Package ‘RLSeq’

February 23, 2024

Type    Package
Title   RLSeq: An analysis package for R-loop mapping data
Version 1.8.0

Description RLSeq is a toolkit for analyzing and evaluating R-loop mapping datasets. RLSeq serves two primary purposes: (1) to facilitate the evaluation of dataset quality, and (2) to enable R-loop analysis in the context of publicly-available data sets from RLBase. The package is intended to provide a simple pipeline, called with the `RLSeq()` function, which performs all main analyses. Individual functions are also accessible and provide custom analysis capabilities. Finally an HTML report is generated with `report()`.

License    MIT + file LICENSE
Encoding   UTF-8
Roxygen    list(markdown = TRUE)
RoxygenNote 7.2.1

Suggests AnnotationDbi, BiocStyle, covr, lintr, rcmdcheck, DT, htr, jsonlite, kableExtra, kernlab, knitr, magick, MASS, org.Hs.eg.db, R.utils, randomForest, readr, rmarkdown, rpart, testthat (>= 3.0.0), tibble, tidyr, TxDb.Hsapiens.UCSC.hg19.knownGene, futile.logger

Config/testthat/edition 3
Depends  R (>= 4.2.0)
Imports dplyr, ggplot2, RColorBrewer, grid, regioneR, valr, caretEnsemble, GenomicFeatures, rtracklayer, GenomicRanges, GenomeInfoDb, ComplexHeatmap, AnnotationHub, VennDiagram, callr, circlize, ggplotify, ggprism, methods, stats, RLHub, aws.s3, pheatmap

VignetteBuilder knitr

biocViews Sequencing, Coverage, Epigenetics, Transcriptomics, Classification

URL https://github.com/Bishop-Laboratory/RLSeq,
https://bishop-laboratory.github.io/RLSeq/

BugReports https://github.com/Bishop-Laboratory/RLSeq/issues
BiocType Software

git_url https://git.bioconductor.org/packages/RLSeq

git_branch RELEASE_3_18

git_last_commit 299fafc

git_last_commit_date 2023-10-24

Repository Bioconductor 3.18

Date/Publication 2024-02-22

Author Henry Miller [aut, cre, cph] (<https://orcid.org/0000-0003-3756-3918>),
Daniel Montemayor [ctb] (<https://orcid.org/0000-0001-8702-5646>),
Simon Levy [ctb] (<https://orcid.org/0000-0002-4623-5716>),
Anna Vines [ctb] (<https://orcid.org/0000-0002-5149-7737>),
Alexander Bishop [ths, cph] (<https://orcid.org/0000-0002-5742-4387>)

Maintainer Henry Miller <millerh1@uthscsa.edu>

**R topics documented:**

- analyzeRLFS .......................................................... 3
- auxdata .............................................................. 4
- available_genomes ............................................... 6
- checkRLFSAnno ...................................................... 7
- corrAnalyze .......................................................... 8
- corrHeatmap .......................................................... 9
- featureEnrich ....................................................... 10
- feature_ggplot ...................................................... 11
- geneAnnotation ...................................................... 12
- genomeMasks ........................................................ 13
- getChromSizes ....................................................... 14
- getGSSignal .......................................................... 14
- getRLFSAnno ........................................................ 15
- noiseAnalyze ........................................................ 15
- noiseComparisonPlot .............................................. 16
- peak_stats ........................................................... 17
- plotEnrichment ....................................................... 18
- plotFingerprint ...................................................... 19
- plotRLFSRes .......................................................... 20
- plotRLRegionOverlap .............................................. 21
- plotTxFeatureOverlap ............................................ 21
- predictCondition ................................................... 22
- randomWindows ...................................................... 24
- report ................................................................. 25
- rlbaseNoiseAnalyze ................................................ 26
- RLRanges-class ...................................................... 27
- RLRangesFromRLBase .............................................. 28
- rlRegionTest ........................................................ 29
- rlresult .............................................................. 30
analyzeRLFS

Description

Analyzes the enrichment of ranges within R-loop forming sequences (RLFS). See details.

Usage

analyzeRLFS(
  object,
  mask = NULL,
  quiet = FALSE,
  useMask = TRUE,
  noZ = FALSE,
  ntimes = 100,
  stepsize = 50,
  ...
)

Arguments

- **object**: An `RLRanges` object.
- **mask**: GRanges object containing masked genomic ranges. Not needed unless masked genome unavailable (see `genomeMasks`). Custom masks can be generated using `regioneR::getMask`.
- **quiet**: If TRUE, messages are suppressed. Default: FALSE.
- **useMask**: If FALSE, masked genome is not used. This is not recommended unless a mask is unavailable as it can lead to spurious results. Default: TRUE.
- **noZ**: If TRUE, Z-score distribution is not calculated. Default: FALSE.
- **ntimes**: Number of permutations to perform (default: 100).
- **stepsize**: The step size for calculating the Z score distribution. Default: 50. See also `regioneR::localZScore`.
- **...**: Arguments passed to `regioneR::permTest`. 
Details

R-loop forming sequences are regions of the genome with sequences that are favorable for R-loop formation. They are computationally predicted with the QmRLFS-finder software program and serve as a data-independent test of whether a sample has mapped R-loops robustly or not.

Method:
Permutation testing is implemented via regioneR::permTest such that, for each permutation, R-loop peaks were randomized using regioneR::circularRandomizeRegions and then the number of overlaps with RLFS are counted. 100 permutations are used by default to build an empirical distribution for peak/RLFS overlap. Then the true number of overlaps from non-randomized peaks and RLFS are compared to the null distribution to calculate Z-score and significance of enrichment. Finally, a Z-score distribution was calculated (using regioneR::localZScore) 5kb upstream and downstream of the average RLFS midpoint.

These results are subsequently used in the binary classification of the sample as "POS" (maps R-loops) or "NEG" (does not map R-loops). See also predictCondition.

