Numeric vector giving number of bins in both vertical and horizontal directions. Set to 100 by default.
binwidth	Numeric vector giving bin width in both vertical and horizontal directions. Overrides bins if both set.
hex	Logical, whether to use hexagon rather than rectangular bins. Requires the hexbin package.
plot_influential	Logical, whether to plot influential points with different palette if binned = TRUE.
...	Other arguments to pass to geom_density2d .

Value

A ggplot object.

Examples

```
library(SpatialFeatureExperiment)
library(SingleCellExperiment)
library(SFEData)
library(bluster)
library(scater)
```

```
sfe <- McKellarMuscleData("full")
sfe <- sfe[, colData(sfe)$in_tissue]
sfe <- logNormCounts(sfe)
colGraph(sfe, "visium") <- findVisiumGraph(sfe)
sfe <- runUnivariate(sfe, type = "moran.plot", features = "Myh1")
clust <- clusterMoranPlot(sfe, "Myh1", BLUSPARAM = KmeansParam(2))
moranPlot(sfe, "Myh1", graphName = "visium", color_by = clust[, 1])
```

plotCellBin2D

Plot cell density as 2D histogram

Description

This function plots cell density in histological space as 2D histograms, especially helpful for larger smFISH-based datasets.

Usage

```
plotCellBin2D(sfe, bins = 200, binwidth = NULL, hex = FALSE)
```

Arguments

sfe	A SpatialFeatureExperiment object.
bins	Numeric vector giving number of bins in both vertical and horizontal directions. Set to 100 by default.
binwidth	Numeric vector giving bin width in both vertical and horizontal directions. Overrides bins if both set.
hex	Logical, whether to use hexagon rather than rectangular bins. Requires the hexbin package.

Value

A ggplot object.

Examples

```
library(SFEData)
sfe <- HeNSCLCData()
plotCellBin2D(sfe)
```

plotColDataBin2D *Plot colData and rowData with 2D histograms*

Description

To avoid overplotting in large datasets. The 2D histogram is more informative of point density on the plot than the scatter plot where there are so many points plotted that they effectively form a solid block.

Usage

```
plotColDataBin2D(  
  sfe,  
  x,  
  y,  
  subset = NULL,  
  bins = 100,  
  binwidth = NULL,  
  hex = FALSE,  
  name_true = NULL,  
  name_false = NULL  
)
```

```
plotRowDataBin2D(  
  sfe,  
  x,  
  y,  
  subset = NULL,  
  bins = 100,  
  binwidth = NULL,  
  hex = FALSE,  
  name_true = NULL,  
  name_false = NULL  
)
```

Arguments

sfe	A SpatialFeatureExperiment object.
x	Name of the column in colData or rowData to plot on the x axis of the plot.
y	Name of the column in colData or rowData to plot on the y axis of the plot.
subset	Name of a logical column in colData or rowData, indicating cells or genes to plot with a different palette. Since the 2D histogram is effectively an opaque heatmap, don't use this argument unless the two groups are largely non-overlapping in the variables being plotted.
bins	Numeric vector giving number of bins in both vertical and horizontal directions. Set to 100 by default.

binwidth	Numeric vector giving bin width in both vertical and horizontal directions. Overrides bins if both set.
hex	Logical, whether to use hexagon rather than rectangular bins. Requires the hexbin package.
name_true	Character, name to show on the legend for cells or genes indicated TRUE in the subset argument.
name_false	Character, name to show on the legend for cells or genes indicated FALSE in the subset argument.

Value

A ggplot object

Examples

```
library(SFEData)
sfe <- McKellarMuscleData()
sfe <- sfe[, sfe$in_tissue]
plotColDataBin2D(sfe, "nCounts", "nGenes")
```

plotColDataHistogram *Plot histograms for colData and rowData columns*

Description

Plot histograms for colData and rowData columns

Usage

```
plotColDataHistogram(
  sfe,
  feature,
  fill_by = NULL,
  subset = NULL,
  bins = 100,
  binwidth = NULL,
  scales = "free",
  ncol = 1,
  position = "identity"
)
```

```
plotRowDataHistogram(
  sfe,
  feature,
  fill_by = NULL,
  subset = NULL,
  bins = 100,
```

```

  binwidth = NULL,
  scales = "free",
  ncol = 1,
  position = "identity"
)

