

# Package ‘PloGO2’

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**Type** Package

**Title** Plot Gene Ontology and KEGG pathway Annotation and Abundance

**Version** 1.14.0

**Description** Functions for enrichment analysis and plotting gene ontology or KEGG pathway information for multiple data subsets at the same time. It also enables incorporating multiple conditions and abundance data.

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abundancePlot	<i>Function to summarize and plot abundance information from an annotations results list</i>
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---

### Description

Generates one GO/pathway abundance plot for each file, and (provided the number of GO/pathway categories is reasonably small) also an abundance plot for each GO/pathway category across all files provided. The abundance values are \*added\* for all values in one category. Hence if the initial values represented percentages such as NSAF, the final values represent percentages of the respective category.

### Usage

```
abundancePlot(res.list, log = FALSE, printLimit = 16, Group=NULL, Plot=FALSE,
CountCutOff=3, ...)
```

### Arguments

res.list	The result returned by processAnnotation
log	TRUE/FALSE: use raw or log abundance data
printLimit	The most number of category to be plotted
Group	The groups
Plot	To plot or not
CountCutOff	The minimum number of proteins in a category for it to be plotted
...	Parameters to pass

### Value

A list object, with the following values:

abundance	The abundance matrix
ag.mat	The aggregated abundance matrix
list.levelplots	A list of abundance levelplots trellis object
list.barplots	A list of abundance barcharts trellis object

**Author(s)**

D.Pascovici and J.Wu

**See Also**

See Also as [processAnnotation](#)

**Examples**

```
# get list of ID's
GOIDlist <- GOTermList("MF", 2)

# find existing files
path <- system.file("files", package="PloG02")
file.names <- paste(path, c("00100.txt", "01111.txt", "10000.txt",
"11111.txt","Control.txt"), sep="/")
datafile <- file.path(path, "NSAF.csv")

# summarize annotation
res.list <- processAnnotation(file.names, GOIDlist, data.file.name = datafile)

abundance.res <- abundancePlot(res.list)

# Plot levelplots
list.levelplots <- abundance.res$list.levelplots

for(i in seq_along(list.levelplots)) {
png(paste(names(list.levelplots)[i], ".png"), 2000, 4000, res=200)
print(list.levelplots[[i]])
dev.off()
}

# KEGG pathway
path <- system.file("files", package="PloG02")
file.names <- file.path(path,"PWFiles", c("AllData.txt","black.txt","blue.txt","brown.txt","green.txt",
"red.txt","turquoise.txt") )
datafile <- file.path(path,"Abundance_data.csv")
Group <- names(read.csv(datafile))[-1]

AnnotIDlist <- c("osa01100","osa01110","osa01230","osa00300","osa00860")

res.list <- processAnnotation(file.names, AnnotIDlist, data.file.name = datafile)

abundance.res <- abundancePlot(res.list, Group=Group, Plot=TRUE)

# Plot levelplots
list.levelplots <- abundance.res$list.levelplots

for(i in seq_along(list.levelplots)) {
png(paste(names(list.levelplots)[i], ".png"), 2000, 4000, res=200)
print(list.levelplots[[i]])
}
```

```

dev.off()
}
# Plot barchats by categories
list.barplots <- abundance.res$list.barplots

for(i in seq_along(list.barplots)) {
png(paste(names(list.barplots)[i], ".png"), 2000, 2000, res=200)
print(list.barplots[[i]])
dev.off()

}

```

---

annotationPlot	<i>Function to summarize and plot extracted GO or pathway annotation</i>
----------------	--

---

### Description

Summarize the information from the GO or pathway annotation list into a table of counts and percentages, and possibly print a few images.

### Usage

```
annotationPlot(res.list, percentages = FALSE, plot = TRUE, trimzero = FALSE, type=c("GO", "pathway"))
```

### Arguments

res.list	The list of summarized annotation as generated by <a href="#">processAnnotation</a> .
percentages	TRUE or FALSE.
plot	TRUE or FALSE: should plots be printed.
trimzero	TRUE or FALSE: should GO categories with no counts be removed. This is not really relevant when a small number of GO categories has been selected.
type	The type of annotation.

### Value

counts	A matrix of counts, GO categories (rows) by samples (cols).
percentages	A matrix of percentages, GO categories (rows) by samples (cols).