Value

An RLRanges object with RLFS analysis results accessible via RLSeq::rlresult(object, "rlfsRes"). Contains the following structure:

- perTestResults
  - An object of the class permTestResultsList from regioneR with the results of permutation testing. See also regioneR::permTest for full description.

- Z-scores
  - An object of the class localZScoreResultsList from regioneR. Contains the results of local Z-score analysis +/-5kb around each RLFS. See also regioneR::localZScore.

Examples

```r
# Example dataset
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# Perform RLFS analysis (remove ntimes=2 and noZ=TRUE for a typical analysis)
rlr <- analyzeRLFS(rlr, ntimes = 2, noZ = TRUE)
```

Auxiliary Data

A list containing data used by RLSeq functions. It can also be useful for checking the available modes and genomes in RLSeq. See also the data-raw/auxdata.R script that was used to create it.

Usage

auxdata
Format

An object of class list of length 12.

Details

Structure:
A named list containing the following entries:

- db_cols
  A tbl with colors associated with each database in RLHub useful for plotting. See also RLHub::annotations.

- annotypes
  A tbl containing the annotation databases and annotation types available from RLBase. See also RLHub::annotations.

- ip_cols
  A tbl containing the colors associated with each "Immunoprecipitation type" (ip_type) in RLBase. See also RLHub::rlbase_samples.

- mode_cols
  A tbl containing the colors associated with each R-loop mapping mode in RLHub::rlbase_samples.

- heat_cols
  A tbl containing the colors associated with user-supplied data and RLBase data when running corrHeatmap.

- label_cols
  A tbl containing the colors associated with the labels in RLBase. See also RLHub::rlbase_samples.

- prediction_cols
  A tbl containing the colors associated with the predictions in RLBase. See also RLHub::rlbase_samples.

- prediction_label_cols
  A tbl containing the colors associated with the prediction-label combinations in RLBase. See also RLHub::rlbase_samples.

- available_modes
  A tbl containing the modes available in RLBase and associated metadata. See also RLHub::rlbase_samples.

- available_genomes
  A character showing all the official UCSC genomes available for use with RLSeq. See also available_genomes.

- misc_modes
  A character showing the R-loop mapping modes that are lumped into the 'misc' category for simplification of plotting.

Examples
auxdata
Available Genomes

Description

Contains metadata about all the genomes available in UCSC. It contains derived metadata, such as the effective genome sizes as well. See also the data-raw/available_genomes.R script to see processing steps.

Usage

available_genomes

Format

An object of class data.frame with 199 rows and 27 columns.

Details

Structure:

available_genomes is a data.frame with the following columns:

- UCSC_orgID
  - Official UCSC ID of the genome
- description
  - Verbose description of the assembly, source, and year/month of entry.
- nibPath
  - Endpoint of the genome in UCSC gdbb.
- organism
  - Name of the organism.
- defaultPos
  - Default location of genome browser view for this genome.
- active
  - Description not available.
- orderKey
  - Description not available.
- genome
  - The name of the genome.
- scientificName
  - The scientific name of the organism.
- htmlPath
  - Path in UCSC gdbb to the description.html file for the genome.
- hgNearOk
  - Description not available.
checkRLFSAnno

- hgPbOk
  - Description not available.
- sourceName
  - Name of organization providing the genome.
- taxId
  - The taxonomy ID of the organism.
- genes_available
  - If TRUE, the gene annotations are available in GTF format.
- year
  - The year the genome assembly was added.
- eff Genome_size_XXbp
  - The effective genome size of this genome. Calculated at various read lengths with khmer and used to improve the accuracy of analysis. See the data-raw/available_genomes.R script to see how this calculation was performed.
- genome_length
  - The total length of the genome.
- rlfs_available
  - If TRUE, R-loop forming sequences annotations are available in the RLBase AWS S3 repository.

Examples

available_genomes

checkRLFSAnno  Check RLFS

Description

Helper function that checks whether a genome has RLFS available.

Usage

checkRLFSAnno(genome)

Arguments

genome the UCSC genome name to check

Value

A logical, TRUE if available, FALSE if not
corrAnalyze

**Analyze Correlations**

**Description**

Finds the pairwise correlation in signal around gold-standard R-Loop sites between the query sample and the coverage tracks in the RLBase database. See details.

**Usage**

corrAnalyze(object, force = FALSE)

**Arguments**

- **object** An RLRanges object.
- **force** Force corrAnalyze to run, even if on Windows. Default: FALSE.

**Details**

Currently, this does not work on windows.

**Method:**

The corrAnalyze function performs a correlation test that can be used to assess sample-sample similarity by calculating coverage signal (from genomic alignments) around “gold standard” R-loop sites (PMID: 33411340). The resulting correlation matrix is useful for determining how well a supplied sample correlates with previously-published datasets.

During the RLBase-data workflow, the signal for each R-loop mapping sample within “gold standard” R-loop sites was calculated see RLHub::gs_signal.

The corrAnalyze function loads RLHub::gs_signal and accepts an RLRanges object with a valid coverage slot. It then does the following:

1. The coverage is quantified within the “gold standard” sites and added as a column to the signal matrix from RLHub::gs_signal.
2. Then, the stats::cor function is used to calculate the Pearson correlation pairwise between all samples, yielding a correlation matrix.
3. Finally, the correlation matrix is stashed in the in the correlationMat slot of the RLResults and returned.

**Value**

An RLRanges object with correlation results included as a matrix. The correlation matrix is accessed via rlresults(object, "correlationMat").
Example RLRanges object
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# corrAnalyze does not work on Windows OS
if (.Platform$OS.type != "windows") {
    # run corrAnalyze
    rlr <- corrAnalyze(rlr)
}

corrHeatmap

Plot Correlation Results

Description
Plots a heatmap to visualize the pairwise Pearson correlation matrix generated via corrAnalyze.

Usage
corrHeatmap(object, returnData = FALSE, complex = TRUE, ...)