```

Arguments

sfe	A SpatialFeatureExperiment object.
feature	Names of columns in colData or rowData to plot. When multiple features are specified, they will be plotted in separate facets.
fill_by	Name of a categorical column in colData or rowData to fill the histogram.
subset	Name of a logical column to only plot a subset of the data.
bins	Numeric vector giving number of bins in both vertical and horizontal directions. Set to 100 by default.
binwidth	Numeric vector giving bin width in both vertical and horizontal directions. Overrides bins if both set.
scales	Should scales be fixed ("fixed", the default), free ("free"), or free in one dimension ("free_x", "free_y")?
ncol	Number of columns in the faceting.
position	Position adjustment, either as a string naming the adjustment (e.g. "jitter" to use position_jitter), or the result of a call to a position adjustment function. Use the latter if you need to change the settings of the adjustment.

Value

A ggplot object

Examples

```

library(SFEData)
sfe <- McKellarMuscleData()
plotColDataHistogram(sfe, c("nCounts", "nGenes"), fill_by = "in_tissue",
  bins = 50)
plotColDataHistogram(sfe, "nCounts", subset = "in_tissue")
sfe2 <- sfe[, sfe$in_tissue]
plotColDataHistogram(sfe2, c("nCounts", "nGenes"), bins = 50)

```

plotColGraph

Plot spatial graphs

Description

A ggplot version of `spdep::plot.nb`, reducing boilerplate for SFE objects.

Usage

```
plotColGraph(
  sfe,
  colGraphName = 1L,
  colGeometryName = NULL,
  sample_id = NULL,
  weights = FALSE,
  segment_size = 0.5,
  geometry_size = 0.5,
  ncol = NULL
)
```

```
plotAnnotGraph(
  sfe,
  annotGraphName = 1L,
  annotGeometryName = 1L,
  sample_id = NULL,
  weights = FALSE,
  segment_size = 0.5,
  geometry_size = 0.5,
  ncol = NULL
)
```

Arguments

<code>sfe</code>	A <code>SpatialFeatureExperiment</code> object.
<code>colGraphName</code>	Name of graph associated with columns of the gene count matrix to be plotted.
<code>colGeometryName</code>	Name of a <code>colGeometry sf</code> data frame whose numeric columns of interest are to be used to compute the metric. Use <code>colGeometryNames</code> to look up names of the <code>sf</code> data frames associated with cells/spots.
<code>sample_id</code>	Sample(s) in the SFE object whose cells/spots to use. Can be "all" to compute metric for all samples; the metric is computed separately for each sample.
<code>weights</code>	Whether to plot weights. If TRUE, then transparency (alpha) of the segments will represent edge weights.
<code>segment_size</code>	Thickness of the segments that represent graph edges.
<code>geometry_size</code>	Point size (for POINT geometries) or line thickness (for LINESTRING and POLYGON) to plot the geometry in the background.
<code>ncol</code>	Number of columns if plotting multiple features. Defaults to NULL, which means using the same logic as <code>facet_wrap</code> , which is used by <code>patchwork</code> 's <code>wrap_plots</code> by default.
<code>annotGraphName</code>	Name of the annotation graph to plot.
<code>annotGeometryName</code>	Name of the <code>annotGeometry</code> , which is associated with the graph specified with <code>annotGraphName</code> , for spatial coordinates of the graph nodes and for context.

Value

A ggplot2 object.

Examples

```
library(SpatialFeatureExperiment)
library(SFEData)
library(sf)
sfe <- McKellarMuscleData("small")
colGraph(sfe, "visium") <- findVisiumGraph(sfe)
plotColGraph(sfe, colGraphName = "visium", colGeometryName = "spotPoly")
# Make the myofiber segmentations a valid POLYGON geometry
ag <- annotGeometry(sfe, "myofiber_simplified")
ag <- st_buffer(ag, 0)
ag <- ag[!st_is_empty(ag), ]
annotGeometry(sfe, "myofiber_simplified") <- ag
annotGraph(sfe, "myofibers") <-
  findSpatialNeighbors(sfe,
    type = "myofiber_simplified", MARGIN = 3,
    method = "tri2nb", dist_type = "idw"
  )
plotAnnotGraph(sfe,
  annotGraphName = "myofibers",
  annotGeometryName = "myofiber_simplified",
  weights = TRUE
)
```

plotCorrelogram

Plot correlogram

Description

Use ggplot2 to plot correlograms computed by `runUnivariate`, pulling results from `rowData`. Correlograms of multiple genes with error bars can be plotted, and they can be colored by any numeric or categorical column in `rowData` or a vector with the same length as `nrow` of the SFE object. The coloring is useful when the correlograms are clustered to show types of length scales or patterns of decay of spatial autocorrelation. For `method = "I"`, the error bars are twice the standard deviation of the estimated Moran's I value.