### Author(s)

D. Pascovici

### See Also

See Also [processAnnotation](#)

## Examples

```
## Not run:
# get list of ID's
GOIDlist <- GOTermList("BP", 2)

# find existing files
dir <- system.file("files", package="PloG02")
file.names <- paste(dir,c("00100.txt", "01111.txt", "10000.txt",
"11111.txt","Control.txt"), sep="/")

# summarize annotation
res.list <- processAnnotation(file.names, GOIDlist)

annotationPlot(res.list, plot=FALSE)

## End(Not run)
# KEGG pathway
dir <- system.file("files", package="PloG02")
fname <- file.path(dir,"PWFiles", c("red.txt", "blue.txt", "yellow.txt", "green.txt", "turquoise.txt") )
datafile <- file.path(dir,"Abundance_data.csv")

AnnotIDlist <- c("osa01100","osa01110","osa01230","osa00300","osa00860")

res.list <- processAnnotation(fname, AnnotIDlist, data.file.name = datafile)

annotationPlot(res.list, plot=TRUE, type="pathway")
```

---

compareAnnot

*Function to compare annotation percentages*

---

## Description

Compare annotation percentages by means of Fisher's exact test. A reference must be selected, and that name must be amongst the annotation result list names.

## Usage

```
compareAnnot(res.list, referenceName, removeZeros = FALSE, correction = TRUE)
```

## Arguments

res.list	List returned by the processAnnotation function
referenceName	Name of the condition to compare with
removeZeros	Remove the categories with no annotation in them from the result
correction	TRUE or FALSE: apply the BH fdr correction to the p-values in each column

**Value**

A matrix of p-values or NA. A comparison is made for each sample other than the reference with the reference, and each GO annotation category. The result is not recorded if the annotation numbers were small for that category (<5). This is strictly not needed for Fisher's exact test (though was needed for the chi-square approximation used initially).

**Author(s)**

D.Pascovici

**See Also**

See Also [processAnnotation](#)

**Examples**

```
## Not run:
# get list of ID's
GOIDlist <- GOTermList("BP", 2)

# find existing files
dir <- system.file("files", package="PloGO2")
file.names <- paste(dir,c("00100.txt", "01111.txt", "10000.txt",
"11111.txt","Control.txt"), sep="/")

# summarize annotation
res.list <- processAnnotation(file.names, GOIDlist)

# compare with Control
compareAnnot(res.list, "Control")

compareAnnot(res.list, "Control", correction=FALSE)

## End(Not run)
```

---

ExcelToPloGO

*Function to add GO annotation to an Excel spread sheet*

---

**Description**

Function to add GO annotation to an Excel spread sheet

**Usage**

```
ExcelToPloGO(fname, colName = "Uniprot", termFile= NA, compareWithReference="none", data.file.name = "
```

**Arguments**

fname	Name of the Excel spread sheet to be annotated
colName	The name of the column containing the protein identifiers
termFile	The name of the file containing the GO categories
compareWithReference	The name of the tab that serves as basis for enrichment comparison.
data.file.name	Abundance data for PloGO if any
outFolder	The output files folder

**Value**

A list object, with the following values:

Counts	The GO counts matrix summarized for all files
Percentages	The GO percentages matrix summarized for all files
Abundance	The GO percentages matrix summarized for all files, <b>ONLY</b> generated if an abundance file was provided
FisherPval	The Fisher p-values matrix summarized for all files, <b>ONLY</b> generated if a reference file was provided
res.list	The full list result of processAnnotation function
list.levelplots	The abundance levelplots if the data.file.name is not "none"
list.barplots	The abundance barcharts if the data.file.name is not "none"

**Author(s)**

D.Pascovici

**Examples**

```
## Not run:
# where sample files are stored
path <- system.file("files", package = "PloGO2")
termFile = paste(path, "GODefault.txt", sep="/")
xlfile <- paste(path, "ResultsWGCNA_Input4PloGO2.xlsx", sep="/")

res <- ExcelToPloGO(xlfile, termFile=termFile, compareWithReference="AllData")

## End(Not run)
```

---

ExcelToPloPathway      *Function to add KEGG pathway annotation to an Excel spread sheet*

---

### Description

Function to add KEGG pathway annotation to an Excel spread sheet

### Usage

```
ExcelToPloPathway(fname, colName = "Uniprot", compareWithReference = "none", DB.name = "pathwayDB.csv")
```

### Arguments

fname	Name of the Excel spread sheet to be annotated
colName	The name of the column containing the protein identifiers
compareWithReference	The name of the tab that serves as basis for enrichment comparison.
DB.name	The DB file name for the pathway
data.file.name	Abundance data for PloGO if any
outFolder	The output files folder

### Value

A list object, with the following values:

Counts	The pathway counts matrix summarized for all files
Percentages	The pathway percentages matrix summarized for all files
Abundance	The pathway percentages matrix summarized for all files, ONLY generated if an abundance file was provided
aggAbundance	Aggregated abundance matrix by combination of tab and abundance file columns for all pathways
FisherPval	The Fisher p-values matrix summarized for all files, ONLY generated if a reference file was provided
res.list	The full list result of processAnnotation function
list.levelplots	The abundance levelplots if the data.file.name is not "none"
list.barplots	The abundance barcharts if the data.file.name is not "none"