Arguments

object An RLRanges with corrAnalyze already run.
returnData If TRUE, plot data is returned instead of plotting. Default: FALSE
complex If TRUE, ComplexHeatmap::Heatmap will be used for plotting. Otherwise, pheatmap::pheatmap is used. Default: TRUE
...

Value
A plot object or plotting data (if returnData is TRUE).

Examples
# Example RLRanges data with corrAnalyze() already run.
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# Corr heatmap
corrHeatmap(rlr)
featureEnrich

Test Genomic Feature Enrichment

Description
Tests the enrichment of genomic features in supplied peaks. See details.

Usage

```r
featureEnrich(
  object,
  annotype = c("primary", "full"),
  annotations = NULL,
  downsample = 10000,
  quiet = FALSE
)
```

Arguments

- `object`: An RLRanges object.
- `annotype`: The type of annotations to use. Can be one of "primary" or "full". Default: "primary". See RLHub::annotations for greater detail.
- `annotations`: A custom annotation list of the same structure described in RLHub::annotations.
- `downsample`: If a numeric, data will be down sampled to the requested number of peaks. This improves the speed of genomic shuffling and helps prevent p-value inflation. If FALSE, then downsampling will not be performed. Default: 10000.
- `quiet`: If TRUE, messages will be suppressed. Default: FALSE

Details

**Method:**
Annotations relevant to R-loops were curated as part of the RLBase-data workflow and are provided via RLHub::annotations.

In featureEnrich, each annotation "type" (e.g., "Exons", "Introns", etc) is compared to the supplied RLRanges, yielding enrichment statistics with the following procedure:

1. For each annotation type, the peaks are overlapped with the annotations.
2. Then, valr::bed_reldist is used to find the relative distance distribution between the peaks and the annotations for both the supplied RLRanges and shuffled RLRanges (via valr::bed_shuffle). Significance of the relative distance is calculated via stats::ks.test.
3. Then, Fisher’s exact test is implemented via valr::bed_fisher to obtain the significance of the overlap and the odds ratio.

**Value**
An RLRanges object containing the results of the enrichment test accessed via rlresult(object, "featureEnrichment"). The results are in tbl format. For a full description of all columns in the output table see RLHub::feat_enrich_samples.
Examples

# Example RLRanges dataset
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# RL Region Test
featureEnrich(rlr)

# With custom annotations
small_anno <- list(
  "Centromeres" = readr::read_csv(
    system.file("extdata", "Centromeres.csv.gz", package = "RLSeq"),
    show_col_types = FALSE
  )
)
featureEnrich(rlr, annotations = small_anno)


description

The core plotting component of plotEnrichment

Usage

feature_ggplot(x, usamp, limits, splitby)

Arguments

x A tbl containing data for plotting.
usamp The name of the user-supplied sample
limits Specify limits on data. Useful for controlling infinite estimation of odds ratio resulting from fisher's exact test. To remove limits, set c(-Inf, Inf). Default: c(-10, 15).
splitby Metadata by which to split plots. Can be "none", "prediction", or "label".

Value

A ggplot2 object.
geneAnnotation  
Annotate R-Loops with Genes

Description
Annotates RLRanges with entrez ids for overlapping genes. See details.

Usage
geneAnnotation(object, txdb = NULL)

Arguments

object  
An RLRanges object.

txdb  
The TxDb or EnsDb object containing gene annotations. If not supplied, annotations will be automatically downloaded from AnnotationHub. See also GenomicFeatures::TxDb.

Details
The geneAnnotation function provides a simple procedure for annotating RLRanges with gene IDs by overlap.

Annotations:
First, gene annotations are automatically downloaded using AnnotationHub::query with the following pattern:

```
AnnotationHub::query(
    x = ah,
    pattern = c("TxDb", "UCSC", "knownGene", genome)
)
```

Where genome is the UCSC genome id for the RLRanges object. If these annotations are unavailable, they should be provided using the txdb parameter. See also GenomicFeatures::TxDb.

Overlaps:
The annotations are subsequently overlapped with the ranges in the supplied RLRanges object using valr::bed_intersect and saved in the RLResults object as a tbl with a mapping of peak names to gene_id (entrez gene IDs).

Value
An RLRanges object with gene overlaps included. The results are available via rlresult(object, "geneAnnoRes"). The result object is a tbl with a mapping of peak_name (peak names from names(object)) to gene_id (entrez gene IDs).
genomeMasks

Examples

```r
# Example RLRanges data
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# Perform gene annotation
rlr <- geneAnnotation(rlr)

# Supply custom TxDb if needed
if (GenomeInfoDb::genome(rlr)[1] == "hg19") {
  library(TxDB.Hsapiens.UCSC.hg19.knownGene)
  rlr <- geneAnnotation(rlr, txdb = TxDb.Hsapiens.UCSC.hg19.knownGene)
}
```

---

<table>
<thead>
<tr>
<th>genomeMasks</th>
<th>Genome Masks</th>
</tr>
</thead>
</table>

Description

A collection of genome masks for use with `analyzeRLFS`. See the `data-raw/genome_masks.R` script for the processing steps.

Usage

genomeMasks

Format

An object of class list of length 8.

Details

Structure:

`genomeMasks` is a named list of GRanges objects. Each entry in the list follows the naming convention: `<genome>.masked`, where `<genome>` is an official UCSC genome ID. Each entry contains a GRanges object with the masked ranges from `<genome>`. The genomes provided correspond to the masked genomes available in `BSgenome::available.genomes`.