Usage

```
plotCorrelogram(
  sfe,
  features,
  sample_id = NULL,
  method = "I",
  color_by = NULL,
  facet_by = c("sample_id", "features"),
```

```

ncol = NULL,
colGeometryName = NULL,
annotGeometryName = NULL,
plot_signif = TRUE,
p_adj_method = "BH",
divergent = FALSE,
diverge_center = NULL,
show_symbol = TRUE
)

```

Arguments

sfe	A SpatialFeatureExperiment object.
features	Features to plot, must be in rownames of the gene count matrix, colnames of colData or a colGeometry.
sample_id	Sample(s) in the SFE object whose cells/spots to use. Can be "all" to compute metric for all samples; the metric is computed separately for each sample.
method	"corr" for correlation, "I" for Moran's I, "C" for Geary's C
color_by	Name of a column in rowData(sfe) or in the featureData of colData (see colFeatureData), colGeometry, or annotGeometry by which to color the correlogram of each feature. Alternatively, a vector of the same length as features.
facet_by	Whether to facet by sample_id (default) or features. If faceting by sample_id, then different features will be plotted in the same facet for comparison. If faceting by features, then different samples will be compared for each feature. Ignored if only one sample is specified.
ncol	Number of columns if faceting.
colGeometryName	Name of a colGeometry sf data frame whose numeric columns of interest are to be used to compute the metric. Use colGeometryNames to look up names of the sf data frames associated with cells/spots.
annotGeometryName	Name of a annotGeometry of the SFE object, to annotate the gene expression plot.
plot_signif	Logical, whether to plot significance symbols: $p < 0.001$: ***, $p < 0.01$: **, $p < 0.05$ *, $p < 0.1$: ., otherwise no symbol. The p-values are two sided, based on the assumption that the estimated Moran's I is normally distributed with mean from a randomized version of the data. The mean and variance come from moran.test for Moran's I and geary.test for Geary's C. Take the results with a grain of salt if the data is not normally distributed.
p_adj_method	Multiple testing correction method as in p.adjust , to correct for multiple testing (number of lags times number of features) in the Moran's I estimates if plot_signif = TRUE.
divergent	Logical, whether a divergent palette should be used.
diverge_center	If divergent = TRUE, the center from which the palette should diverge. If NULL, then not centering.

show_symbol Logical, whether to show human readable gene symbol on the plot instead of Ensembl IDs when the row names are Ensembl IDs. There must be a column in rowData(sfe) called "symbol" for this to work.

Value

A ggplot object.

Examples

```
library(SpatialFeatureExperiment)
library(SFEData)
library(bluster)
library(scater)
sfe <- McKellarMuscleData("small")
sfe <- logNormCounts(sfe)
colGraph(sfe, "visium") <- findVisiumGraph(sfe)
inds <- c(1, 3, 4, 5)
features <- rownames(sfe)[inds]
sfe <- runUnivariate(sfe,
  type = "sp.correlogram", features = features,
  exprs_values = "counts", order = 5
)
clust <- clusterCorrelograms(sfe,
  features = features,
  BLUSPARAM = KmeansParam(2)
)
# Color by features
plotCorrelogram(sfe, features)
# Color by something else
plotCorrelogram(sfe, features, color_by = clust$cluster)
# Facet by features
plotCorrelogram(sfe, features, facet_by = "features")
```

plotDimLoadings

Plot top PC loadings of genes

Description

Just like Seurat's VizDimLoadings function. I haven't found an equivalent for SCE but find it useful. But I'm not trying to reproduce that Seurat function exactly. For instance, I don't like it when Seurat imposes a ggplot theme, and I don't like the cowplot theme. Maybe I should rewrite it in base R but for now I'm using Tidyverse.

