

# Package ‘RiboCrypt’

January 25, 2022

**Type** Package

**Title** Interactive visualization in genomics

**Version** 1.0.0

**License** MIT + file LICENSE

**Description** R Package for interactive visualization and browsing NGS data.

It contains a browser for both transcript and genomic coordinate view.

In addition a QC and general metaplots are included, among others differential translation plots and gene expression plots. The package is still under development.

**biocViews** Software, Sequencing, RiboSeq, RNASeq,

**Encoding** UTF-8

**LazyData** true

**BugReports** <https://github.com/m-swirski/RiboCrypt/issues>

**URL** <https://github.com/m-swirski/RiboCrypt>

**Depends** R (>= 3.6.0), ORFik (>= 1.13.12)

**Imports** BiocGenerics, BiocParallel, Biostrings, data.table, dplyr,  
GenomeInfoDb, GenomicFeatures, GenomicRanges, ggplot2, IRanges,  
plotly, rlang

**Suggests** testthat, rmarkdown, knitr, BiocStyle, BSgenome,  
BSgenome.Hsapiens.UCSC.hg19

**RoxygenNote** 7.1.2

**VignetteBuilder** knitr

**git\_url** <https://git.bioconductor.org/packages/RiboCrypt>

**git\_branch** RELEASE\_3\_14

**git\_last\_commit** c07b93f

**git\_last\_commit\_date** 2021-10-26

**Date/Publication** 2022-01-25

**Author** Michal Swirski [aut, cre],  
Haakon Tjeldnes [ctb]

**Maintainer** Michal Swirski <michal.swirski@uw.edu.pl>

## R topics documented:

multiOmicsPlot_animate . . . . .	2
multiOmicsPlot_list . . . . .	4
multiOmicsPlot_ORFikExp . . . . .	6
RiboCrypt.template.experiment . . . . .	8

<b>Index</b>	<b>9</b>
--------------	----------

---

multiOmicsPlot\_animate  
*Multi-omics animation using list input*

---

### Description

The animation will move with a play button, there is 1 transition per library given.

### Usage

```
multiOmicsPlot_animate(
  target_range,
  annotation = target_range,
  reference_sequence,
  reads,
  withFrames = NULL,
  colors = NULL,
  kmers = NULL,
  kmers_type = c("mean", "sum")[1],
  ylabels = NULL,
  proportions = NULL,
  width = NULL,
  height = NULL,
  plot_name = "default",
  plot_title = NULL,
  display_sequence = FALSE,
  annotation_names = NULL,
  start_codons = "ATG",
  stop_codons = c("TAA", "TAG", "TGA"),
  custom_motif = NULL
)
```

### Arguments

`target_range` the whole region to visualize, a [GRangesList](#) or [GRanges](#) object

`annotation` the whole annotation which your target region is a subset, a [GRangesList](#) or [GRanges](#) object

`reference_sequence` the genome reference, a [FaFile](#) or [FaFile](#) convertible object

reads	the NGS libraries, as a list of <a href="#">GRanges</a> with or without score column for replicates.
withFrames	a logical vector, default NULL. Alternative: a length 1 or same length as list length of "reads" argument.
colors	character, default NULL (automatic colouring). If "withFrames" argument is TRUE, colors are set to c("red", "green", "blue") for the 3 frames. Alternative: Character vector of length 1 or length of "reads" list argument.
kmers	numeric (integer), bin positions into kmers.
kmers_type	character, function used for kmers sliding window. default: "mean", alternative: "sum"
ylabels	character, default NULL. Name of libraries in "reads" list argument.
proportions	numeric, default NULL. Width of plot.
width	numeric, default NULL. Width of plot.
height	numeric, default NULL. Height of plot.
plot_name	= character, default "default" (will create name from target_range name). Alternative: custom name for region.
plot_title	character, default NULL. A title for plot.
display_sequence	logical, default FALSE. If TRUE, display nucleotide sequence in plot.
annotation_names	character, default NULL. Alternative naming for annotation.
start_codons	character vector, default "ATG"
stop_codons	character vector, default c("TAA", "TAG", "TGA")
custom_motif	character vector, default NULL.