Examples

genomeMasks
getChromSizes  
*Get Chrom Sizes*

**Description**
Helper function which extracts chrom sizes from an RLRanges object.

**Usage**
```r
getChromSizes(object)
```

**Arguments**
- `object` An RLRanges object.

**Value**
A tibble containing chrom sizes.

---

getGSSignal  
*Get GS Signal*

**Description**
Extract signal around "gold-standard" R-loop sites.

**Usage**
```r
g getSsignal(coverage, gssignal)
```

**Arguments**
- `coverage` The path to a .bigWig file (can be a URL).
- `gssignal` The GS signal obtained from RLHub.

**Value**
A named list containing the results of correlation analysis.
**getRLFSAnno**  
*Get RLFS*

**Description**
Helper function that retrieves R-loop-forming sequences as GRanges

**Usage**
```r
getRLFSAnno(object)
```

**Arguments**
- `object` An RLRanges object.

**Value**
A GRanges object with RLFS for that species.

---

**noiseAnalyze**  
*Analyze sample noise*

**Description**
Analyzes the noiseness of the supplied sample using the method described by Diaz et al.. See details.

**Usage**
```r
noiseAnalyze(object, windows = NULL, force = FALSE)
```

**Arguments**
- `object` An RLRanges object.
- `windows` Genomics windows to use for quantifying signal. Will be automatically supplied if not provided. It is recommended NOT to specify this option for most analysis types, as doing so will impair the ability to compare to RLBase samples. Default: NULL.
- `force` Force `noiseAnalyze` to run, even if on Windows. Default: FALSE.
Details

Currently, this does not work on windows.

Method:
The method used for noise analysis is a minor modification of the method developed by Diaz et al., 2012 and also implemented by the deepTools function, plotFingerprint. Briefly, if user-supplied RLRanges contain a bigWig coverage file, then the coverage is quantified within random genomic regions (randomWindows). The regions are then ranked. A good signal-to-noise ratio will yield a distribution where most bins have little coverage but a few have very high coverage. Use downstream tools like plotNoise and plotCompareNoise to visualize these results.

Value

An RLRanges object with noise analysis results included as a tbl. The result is accessed via rlresults(object, "noiseAnalysis").

Examples

```r
# Example RLRanges object
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# noiseAnalyze does not work on Windows OS
if (.Platform$OS.type != "windows") {
    # run noiseAnalyze
    rlr <- noiseAnalyze(rlr)
}
```

---

noiseComparisonPlot  
**Creates a metaplot for comparing noise analysis results with RLBase**

Description

Plots the average standardized signal from noiseAnalyze alongside the samples in RLBase. For this plot, lower average signal indicates better signal to noise ratio. **Note:** This plot may be misleading if you supplied custom windows when running noiseAnalyze.

Usage

`noiseComparisonPlot(object, mode = "auto", simple = TRUE, returnData = FALSE)`

Arguments

- **object**
  An RLRanges object with noiseAnalyze already run.

- **mode**
  A character containing the R-loop data mode to compare against. See details for more information.
peak_stats

  simple     A logical which specifies whether the plot should only show samples where the prediction and label are the same. Default: TRUE.
  returnData If TRUE, plot data is returned instead of plotting. Default: FALSE

Details

  Mode:
  The mode parameter specifies the R-loop modality to compare the user-supplied sample against in the plot. The default, "auto" specifies that the mode from the supplied RLRanges object will be used. Only one mode can be specified. For a list of applicable modes, see auxdata$available_modes.

  Plot:
  The plot is a violin / jitter plot showing the distribution of average values from the noiseAnalyze output across RLBase samples of the selected mode. The user-supplied sample is annotated on the plot.

Value

  A ggplot2::ggplot object or a tbl if returnData is TRUE.

Examples

  rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

  # Plot RL-Region overlap
  noiseComparisonPlot(rlr)

  # Return data only
  noiseComparisonPlot(rlr, returnData = TRUE)

---

peak_stats Build peak statistics tibble

Description

  A helper function for building the peak statistics tibble

Usage

  peak_stats(x, xshuff, y, chromSizeTbl, quiet = FALSE)

Arguments

  x     The R-loop peaks to test.
  xshuff x, but shuffled around the genome to build a control peakset.
  y     The annotations against which to test x.
  chromSizeTbl A tbl containing the sizes of each chromosome in x and y.
  quiet  If TRUE, messages will be suppressed. Default: FALSE
plotEnrichment

Value

A tibble containing the test results.

plotEnrichment  
Plot Enrichment Test Results

Description

Creates a list of plots, one for each annotation database (see RLHub::annotations). These plots show the feature enrichment for the user-supplied sample in comparison to the samples in RLBase. This will only work if you did not use custom annotations with featureEnrich.

Usage

plotEnrichment(
  object,
  pred_POS_only = TRUE,
  label_POS_only = FALSE,
  splitby = c("none", "prediction", "label"),
  limits = c(-10, 15),
  returnData = FALSE,
  ...
)

Arguments

object          An RLRanges object with featureEnrich already run.
pred_POS_only   If TRUE, only "POS" predicted samples included (see also predictCondition). Default: TRUE.
label_POS_only  If TRUE, only "POS" labeled samples included (samples which are expected to robustly map R-loops, e.g., "D210N" condition R-ChIP data). Default: FALSE.
splitby         Metadata by which to split plots. Can be "none", "prediction", or "label".
limits          Specify limits on data range. This is used for controlling the infinite estimation of odds ratio resulting from fisher’s exact test. To remove limits, set c(-Inf, Inf). Default: c(-10, 15).
returnData      If TRUE, plot data is returned instead of plot objects. Default: FALSE
...              For internal use.

Value

A named list of ggplot2::ggplot objects. Names correspond to the annotations provided. See also featureEnrich.
Examples

# Example dataset with featureEnrich() already run.
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# Make plots, split by prediction
plotEnrichment(rlr, pred_POS_only = FALSE, splitby = "prediction")

plotFingerprint  
Plot noise analysis results as a fingerprint plot

Description

Plots the results of the noise analysis in noiseAnalyze. Creates a fingerprint plot like those developed by Diaz et al, 2012 and those provided by deepTools.

Usage

plotFingerprint(object)

Arguments

object  
An RLRanges object with noiseAnalyze already run.