Usage

```
plotDimLoadings(
  sce,
  dims = 1:4,
```

```

nfeatures = 10,
show_symbol = TRUE,
symbol_col = "symbol",
reduction = "PCA",
balanced = TRUE,
ncol = 2
)

```

Arguments

sce	A SingleCellExperiment object, or anything that inherits from SingleCellExperiment.
dims	Numeric vector specifying which PCs to plot.
nfeatures	Number of genes to plot.
show_symbol	Logical; if the row names of the matrix are Ensembl accessions, indicate whether to show more human readable gene symbols in the plot instead. Ignored if the column specified in symbol_col is absent from rowData.
symbol_col	If the row names of the gene expression matrix are Ensembl accessions to avoid ambiguity in analysis. If not found in rowData, then rownames of the gene count matrix will be used.
reduction	Name of the dimension reduction to use. It must have an attribute called "percentVar". Defaults to "PCA".
balanced	Return an equal number of genes with + and - scores. If FALSE, returns the top genes ranked by the scores absolute values.
ncol	Number of columns in the faceted plot.

Value

A ggplot object. Loadings for different PCs are plotted in different facets so one ggplot object is returned.

Examples

```

library(SFEData)
library(scater)
sfe <- McKellarMuscleData("small")
sfe <- runPCA(sfe, ncomponents = 10, exprs_values = "counts")
plotDimLoadings(sfe, dims = 1:2)

```

plotLocalResult

Plot local results

Description

Plot results of local spatial analyses in space, such as local Getis-Ord G_i^* values.

Usage

```

plotLocalResult(
  sfe,
  type,
  features,
  attribute = NULL,
  sample_id = NULL,
  colGeometryName = NULL,
  annotGeometryName = NULL,
  ncol = NULL,
  ncol_sample = NULL,
  annot_aes = list(),
  annot_fixed = list(),
  aes_use = c("fill", "color", "shape", "linetype"),
  divergent = FALSE,
  diverge_center = NULL,
  annot_divergent = FALSE,
  annot_diverge_center = NULL,
  size = 0,
  shape = 16,
  linetype = 1,
  alpha = 1,
  color = NA,
  fill = "gray80",
  show_symbol = TRUE,
  scattermore = FALSE,
  pointsize = 0,
  ...
)

```

Arguments

sfe	A SpatialFeatureExperiment object.
type	Which local spatial results. Use localResultNames to see which types of results have already been calculated.
features	Character vector of vectors. To see which features have the results of a given type, see localResultFeatures .
attribute	Which field in the local results of the type and features. If the result of each feature is a vector, the this argument is ignored. But if the result is a data frame or a matrix, then this is the column name of the result, such as "Ii" for local Moran's I. For each local spatial analysis method, there's a default attribute. See Details. Use localResultAttrs .
sample_id	Sample(s) in the SFE object whose cells/spots to use. Can be "all" to compute metric for all samples; the metric is computed separately for each sample.
colGeometryName	Name of a colGeometry sf data frame whose numeric columns of interest are to be used to compute the metric. Use colGeometryNames to look up names of

	the sf data frames associated with cells/spots.
annotGeometryName	Name of a annotGeometry of the SFE object, to annotate the gene expression plot.
ncol	Number of columns if plotting multiple features. Defaults to NULL, which means using the same logic as facet_wrap, which is used by patchwork's wrap_plots by default.
ncol_sample	If plotting multiple samples as facets, how many columns of such facets. This is distinct from ncols, which is for multiple features. When plotting multiple features for multiple samples, then the result is a multi-panel plot each panel of which is a plot for each feature faceted by samples.
annot_aes	A named list of plotting parameters for the annotation sf data frame. The names are which geom (as in ggplot2, such as color and fill), and the values are column names in the annotation sf data frame. Tidyeval is NOT supported.
annot_fixed	Similar to annot_aes, but for fixed aesthetic settings, such as color = "gray". The defaults are the same as the relevant defaults for this function.
aes_use	Aesthetic to use for discrete variables. For continuous variables, it's always "fill" for polygons and point shapes 21-25. For discrete variables, it can be fill, color, shape, or linetype, whenever applicable. The specified value will be changed to the applicable equivalent. For example, if the geometry is point but "linetype" is specified, then "shaped" will be used instead.
divergent	Logical, whether a divergent palette should be used.
diverge_center	If divergent = TRUE, the center from which the palette should diverge. If NULL, then not centering.
annot_divergent	Just as divergent, but for the annotGeometry in case it's different.
annot_diverge_center	Just as diverge_center, but for the annotGeometry in case it's different.
size	Fixed size of points or width of lines, including outlines of polygons. For polygons, this defaults to 0, meaning no outlines. For points and lines, this defaults to 0.5. Ignored if size_by is specified.
shape	Fixed shape of points, ignored if shape_by is specified and applicable.
linetype	Fixed line type, ignored if linetype_by is specified and applicable.
alpha	Transparency.
color	Fixed color for colGeometry if color_by is not specified or not applicable, or for annotGeometry if annot_color_by is not specified or not applicable.
fill	Similar to color, but for fill.
show_symbol	Logical, whether to show human readable gene symbol on the plot instead of Ensembl IDs when the row names are Ensembl IDs. There must be a column in rowData(sfe) called "symbol" for this to work.
scattermore	Logical, whether to use the scattermore package to greatly speed up plotting numerous points. Only used for POINT colGeometries. If the geometry is not POINT, then the centroids are used. Recommended for plotting hundreds of