**Value**

the plot object

**Examples**

```
library(ORFik)
df <- ORFik.template.experiment()[3,] #Use third library in experiment only
if (requireNamespace("BSgenome.Hsapiens.UCSC.hg19")) {
  cds <- loadRegion(df, "cds")
  multiOmicsPlot_ORFikExp(extendLeaders(extendTrailers(cds[1], 30), 30), df = df,
    reference_sequence = BSgenome.Hsapiens.UCSC.hg19::Hsapiens,
    frames_type = "columns")
}
```

---

multiOmicsPlot\_list    *Multi-omics plot using list input*

---

## Description

Customizable html plots for visualizing genomic data.

## Usage

```
multiOmicsPlot_list(
  target_range,
  annotation = target_range,
  reference_sequence,
  reads,
  withFrames = NULL,
  frames_type = "lines",
  colors = NULL,
  kmers = NULL,
  kmers_type = c("mean", "sum")[1],
  ylabels = NULL,
  proportions = NULL,
  width = NULL,
  height = NULL,
  plot_name = "default",
  plot_title = NULL,
  display_sequence = FALSE,
  annotation_names = NULL,
  start_codons = "ATG",
  stop_codons = c("TAA", "TAG", "TGA"),
  custom_motif = NULL,
  BPPARAM = bpparam()
)
```

## Arguments

target_range	the whole region to visualize, a <a href="#">GRangesList</a> or <a href="#">GRanges</a> object
annotation	the whole annotation which your target region is a subset, a <a href="#">GRangesList</a> or <a href="#">GRanges</a> object
reference_sequence	the genome reference, a <a href="#">FaFile</a> or <a href="#">FaFile</a> convertible object
reads	the NGS libraries, as a list of <a href="#">GRanges</a> with or without score column for replicates.
withFrames	a logical vector, default NULL. Alternative: a length 1 or same length as list length of "reads" argument.

frames_type	character, default "lines". Alternative: - columns - stacks - area
colors	character, default NULL (automatic colouring). If "withFrames" argument is TRUE, colors are set to c("red", "green", "blue") for the 3 frames. Alternative: Character vector of length 1 or length of "reads" list argument.
kmers	numeric (integer), bin positions into kmers.
kmers_type	character, function used for kmers sliding window. default: "mean", alternative: "sum"
ylabels	character, default NULL. Name of libraries in "reads" list argument.
proportions	numeric, default NULL. Width of plot.
width	numeric, default NULL. Width of plot.
height	numeric, default NULL. Height of plot.
plot_name	= character, default "default" (will create name from target_range name). Alternative: custom name for region.
plot_title	character, default NULL. A title for plot.
display_sequence	logical, default FALSE. If TRUE, display nucleotide sequence in plot.
annotation_names	character, default NULL. Alternative naming for annotation.
start_codons	character vector, default "ATG"
stop_codons	character vector, default c("TAA", "TAG", "TGA")
custom_motif	character vector, default NULL.
BPPARAM	how many cores/threads to use? default: BiocParallel::bpparam(). To see number of threads used, do BiocParallel::bpparam()\$workers. You can also add a time remaining bar, for a more detailed pipeline.

**Value**

the plot object

**Examples**

```
library(ORFik)
df <- ORFik.template.experiment()[3,] #Use third library in experiment only
if (requireNamespace("BSgenome.Hsapiens.UCSC.hg19")) {
  cds <- loadRegion(df, "cds")
  multiOmicsPlot_ORFikExp(extendLeaders(extendTrailers(cds[1], 30), 30), df = df,
    reference_sequence = BSgenome.Hsapiens.UCSC.hg19::Hsapiens,
    frames_type = "columns")
}
```

---

 multiOmicsPlot\_ORFikExp

*Multi-omics plot using ORFik experiment input*


---

## Description

Customizable html plots for visualizing genomic data.