Details

The term "Fingerprint plot" comes from deepTools.

Value

A ggplot object. See also ggplot2::ggplot.

1. noiseComparisonPlot

• A plot showing the noise analysis results from the user-supplied sample compared to similar samples from RLBase.

Examples

# Example RLRanges dataset with analyzeRLFS() already run.
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# Plot RLFS res
plotFingerprint(rlr)
**plotRLFSRes**  

**Plot RLFS analysis results**

**Description**

Plots the results of the R-loop-forming sequences (RLFS) analysis. The plot is a metaplot of the Z score distribution around RLFS with the p value from permutation testing annotated. See also `analyzeRLFS`.

**Usage**

```r
plotRLFSRes(object, plotName = NULL, fft = FALSE, ...)
```

**Arguments**

- `object`: An RLRanges object with `analyzeRLFS` already run. Alternatively, can be the RLFS results object from an RLRanges (from `rlresult(object, "rlfsRes")`).
- `plotName`: A Sample name used for plotting. If blank, the RLRanges `sampleName` metadata entry is used (see RLRanges).
- `fft`: If TRUE, the Fourier transform of the Z-score is plotted instead. Default: FALSE.
- `...`: Additional parameters passed to `ggplot2::ggplot`.

**Value**

A ggplot object. See also `ggplot2::ggplot`.

**Examples**

```r
# Example RLRanges dataset with analyzeRLFS() already run.
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# Plot RLFS res
plotRLFSRes(rlr)

# Plot the Fourier transform instead
plotRLFSRes(rlr, fft = TRUE)
```
Description

Convenience function for plotting the overlap between RLRanges and R-loop regions (RL regions) as calculated by rlRegionTest.

Usage

plotRLRegionOverlap(object, returnData = FALSE, rlregions_table = NULL, ...)

Arguments

object
An RLRanges object with rlRegionTest already run.

returnData
If TRUE, plot data is returned instead of plotting. Default: FALSE

rlregions_table
The table of RLRegions to overlap sample ranges with. Obtained from RLHub using RLHub::rlregions_meta. Loaded from RLHub if not supplied. Default: NULL.

Additional arguments passed to VennDiagram::venn.diagram

Value

A ggplot2::ggplot object containing the venn diagram. Built using ggplotify::as.ggplot.

Examples

# Example dataset with rlRegionTest() already run.
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# Plot RL-Region overlap
plotRLRegionOverlap(rlr)

# Return data only
plotRLRegionOverlap(rlr, returnData = TRUE)

Description

Plots the results of txFeatureOverlap alongside the average from public R-loop datasets. This allows comparison of user-supplied samples with data that is expected to be similar.
Usage

plotTxFeatureOverlap(object, mode = "auto", returnData = FALSE)

Arguments

object An RLRanges object with txFeatureOverlap already run.
mode A character containing the R-loop data mode to compare against. See details for more information.
returnData If TRUE, plot data is returned instead of plotting. Default: FALSE

Details

Mode:
The `mode` parameter specifies the R-loop modality to compare the user-supplied sample against in the plot. The default, "auto" specifies that the `mode` from the supplied RLRanges object will be used. Only one mode can be specified. For a list of applicable modes, see auxdata$available_modes.

Plot:
The plot is a stacked bar chart showing the proportion of peaks overlapping transcript features for the supplied RLRanges object. Additionally, the average of the txFeatureOverlap analysis for all samples within the specified modes are also shown as a background comparison. This style of analysis enables a user to see the transcript features overlapping their peaks and compare those results to the average within relevant public datasets.

Value

A ggplot2::ggplot object or a tbl if returnData is TRUE.

Examples

rlr <- readRDS(system.file("extdata", "rlrsml.rds", package = "RLSeq"))

# Plot RL-Region overlap
plotTxFeatureOverlap(rlr)

# Return data only
plotRLRegionOverlap(rlr, returnData = TRUE)

---

predictCondition  Predict Condition

Description

Uses the results of analyzeRLFS to predict whether a sample is "POS" (robust R-loop mapping) or "NEG" (poor R-loop mapping). See details.
Usage

predictCondition(object, rlfsRes = NULL, ...)

Arguments

object       An RLRanges object with `analyzeRLFS` already run. Ignored if rlfsRes is provided.
rlfsRes      If object not supplied, provide the rlfsRes list which is obtained from `rlresult(object, "rlfsRes")`.
...          Internal use only.

Details

Following R-loop forming sequences (RLFS) analysis, the quality model (see `RLHub::models`) is implemented for predicting the sample condition in coordination with other results from `analyzeRLFS`. A prediction of “POS” indicates robust R-loop mapping, whereas “NEG” indicates poor R-loop mapping. The succeeding sections describe this process in greater detail.

Application of binary classification model:
First, the binary classifier is applied, yielding a preliminary prediction of quality. This is accomplished via the following steps:

1. Calculate the Fourier transform of the Z-score distribution (see `analyzeRLFS`).
2. Reduce the dimensions to the engineered feature set (see table below).
3. Apply the preprocessing model (see `RLHub::models`) to normalize these features
4. Apply the classifier (see `RLHub::models`) to render a quality prediction.

Engineered feature set:
Abbreviations: Z, Z-score distribution; ACF, autocorrelation function; FT, Fourier Transform.