thousands or more cells where the cell polygons can't be seen when plotted due to the large number of cells and small plot size such as when plotting multiple panels for multiple features.

`pointsize` Radius of rasterized point in `scat` termore. Default to 0 for single pixels (fastest).

... Other arguments passed to `wrap_plots`.

Details

Many local spatial analyses return a data frame or matrix as the results, whose columns can be the statistic of interest at each location, its variance, expected value from permutation, p-value, and etc. The `attribute` argument specifies which column to use when there are multiple columns. Below are the defaults for each local method supported by this package what they mean:

`localmoran` **and** `localmoran_perm` I_i , local Moran's I statistic at each location.

`localC_perm` `localC`, the local Geary C statistic at each location.

`localG` **and** `localG_perm` `localG`, the local Getis-Ord G_i or G_i^* statistic. If `include_self = TRUE` when `calculateUnivariate` or `runUnivariate` was called, then it would be G_i^* . Otherwise it's G_i .

`LOSH` **and** `LOSH.mc` H_i , local spatial heteroscedasticity

`moran.plot` `wx`, the average of the value of each neighbor of each location. Moran plot is best plotted as a scatter plot of `wx` vs `x`. See `moranPlot`.

Other local methods not listed above return vectors as results. For instance, `localC` returns a vector by default, which is the local Geary's C statistic.

Value

A `ggplot2` object if plotting one feature. A patchwork object if plotting multiple features.

Note

While this function shares internals with `plotSpatialFeature`, there are some important differences. In `plotSpatialFeature`, the `annotGeometry` is indeed only used for annotation and the protagonist is the `colGeometry`, since it's easy to directly use `ggplot2` to plot the data in `annotGeometry sf` data frames while overlaying `annotGeometry` and `colGeometry` involves more complicated code. In contrast, in this function, local results for `annotGeometry` can be plotted separately without anything related to `colGeometry`. Note that when `annotGeometry` local results are plotted without `colGeometry`, the `annot_*` arguments are ignored. Use the other arguments for aesthetics as if it's for `colGeometry`.

Examples

```
library(SpatialFeatureExperiment)
library(SFEData)
library(scater)
sfe <- McKellarMuscleData("small")
colGraph(sfe, "visium") <- findVisiumGraph(sfe)
feature_use <- rownames(sfe)[1]
```

```

sfe <- logNormCounts(sfe)
sfe <- runUnivariate(sfe, "localmoran", feature_use)
# Which types of results are available?
localResultNames(sfe)
# Which features for localmoran?
localResultFeatures(sfe, "localmoran")
# Which columns does the localmoran results have?
localResultAttrs(sfe, "localmoran", feature_use)
plotLocalResult(sfe, "localmoran", feature_use, "Ii",
  colGeometryName = "spotPoly"
)

# For annotGeometry
# Make sure it's type POLYGON
annotGeometry(sfe, "myofiber_simplified") <-
  sf::st_buffer(annotGeometry(sfe, "myofiber_simplified"), 0)
annotGraph(sfe, "poly2nb_my") <-
  findSpatialNeighbors(sfe,
    type = "myofiber_simplified", MARGIN = 3,
    method = "poly2nb", zero.policy = TRUE
  )
sfe <- annotGeometryUnivariate(sfe, "localmoran",
  features = "area",
  annotGraphName = "poly2nb_my",
  annotGeometryName = "myofiber_simplified",
  zero.policy = TRUE
)
plotLocalResult(sfe, "localmoran", "area", "Ii",
  annotGeometryName = "myofiber_simplified",
  size = 0.3, color = "cyan"
)
plotLocalResult(sfe, "localmoran", "area", "Z.Ii",
  annotGeometryName = "myofiber_simplified"
)
# don't use annot_* arguments when annotGeometry is plotted without colGeometry

```

plotMoranMC

Plot Moran/Geary monte carlo results

Description

Plot the simulations as a density plot or histogram compared to the observed Moran's I or Geary's C, with ggplot2 so it looks nicer. Unlike the plotting function in spdep, this function can also plot the same feature in different samples as facets or plot different features or samples together for comparison.