## Usage

```
multiOmicsPlot_ORFikExp(
  target_range,
  annotation = target_range,
  df,
  reference_sequence = findFa(df),
  reads = outputLibs(df, type = "pshifted", output.mode = "envirlist", naming = "full"),
  withFrames = libraryTypes(df, uniqueTypes = FALSE) %in% c("RFP", "RPF", "LSU"),
  frames_type = "lines",
  colors = NULL,
  kmers = NULL,
  kmers_type = c("mean", "sum")[1],
  ylabels = bamVarName(df),
  proportions = NULL,
  width = NULL,
  height = NULL,
  plot_name = "default",
  plot_title = NULL,
  display_sequence = FALSE,
  annotation_names = NULL,
  start_codons = "ATG",
  stop_codons = c("TAA", "TAG", "TGA"),
  custom_motif = NULL,
  BPPARAM = bpparam()
)
```

## Arguments

target_range	the whole region to visualize, a <a href="#">GRangesList</a> or <a href="#">GRanges</a> object
annotation	the whole annotation which your target region is a subset, a <a href="#">GRangesList</a> or <a href="#">GRanges</a> object
df	an ORFik <a href="#">experiment</a> or a list containing ORFik experiments. Usually a list when you have split Ribo-seq and RNA-seq etc.
reference_sequence	the genome reference, default <code>ORFik::findFa(df)</code>

reads	the NGS libraries, as a list of <a href="#">GRanges</a> with or without score column for replicates. Default: <code>outputLibs(df, type = "pshifted", output.mode = "envirlist", naming = "full")</code>
withFrames	a logical vector, default <code>libraryTypes(df, uniqueTypes = FALSE) %in% c("RFP", "RPF", "LSU")</code> Alternative: a length 1 or same length as list length of "reads" argument.
frames_type	character, default "lines". Alternative: - columns - stacks - area
colors	character, default NULL (automatic colouring). If "withFrames" argument is TRUE, colors are set to <code>c("red", "green", "blue")</code> for the 3 frames. Alternative: Character vector of length 1 or length of "reads" list argument.
kmers	numeric (integer), bin positions into kmers.
kmers_type	character, function used for kmers sliding window. default: "mean", alternative: "sum"
ylabels	character, default <code>bamVarName(df)</code> . Name of libraries in "reads" list argument.
proportions	numeric, default NULL. Width of plot.
width	numeric, default NULL. Width of plot.
height	numeric, default NULL. Height of plot.
plot_name	character, default "default" (will create name from <code>target_range</code> name). Alternative: custom name for region.
plot_title	character, default NULL. A title for plot.
display_sequence	logical, default FALSE. If TRUE, display nucleotide sequence in plot.
annotation_names	character, default NULL. Alternative naming for annotation.
start_codons	character vector, default "ATG"
stop_codons	character vector, default <code>c("TAA", "TAG", "TGA")</code>
custom_motif	character vector, default NULL.
BPPARAM	how many cores/threads to use? default: <code>BiocParallel::bpparam()</code> . To see number of threads used, do <code>BiocParallel::bpparam()\$workers</code> . You can also add a time remaining bar, for a more detailed pipeline.

**Value**

the plot object

**Examples**

```
library(ORFik)
df <- ORFik.template.experiment()[3,] #Use third library in experiment only
if (requireNamespace("BSgenome.Hsapiens.UCSC.hg19")) {
  cds <- loadRegion(df, "cds")
  multiOmicsPlot_ORFikExp(extendLeaders(extendTrailers(cds[1], 30), 30), df = df,
```

```
reference_sequence = BSgenome.Hsapiens.UCSC.hg19::Hsapiens,  
frames_type = "columns")  
}
```

---

RiboCrypt.template.experiment

*An ORFik experiment to see how it looks*

---

## Description

Toy-data created to resemble human genes:

Number of genes: 6

Ribo-seq: 2 libraries RNA-seq: 2 libraries CAGE: 1 library PAS (poly-A): 1 library

## Usage

```
RiboCrypt.template.experiment(as.temp = FALSE)
```

## Arguments

as.temp	logical, default FALSE, load as ORFik experiment. If TRUE, loads as data.frame template of the experiment.
---------	--

## Value

an ORFik experiment

## Examples

```
ORFik.template.experiment()
```



# Index

## \* **ORFik\_experiment**

RiboCrypt.template.experiment, 8

experiment, 6

FaFile, 2, 4

GRanges, 2–4, 6, 7

GRangesList, 2, 4, 6

multiOmicsPlot\_animate, 2

multiOmicsPlot\_list, 4

multiOmicsPlot\_ORFikExp, 6

RiboCrypt.template.experiment, 8