<table>
<thead>
<tr>
<th>feature</th>
<th>description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Z1</td>
<td>mean of Z</td>
</tr>
<tr>
<td>Z2</td>
<td>variance of Z</td>
</tr>
<tr>
<td>Zac1</td>
<td>mean of Z ACF</td>
</tr>
<tr>
<td>Zac2</td>
<td>variance of Z ACF</td>
</tr>
<tr>
<td>ReW1</td>
<td>mean of FT of Z (real part)</td>
</tr>
<tr>
<td>ReW2</td>
<td>variance of FT of Z (real part)</td>
</tr>
<tr>
<td>ImW1</td>
<td>mean of FT of Z (imaginary part)</td>
</tr>
<tr>
<td>ImW2</td>
<td>variance of FT of Z (imaginary part)</td>
</tr>
<tr>
<td>ReWac1</td>
<td>mean of FT of Z ACF (real part)</td>
</tr>
<tr>
<td>ReWac2</td>
<td>variance of FT of Z ACF (real part)</td>
</tr>
<tr>
<td>ImWac1</td>
<td>mean of FT of Z ACF (imaginary part)</td>
</tr>
<tr>
<td>ImWac2</td>
<td>variance of FT of Z ACF (imaginary part)</td>
</tr>
</tbody>
</table>

Final quality prediction:
The results from the binary classifier are combined with other results from `analyzeRLFS` to yield a final prediction. To yield a prediction of “POS” all the following must be TRUE:
1. The RLFS Permutation test P value is significant ($p < .05$). Stored as \texttt{PVal Significant} in the results object.
2. The Z-score distribution at 0bp is $> 0$. Stored as \texttt{ZApex > 0} in the results object.
3. The Z-score distribution at 0bp is $> \text{the start and the end}$. Stored as \texttt{ZApex > ZEdges} in the results object.
4. binary The classifier predicts a label of “POS”. Stored as \texttt{Predicted 'POS'} in the results object.

**Value**

An RLRanges object with predictions accessible via \texttt{rlresult(object, "predictRes")}.

**Structure:**

The results object is a named list of the structure:

- **Features**
  - A tbl with three columns that describe the engineered features used for prediction:
    - \texttt{feature}: the name of the feature (see \texttt{details}).
    - \texttt{raw_value}: The raw value of that feature in the supplied object.
    - \texttt{processed_value}: The normalized value of that feature after preprocessing (see \texttt{details}).

- **Criteria**
  - The four criteria which must all be \texttt{TRUE} to render a prediction of "POS" (see \texttt{details}).

- **prediction**
  - The final prediction. "POS" indicates robust R-loop mapping, "NEG" indicates poor R-loop mapping.

**Examples**

```r
# Example data with analyzeRLFS already run
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# predict condition
rlr <- predictCondition(rlr)

# With rlfsRes
predRes <- predictCondition(rlfsRes = rlresult(rlr, "rlfsRes"))
```

**randomWindows**

**Random Genomic Windows**

**Description**

A collection of random genomic windows for use with \texttt{noiseAnalyze}. See the \texttt{data-raw/genome_masks.R} script for the processing steps.
Usage

randomWindows

Format

An object of class list of length 3.

Details

Structure:
randomWindows is a named list of tbl objects containing ~1000 random genomic regions. The names are the genomes to which the regions correspond.

Columns (mirrors bed3 format).

1. chrom - the chromosome in UCSC format
2. start - the starting position
3. end - the end position

Examples

randomWindows

Description

Builds an HTML report to showcase the results available in the supplied RLRanges object (see also RLRresults).

Usage

report(
  object,
  reportPath = "rlreport.html",
  intermediates_dir = NULL,
  quiet = FALSE,
  ...
)
Arguments

object  An RLRanges object.
reportPath  A path indicating the report output HTML file. Default: "rlreport.html"
intermediates_dir  A directory indicating where intermediate files should be stored during report building. If not set, a random directory in tmp/ will be used. Default: NULL.
quiet  If TRUE, messages are suppressed. Default: FALSE.
...

Arguments passed to rmarkdown::render()

Value

TRUE

Examples

# Example data with RLSeq() already run.
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# Get a TMP file (only for example use)
tmp <- tempfile()

# Generate the report
report(rlr, reportPath = tmp)

rlbaseNoiseAnalyze  RLBase Sample Noise Analysis Results

Description

Average signal from noise analysis of RLBase samples. See noiseAnalyze for more detail regarding how signal was initially calculated.

Usage

rlbaseNoiseAnalyze

Format

An object of class tbl_df (inherits from tbl.data.frame) with 704 rows and 2 columns.

Details

Structure:
rlbaseNoiseAnalyze is a tbl with the following columns:

• rlsample
  – The RLBase sample identifier for the sample.
RLRanges-class

- Matches the rlsample column in RLHub::rlbase_samples
  - value
- The mean signal from noise analysis. See noiseAnalyze.

Examples

rlbaseNoiseAnalyze

---

**RLRanges-class**

**Construct RLRanges Dataset**

**Description**

RLRanges is a subclass of GRanges, which stores R-loop peaks and metadata about the R-loop-mapping experiment, along with results from the analyses in RLSeq.

**Usage**

```r
RLRanges(
  peaks = GenomicRanges::GRanges(),
  coverage = character(1),
  genome = character(1),
  mode = character(1),
  label = character(1),
  sampleName = "User-selected sample",
  qcol = NULL
)
```

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>peaks</td>
<td>Path/URL to peak file or a GRanges object. This file should be in &quot;broadPeak&quot; format if possible. If not, then qcol should be specified.</td>
</tr>
<tr>
<td>coverage</td>
<td>Path/URL to the corresponding coverage file (in &quot;bigWig&quot; format). If not supplied, correlation tests will be skipped.</td>
</tr>
<tr>
<td>genome</td>
<td>UCSC genome ID. Acceptable types are listed in auxdata (available_genomes entry).</td>
</tr>
<tr>
<td>mode</td>
<td>Type of R-loop mapping from which peaks and coverage were derived. Acceptable types are listed in auxdata (available_modes entry). Can be unspecified.</td>
</tr>
<tr>
<td>label</td>
<td>&quot;POS&quot; (positive R-loop-mapping sample; e.g., DRIP-Seq S9.6 -RNH1) or &quot;NEG&quot; (negative control sample; e.g., DRIP-Seq S9.6 +RNH1 or Input). Can be unspecified.</td>
</tr>
<tr>
<td>sampleName</td>
<td>A unique name for identifying this sample. Can be unspecified.</td>
</tr>
</tbody>
</table>
qcol

The name of the metadata column which contains the score or significance of each peak. For broadPeak (preferred), this is the qvalue (column 11 after accounting for extra columns created during peakset building). If not specified, the last column will be chosen by default. **NOTE:** if supplying narrowPeak form peaks, the last column will NOT be appropriate and QCol should be specified as 11. If FALSE or if no metadata columns exist, it will be left blank and some operations in `report()` will not fully run.