Usage

```

plotMoranMC(
  sfe,

```

```

    features,
    sample_id = NULL,
    facet_by = c("sample_id", "features"),
    ncol = NULL,
    colGeometryName = NULL,
    annotGeometryName = NULL,
    ptype = c("density", "histogram", "freqpoly"),
    show_symbol = TRUE,
    ...
  )

```

Arguments

sfe	A SpatialFeatureExperiment object.
features	Features to plot, must be in rownames of the gene count matrix, colnames of colData or a colGeometry.
sample_id	Sample(s) in the SFE object whose cells/spots to use. Can be "all" to compute metric for all samples; the metric is computed separately for each sample.
facet_by	Whether to facet by sample_id (default) or features. If faceting by sample_id, then different features will be plotted in the same facet for comparison. If faceting by features, then different samples will be compared for each feature. Ignored if only one sample is specified.
ncol	Number of columns if faceting.
colGeometryName	Name of a colGeometry sf data frame whose numeric columns of interest are to be used to compute the metric. Use colGeometryNames to look up names of the sf data frames associated with cells/spots.
annotGeometryName	Name of a annotGeometry of the SFE object, to annotate the gene expression plot.
ptype	Plot type, one of "density", "histogram", or "freqpoly".
show_symbol	Logical, whether to show human readable gene symbol on the plot instead of Ensembl IDs when the row names are Ensembl IDs. There must be a column in rowData(sfe) called "symbol" for this to work.
...	Other arguments passed to geom_density , geom_histogram , or geom_freqpoly , depending on ptype.

Value

A ggplot2 object.

Examples

```

library(SpatialFeatureExperiment)
library(SFEData)
sfe <- McKellarMuscleData("small")
colGraph(sfe, "visium") <- findVisiumGraph(sfe)

```

```
sfe <- colDataUnivariate(sfe, type = "moran.mc", "nCounts", nsim = 100)
plotMoranMC(sfe, "nCounts")
```

plotSpatialFeature *Plot gene expression in space*

Description

Unlike Seurat and ggspavis, plotting functions in this package uses geom_sf whenever applicable.

Usage

```
plotSpatialFeature(
  sfe,
  features,
  colGeometryName = 1L,
  sample_id = NULL,
  ncol = NULL,
  ncol_sample = NULL,
  annotGeometryName = NULL,
  annot_aes = list(),
  annot_fixed = list(),
  exprs_values = "logcounts",
  aes_use = c("fill", "color", "shape", "linetype"),
  divergent = FALSE,
  diverge_center = NA,
  annot_divergent = FALSE,
  annot_diverge_center = NA,
  size = 0,
  shape = 16,
  linetype = 1,
  alpha = 1,
  color = NA,
  fill = "gray80",
  show_symbol = TRUE,
  scattermore = FALSE,
  pointsize = 0,
  ...
)
```

Arguments

sfe	A SpatialFeatureExperiment object.
features	Features to plot, must be in rownames of the gene count matrix, colnames of colData or a colGeometry.