### Examples

```
path <- system.file("files", package = "PloGO2")

res <- ExcelToPloPathway(file.path(path, "ResultsWGCNA_Input4PloGO2.xlsx"),
  colName="Uniprot", compareWithReference="AllData", DB.name=file.path(path, "pathwayDB.csv"),
  data.file.name = file.path(path, "Abundance_data.csv") )
```

---

genAnnotationFiles	<i>Function to generate a format in Wego native style from a list of Uniprot identifiers</i>
--------------------	--

---

### Description

Given a Excel spreadsheet with multiple tabs, generate a Wego file for each tab using a pre-download DB file.

### Usage

```
genAnnotationFiles(fExcelName, colName="Uniprot",  
DB.name = "pathwayDB.csv", folder="PWFiles",outFolder=tempdir())
```

### Arguments

fExcelName	An excel file containing one or multiple tabs of protein IDs. For example, the proteins in each tab come from the same cluster.
colName	The column name of the protein ID.
DB.name	The database file name, in .csv format.
folder	The folder name for saving the generated files.
outFolder	The output files folder

### Value

The folder path for generated annotation files.

### Author(s)

J. Wu

### Examples

```
path <- system.file("files", package = "PloG02")  
  
genAnnotationFiles(fExcelName = file.path(path, "ResultsWGCNA_Input4PloG02.xlsx"),  
colName="Uniprot",  
DB.name = file.path(path, "pathwayDB.csv"))
```

---

genWegoFile	<i>Function to generate a format in Wego native style from a list of Uniprot identifiers</i>
-------------	--

---

### Description

Connects to the Uniprot Biomart using functionality from the biomaRt package, downloads GO information and organizes it as needed.

### Usage

```
genWegoFile(IDList, fname = "Wego.txt", database = "uniprot", outFolder=tempdir())
```

### Arguments

IDList	A list of ID's, either Ensembl or Uniprot.
fname	The name of the text file to be outputed.
database	"ensembl" or "uniprot".
outFolder	The output files folder

### Value

File path of generated Wego files

### Author(s)

D. Pascovici

### Examples

```
v <- c("Q9HWC9", "Q9HWD0", "Q9I4N8", "Q9HW18", "Q9HWC9", "Q9HWD0")
## Not run: genWegoFile(v, fname = "F1.txt")
```

---

getGoID	<i>Function to map a vector of GO terms to the corresponding GO ID's</i>
---------	--

---

### Description

This function is rather slow and inefficient, as it first extracts all GO terms then matches the current one. However, it is only intended for a one off matching of terms of interest to the respective nodes.

### Usage

```
getGoID(v)
```

**Arguments**

v                    the precise spelling of the go term

**Value**

The list of matched terms, with names the respective GO ID's/

**Author(s)**

D. Pascovici

**Examples**

```
getGoID(c("biological_process", "transport"))

# however the next one is not found as the proper term name has an
# underscore "cellular_component"
getGoID("cellular component")
```

---

GOTermList	<i>Function to extract a list of GO terms at level 2,3 or 4 of the GO hierarchy</i>
------------	---

---

**Description**

At the moment a quick and dirty way to extract all GO nodes at levels 2, 3 or 4 of the GO hierarchy.

**Usage**

```
GOTermList(ontology = "BP", level = 2, node = NULL)
```

**Arguments**

ontology	Either "BP", "CC" or "MF"
level	Either 2, 3 or 4
node	Null or a GO node

**Details**

This is a quick and dirty way to extract a list of GO terms of interest. That can be one of the levels 2, 3 or 4 or all subnodes (children) of a particular node if a valid GO is provided for the node parameter. Should be rewritten.

**Value**

A vector of GO nodes

**Author(s)**

D. Pascovici

**Examples**

```
GOTermList("BP", 2)
GOTermList("CC", 2)
```

---

PloGO

---

*Function to do steps of GO annotation and annotation/abundance plots*


---

**Description**

Summarize GO categories for all the files in the zip, if provided merge data from data file, generate annotation and abundance plots and comparison with reference.