**Value**

An object of class `RLRanges`. These objects are an extension of `GRanges` with the addition of sample metadata entries and `RLResults`.

**Examples**

```r
# Example dataset
rbase <- "https://rlbase-data.s3.amazonaws.com"
cvg <- file.path(rbase, "coverage", "SRX7671349_hg38.bw")
pks <- system.file("extdata", "SRX7671349_hg38.broadPeak", package = "RLSeq")

# Get RLRanges object
rlr <- RLRanges(pks,
               coverage = cvg, genome = "hg38", label = "NEG",
               mode = "RDIP", sampleName = "RDIP-Seq +RNH1", qcol = 9)
```

**Description**

Accessor function which returns any sample in RLBase as an `RLRanges` object for use with RLSeq. For a full list of available samples, see `RLHub::rlbase_samples`.

**Usage**

```r
RLRangesFromRLBase(acc, rlsamples = NULL)
```

**Arguments**

- `acc` The sample ID of the RLBase object. See the `rlsample` column in `RLHub::rlbase_samples`.
- `rlsamples` The tibble provided by `RLHub::rlbase_samples`. Providing these data ahead of time speeds up this operation. Default: NULL.

**Value**

An RLRanges object with all results available.
Examples

rlr <- RLRangesFromRLBase("SRX1070676")

Description

Tests the overlap of user-supplied ranges with R-loop regions (RL regions).

Usage

rlRegionTest(object)

Arguments

object An RLRanges object with genome "hg38".

Details

R-loop regions (RL regions) are consensus sites of R-loop formation. For more information, see RLHub::rlregions. The rlRegionTest is a simple function which finds the overlap of user-supplied samples with RL regions and calculates Fisher’s exact test via valr::bed_fisher.

Value

An RLRanges object with test results accessible via rlresult(object, "rlRegionRes").

Structure:
The structure of the results is a named list containing the following:

- Overlap
  - A tbl showing the overlap between RL regions and user-supplied ranges.
  - Column description:
    * chrom - The chromosome name
    * start__peaks - The starting position of the user-supplied peak in the overlap.
    * end__peaks - Same as above for end position.
    * name__peaks - The name of the user-supplied peak in the overlap (from names(object)).
    * start/end/name__rlregion - Same as above for RL regions.
    * strand__rlregion - The genomic strand of the RL region in the overlap.
    * .overlap - The size of the overlap.
- Test_results
  - A tbl showing the results of the Fisher’s exact test. See valr::bed_fisher.
Examples

```r
# Example RLRanges data
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# RL Region Test
rlRegionTest(rlr)
```

### Description

Functions for creating and accessing the R-loop results (RL Results). These are a type of object used for holding the results of the tests implemented in RLSeq. They can be accessed using the `rlresult` function.

### Usage

```r
rlresult(object, resultName)
```

### Arguments

- `object`: RLRanges object.
- `resultName`: Name of the result slot to access. See details.

### Details

**Slot descriptions:**

- `featureEnrichment`
  - The tbl generated from running `featureEnrich`.
  - The structure and column descriptions are provided in detail within `RLHub::feat_enrich_samples`.
- `correlationMat`
  - The matrix generated from running `corrAnalyze`.
  - Contains pairwise pearson correlations between all samples in `RLBase` and the supplied RLRanges object.
- `rlfsRes`
  - The list generated from running `analyzeRLFS`.
  - See `analyzeRLFS` for description of structure.
- `noiseAnalysis`
  - The tbl generated from running `noiseAnalyze`.
- `txFeatureOverlap`
  - The tbl generated from running `txFeatureOverlap`.
- `geneAnnoRes`
  - The tbl generated from running `geneAnnotation`.
• predictRes
  – The list generated from running `predictCondition`.
• rlRegionRes
  – The list generated from running `rlRegionTest`.

Value
The contents of the requested slot.

Examples
```r
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLseq"))
rlresult(rlr, "predictRes")
```

---

`rlsampleTxOl`  
 **RLBase Sample Transcript Feature Overlaps**

Description
Summary statistics from transcript feature overlap analysis of peaks from all RLBase samples.

Usage
`rlsampleTxOl`

Format
An object of class `tbl_df` (inherits from `tbl`, `data.frame`) with 4754 rows and 4 columns.

Details

**Structure:**
`rlsampleTxOl` is a `tbl` with the following columns:

• `rlsample`
  – The RLBase sample identifier for the sample.
  – Matches the `rlsample` column in `RLHub::rlbase_samples`
• `feature`
  – The transcript feature for which overlap analysis was performed.
  – These features were derived from the Transcript Features collection described in `RLHub::annotations`
• `n`
  – The raw number of peaks from the sample overlapping a feature.
• `pct`
  – The proportion of peaks from the sample overlapping a feature.
Examples

```r
rlsampleTxOl
```

---

### Description

Executes the RLSeq analysis workflow.

### Usage

```r
RLSeq(object, quiet = FALSE, skip = NULL, ...)
```

### Arguments

- **object**
  - An `RLRanges` object.

- **quiet**
  - If `TRUE`, messages are suppressed. Default: `FALSE`.

- **skip**
  - Analysis steps to skip. Default: `NULL`. See `details` for options.

- **...**
  - Arguments passed to `analyzeRLFS`.