colGeometryName	Name of a colGeometry sf data frame whose numeric columns of interest are to be used to compute the metric. Use colGeometryNames to look up names of the sf data frames associated with cells/spots.
sample_id	Sample(s) in the SFE object whose cells/spots to use. Can be "all" to compute metric for all samples; the metric is computed separately for each sample.
ncol	Number of columns if plotting multiple features. Defaults to NULL, which means using the same logic as <code>facet_wrap</code> , which is used by patchwork's wrap_plots by default.
ncol_sample	If plotting multiple samples as facets, how many columns of such facets. This is distinct from <code>ncols</code> , which is for multiple features. When plotting multiple features for multiple samples, then the result is a multi-panel plot each panel of which is a plot for each feature faceted by samples.
annotGeometryName	Name of a annotGeometry of the SFE object, to annotate the gene expression plot.
annot_aes	A named list of plotting parameters for the annotation sf data frame. The names are which geom (as in <code>ggplot2</code> , such as <code>color</code> and <code>fill</code>), and the values are column names in the annotation sf data frame. <code>Tidyeval</code> is NOT supported.
annot_fixed	Similar to <code>annot_aes</code> , but for fixed aesthetic settings, such as <code>color = "gray"</code> . The defaults are the same as the relevant defaults for this function.
exprs_values	Integer scalar or string indicating which assay of <code>x</code> contains the expression values.
aes_use	Aesthetic to use for discrete variables. For continuous variables, it's always "fill" for polygons and point shapes 21-25. For discrete variables, it can be <code>fill</code> , <code>color</code> , <code>shape</code> , or <code>linetype</code> , whenever applicable. The specified value will be changed to the applicable equivalent. For example, if the geometry is point but "linetype" is specified, then "shaped" will be used instead.
divergent	Logical, whether a divergent palette should be used.
diverge_center	If <code>divergent = TRUE</code> , the center from which the palette should diverge. If NULL, then not centering.
annot_divergent	Just as <code>divergent</code> , but for the <code>annotGeometry</code> in case it's different.
annot_diverge_center	Just as <code>diverge_center</code> , but for the <code>annotGeometry</code> in case it's different.
size	Fixed size of points or width of lines, including outlines of polygons. For polygons, this defaults to 0, meaning no outlines. For points and lines, this defaults to 0.5. Ignored if <code>size_by</code> is specified.
shape	Fixed shape of points, ignored if <code>shape_by</code> is specified and applicable.
linetype	Fixed line type, ignored if <code>linetype_by</code> is specified and applicable.
alpha	Transparency.
color	Fixed color for <code>colGeometry</code> if <code>color_by</code> is not specified or not applicable, or for <code>annotGeometry</code> if <code>annot_color_by</code> is not specified or not applicable.
fill	Similar to <code>color</code> , but for fill.

show_symbol	Logical, whether to show human readable gene symbol on the plot instead of Ensembl IDs when the row names are Ensembl IDs. There must be a column in rowData(sfe) called "symbol" for this to work.
scattermore	Logical, whether to use the scattermore package to greatly speed up plotting numerous points. Only used for POINT colGeometries. If the geometry is not POINT, then the centroids are used. Recommended for plotting hundreds of thousands or more cells where the cell polygons can't be seen when plotted due to the large number of cells and small plot size such as when plotting multiple panels for multiple features.
pointsize	Radius of rasterized point in scattermore. Default to 0 for single pixels (fastest).
...	Other arguments passed to <code>wrap_plots</code> .

Details

In the documentation of this function, a "feature" can be a gene (or whatever entity that corresponds to rows of the gene count matrix), a column in colData, or a column in the colGeometry sf data frame specified in the colGeometryName argument.

For continuous variables, the Blues palette from colorbrewer is used if divergent = FALSE, and the roma palette from the scico package if divergent = TRUE. For discrete variables, the dittoSeq palette is used. The defaults are colorblind friendly. For annotation, the PuRd colorbrewer palette is used for continuous variables and the other end of the dittoSeq palette is used for discrete variables.

Value

A ggplot2 object if plotting one feature. A patchwork object if plotting multiple features.

Examples

```
library(SFEData)
library(sf)
sfe <- McKellarMuscleData("small")
# features can be genes or colData or colGeometry columns
plotSpatialFeature(sfe, c("nCounts", rownames(sfe)[1]),
  exprs_values = "counts",
  colGeometryName = "spotPoly",
  annotGeometryName = "tissueBoundary"
)
# Change fixed aesthetics
plotSpatialFeature(sfe, "nCounts",
  colGeometryName = "spotPoly",
  annotGeometryName = "tissueBoundary",
  annot_fixed = list(color = "blue", size = 0.3, fill = NA),
  alpha = 0.7
)
# Make the myofiber segmentations a valid POLYGON geometry
ag <- annotGeometry(sfe, "myofiber_simplified")
ag <- st_buffer(ag, 0)
ag <- ag[!st_is_empty(ag), ]
annotGeometry(sfe, "myofiber_simplified") <- ag
```



```
# Also plot an annotGeometry variable
plotSpatialFeature(sfe, "nCounts",
  colGeometryName = "spotPoly",
  annotGeometryName = "myofiber_simplified",
  annot_aes = list(fill = "area")
)
```

spatialReducedDim *Plot dimension reduction components in space*

Description

Such as plotting the value of projection of gene expression of each cell to a principal component in space. At present, this function does not work for the 3D array of geographically weighted PCA (GWPCA), but a future version will deal with GWPCA results.