### Details

The `RLSeq()` function does all of the following by default:

1. **RLFS Perm Test.** Runs the `analyzeRLFS` function to test the enrichment of user-supplied ranges within R-loop-forming sequences. *Cannot be skipped.*

2. **Predict Condition.** Runs the `predictCondition` function to predict whether the user-supplied sample robustly maps R-loops or not. *Cannot be skipped.*

3. **Noise analysis.** Runs the `noiseAnalyze` function to analyze the signal-noise distribution within the supplied dataset. Skip with `skip="noiseAnalyze"`.

4. **Feature enrichment test.** Runs the `featureEnrich` function to test the enrichment of user-supplied ranges within R-loop-relevant genomic features. Skip with `skip="featureEnrich"`.

5. **Transcript Feature Overlap.** Runs the `txFeatureOverlap` function to get the overlap of transcript features and user-supplied peaks.

6. **Correlation Analysis.** Runs the `corrAnalyze` function to test the correlation of user-supplied R-loop signal with other samples in RLBase around "gold-standard" R-loop regions. Skip with `skip="corrAnalyze"`.

7. **Gene annotation.** Runs the `geneAnnotation` function to find overlap of genes with the user-supplied ranges. Skip with `skip="geneAnnotation"`.

8. **R-loop Region Analysis.** Runs the `rlRegionTest` function to find the overlap of user-supplied ranges with consensus R-loop sites (RL-Regions). Skip with `skip="rlRegionTest"`. 
Table to Regions

Description

Helper function to Convert "table" format to "regions" format.

Usage

`tableToRegions(table)`

Arguments

- `table` A tibble in "Table" format from RLHub.

Value

A tibble in "regions" format.
txFeatureOverlap  

*Calculate overlap with transcript features*

**Description**

Tests the overlap of transcript features with supplied peaks. See *details*.

**Usage**

```r
txFeatureOverlap(object, quiet = FALSE)
```

**Arguments**

- `object`: An RLRanges object.
- `quiet`: If TRUE, messages will be suppressed. Default: FALSE

**Details**

**Method:**

Transcript annotations were curated as part of the RLBase-data workflow and are provided via RLHub::annotations.

In `txFeatureOverlap`, each annotation "type" (e.g., "Exons", "Introns", etc) is compared to the supplied RLRanges, yielding overlap statistics with the following procedure:

1. For each annotation type, the peaks are overlapped with the annotations.
2. Then the number of overlapping peaks is counted and summarised using a priority order. This order determines which feature is assigned to a peak when that peak overlaps multiple features. The order is "TSS", "TTS", "5'UTR", "3'UTR", "Exon", "Intron", "Intergenic".

**Value**

An RLRanges object containing the results of the enrichment test accessed via `rlresult(object, "txFeatureOverlap")`. The results are in tbl format.

**Examples**

```r
# Example RLRanges dataset
rlr <- readRDS(system.file("extdata", "rlrsmall.rds", package = "RLSeq"))

# RL Region Test
taxFeatureOverlap(rlr)
```
urlExists

Check if URL exists

Description

Check if URL exists

Usage

urlExists(urlcon)

Arguments

urlcon URL to check

Value

logical. TRUE if status code 200, FALSE if not

%>% Pipe operator

Description

Pipe operator

Usage

lhs %>% rhs

Arguments

lhs A value or the magrittr placeholder.

rhs A function call using the magrittr semantics.

Value

The result of calling rhs(lhs).
Index

* datasets
  auxdata, 4
  available_genomes, 6
genomeMasks, 13
randomWindows, 24
rlbaseNoiseAnalyze, 26
rlsampleTxOl, 31

* internal
  %>%, 35
%>%, 35
analyzeRLFS, 3, 13, 20, 22, 23, 30, 32
AnnotationHub::query, 12
auxdata, 4, 27
available_genomes, 5, 6
BSgenome::available.genomes, 13
checkRLFSAnno, 7
ComplexHeatmap::Heatmap, 9
corrAnalyze, 8, 9, 30, 32
corrHeatmap, 5, 9
feature_ggplot, 11
featureEnrich, 10, 18, 30, 32
geneAnnotation, 12, 30, 32
genomeMasks, 3, 13
GenomicFeatures::TxDb, 12
getChromSizes, 14
getGSSignal, 14
getRLFSAAnno, 15
ggplot2::ggplot, 17–22
ggplotify::as.ggplot, 21
noiseAnalyze, 15, 16, 17, 19, 24, 26, 27, 30, 32
noiseComparisonPlot, 16
peak_stats, 17
pheatmap::pheatmap, 9
plotEnrichment, 18
plotFingerprint, 19
plotRLFSRes, 20
plotRLRegionOverlap, 21
plotTxFeatureOverlap, 21
predictCondition, 4, 18, 22, 31, 32
randomWindows, 24
regioneR::circularRandomizeRegions, 4
regioneR::getMask, 3
regioneR::localZScore, 3, 4
regioneR::permTest, 3, 4
report, 25
rlbaseNoiseAnalyze, 26
RLHub::annotations, 5, 10, 18, 31, 34
RLHub::feat_enrich_samples, 10, 30
RLHub::gs_signal, 8
RLHub::models, 23
RLHub::rlbase_samples, 5, 27, 28, 31
RLHub::rlregions, 29
RLHub::rlregions_meta, 21
RLRanges, 3, 8, 16, 20, 25, 28, 30, 32, 33
RLRanges (RLRanges-class), 27
RLRanges-class, 27
RLRangesFromRLBase, 28
rLRegionTest, 21, 29, 31, 32
rlresult, 30, 33
RLResults, 8, 12, 25, 28
RLResults (rlresult), 30
RLResults-class (rlresult), 30
rlsampleTxOl, 31
RLSeq, 27, 32
stats::cor, 8
stats::ks.test, 10
tableToRegions, 33
txFeatureOverlap, 21, 22, 30, 32, 34
urlExists, 35

36
valr::bed_fisher, 10, 29
valr::bed_intersect, 12
valr::bed_reldist, 10
valr::bed_shuffle, 10
VennDiagram::venn.diagram, 21