Usage

```
spatialReducedDim(
  sfe,
  dimred,
  ncomponents,
  colGeometryName = 1L,
  sample_id = NULL,
  ncol = NULL,
  ncol_sample = NULL,
  annotGeometryName = NULL,
  annot_aes = list(),
  annot_fixed = list(),
  exprs_values = "logcounts",
  aes_use = c("fill", "color", "shape", "linetype"),
  divergent = FALSE,
  diverge_center = NULL,
  annot_divergent = FALSE,
  annot_diverge_center = NULL,
  size = 0,
  shape = 16,
  linetype = 1,
  alpha = 1,
  color = NA,
  fill = "gray80",
  scattermore = FALSE,
  pointsize = 0,
  ...
)
```

Arguments

sfe	A SpatialFeatureExperiment object.
dimred	A string or integer scalar indicating the reduced dimension result in reducedDims(sfe) to plot.
ncomponents	A numeric scalar indicating the number of dimensions to plot, starting from the first dimension. Alternatively, a numeric vector specifying the dimensions to be plotted.
colGeometryName	Name of a colGeometry sf data frame whose numeric columns of interest are to be used to compute the metric. Use colGeometryNames to look up names of the sf data frames associated with cells/spots.
sample_id	Sample(s) in the SFE object whose cells/spots to use. Can be "all" to compute metric for all samples; the metric is computed separately for each sample.
ncol	Number of columns if plotting multiple features. Defaults to NULL, which means using the same logic as facet_wrap, which is used by patchwork's wrap_plots by default.
ncol_sample	If plotting multiple samples as facets, how many columns of such facets. This is distinct from ncols, which is for multiple features. When plotting multiple features for multiple samples, then the result is a multi-panel plot each panel of which is a plot for each feature faceted by samples.
annotGeometryName	Name of a annotGeometry of the SFE object, to annotate the gene expression plot.
annot_aes	A named list of plotting parameters for the annotation sf data frame. The names are which geom (as in ggplot2, such as color and fill), and the values are column names in the annotation sf data frame. Tidyeval is NOT supported.
annot_fixed	Similar to annot_aes, but for fixed aesthetic settings, such as color = "gray". The defaults are the same as the relevant defaults for this function.
exprs_values	Integer scalar or string indicating which assay of x contains the expression values.
aes_use	Aesthetic to use for discrete variables. For continuous variables, it's always "fill" for polygons and point shapes 21-25. For discrete variables, it can be fill, color, shape, or linetype, whenever applicable. The specified value will be changed to the applicable equivalent. For example, if the geometry is point but "linetype" is specified, then "shaped" will be used instead.
divergent	Logical, whether a divergent palette should be used.
diverge_center	If divergent = TRUE, the center from which the palette should diverge. If NULL, then not centering.
annot_divergent	Just as divergent, but for the annotGeometry in case it's different.
annot_diverge_center	Just as diverge_center, but for the annotGeometry in case it's different.
size	Fixed size of points or width of lines, including outlines of polygons. For polygons, this defaults to 0, meaning no outlines. For points and lines, this defaults to 0.5. Ignored if size_by is specified.

shape	Fixed shape of points, ignored if shape_by is specified and applicable.
linetype	Fixed line type, ignored if linetype_by is specified and applicable.
alpha	Transparency.
color	Fixed color for colGeometry if color_by is not specified or not applicable, or for annotGeometry if annot_color_by is not specified or not applicable.
fill	Similar to color, but for fill.
scattermore	Logical, whether to use the scattermore package to greatly speed up plotting numerous points. Only used for POINT colGeometries. If the geometry is not POINT, then the centroids are used. Recommended for plotting hundreds of thousands or more cells where the cell polygons can't be seen when plotted due to the large number of cells and small plot size such as when plotting multiple panels for multiple features.
pointsize	Radius of rasterized point in scattermore. Default to 0 for single pixels (fastest).
...	Other arguments passed to wrap_plots .

Value

Same as in [plotSpatialFeature](#). A ggplot2 object if plotting one component. A patchwork object if plotting multiple components.

See Also

[scater::plotReducedDim](#)

Examples

```
library(SFEData)
library(scater)
sfe <- McKellarMuscleData("small")
sfe <- logNormCounts(sfe)
sfe <- runPCA(sfe, ncomponents = 2)
spatialReducedDim(sfe, "PCA", 2, "spotPoly",
  annotGeometryName = "tissueBoundary",
  divergent = TRUE, diverge_center = 0
)
# Basically PC1 separates spots not on tissue from those on tissue.
```

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