**Usage**

```
PloGO(zipFile = "none", termFile="none", ontology = "BP", ontologyLevel = 2, reference = "none", data.f
filesPath=".", node=NULL, aggregateFun="sum", logAb=FALSE, ...)
```

**Arguments**

zipFile	Zip containing all the GO files
termFile	A file with the GO terms of interest, if a limited set provided
ontology	Wither BP, MF or CC
ontologyLevel	A small level such as 2 or 3
reference	The file name of the reference file, if any, for instance "Control" for Control.txt
data.file.name	The file containing all the experimental data, for instance "NASF.csv"
datafile.ignore.cols	The number of columns in the experimental file given in data.file.name that should not be used as numerical, for instance 2 if there is an ID field and a Description field. By default 1.
filesPath	If the zip file is not provided, the path to the GO files
node	NULL, ignored at the moment
aggregateFun	Either "sum" or "product"; the aggregation operation for abundance data
logAb	TRUE or FALSE; the abundance data to be logged or not
...	Parameters to pass

**Details**

Process all the GO files provided, in the directory or the zip, and assign ID's to the respective categories. The categories can come from a target list, or from a choice of level and ontology. If a reference is provided, then the numbers of identifiers in each category is compared to the reference by means of Fisher's exact test. If a data file is provided, then the quantitative values are aggregated into the sets.

**Value**

A list object, with the following values:

Counts	The GO counts matrix summarized for all files
Percentages	The GO percentages matrix summarized for all files
Abundance	The GO percentages matrix summarized for all files, ONLY generated if an abundance file was provided
aggregatedAbundance	The aggregated abundance matrix if the data file is provided
FisherPval	The Fisher p-values matrix summarized for all files, ONLY generated if a reference file was provided
res.list	The full list result of processAnnotation function
list.levelplots	The abundance levelplots if the data.file.name is not "none"
list.barplots	The abundance barcharts if the data.file.name is not "none"

**Author(s)**

D. Pascovici

**See Also**

[processAnnotation](#)

**Examples**

```
# where sample files are stored
path <- system.file("files", package = "PloGO2")
# run PloGO with list of id's, data file and reference
res <- PloGO( zipFile=paste(path, "GOfiles.zip", sep="/"),
reference="Control", termFile = paste(path, "GOListDrought.txt", sep="/"),
data.file.name = paste(path, "NSAFDesc.csv", sep="/"),
datafile.ignore.cols = 2)
```

---

PloPathway	<i>Function to do steps of pathway annotation and annotation/abundance plots</i>
------------	--

---

### Description

Summarize pathway categories for all the files in the zip, if provided merge data from data file, generate annotation and abundance plots and comparison with reference.

### Usage

```
PloPathway(zipFile = "none", reference = "none", data.file.name = "none", datafile.ignore.cols = 1, fil
```

### Arguments

zipFile	Zip containing all the GO files
reference	The file name of the reference file, if any, for instance "Control" for Control.txt
data.file.name	The file containing all the experimental data, for instance "NASF.csv"
datafile.ignore.cols	The number of columns in the experimental file given in data.file.name that should not be used as numerical, for instance 2 if there is an ID field and a Description field. By default 1.
filesPath	If the zip file is not provided, the path to the GO files
aggregateFun	Either "sum" or "product"; the aggregation operation for abundance data
logAb	TRUE or FALSE; the abundance data to be logged or not
...	Parameters to pass

### Details

Process all the pathway files provided, in the directory or the zip, and assign ID's to the respective categories. If a reference is provided, then the numbers of identifiers in each category is compared to the reference by means of Fisher's exact test. If a data file is provided, then the quantitative values are aggregated into the sets.

### Value

A list object, with the following values:

Counts	The GO counts matrix summarized for all files
Percentages	The GO percentages matrix summarized for all files
Abundance	The GO percentages matrix summarized for all files, ONLY generated if an abundance file was provided
aggregatedAbundance	The aggregated abundance matrix if the data file is provided

FisherPval	The Fisher p-values matrix summarized for all files, ONLY generated if a reference file was provided
res.list	The full list result of processAnnotation function
list.levelplots	The abundance levelplots if the data.file.name is not "none"
list.barplots	The abundance barcharts if the data.file.name is not "none"

**Author(s)**

J. Wu

**See Also**[processAnnotation](#)**Examples**

```
# where sample files are stored
path <- system.file("files", package = "PloG02")
# run PloPathway with list of id's, data file and reference
res <- PloPathway( zipFile=paste(path, "PWFiles.zip", sep="/"),
reference="Alldata",
data.file.name = paste(path, "Abundance_data.csv", sep="/"),
datafile.ignore.cols = 1)
```

---

plotAbundanceBar	<i>Function to plot abundance barplot</i>
------------------	---

---

**Description**

Plot the aggregated abundance barplot for all files/clusters

**Usage**

```
plotAbundanceBar(mat.abundance, mat.counts, min.count=5)
```

**Arguments**

mat.abundance	A matrix of abundance
mat.counts	A matrix of the counts
min.count	The cutoff for the minimum counts to be included

**Value**

None returned, generate a barplot.

**Author(s)**

J.Wu

**Examples**

```
path <- system.file("files", package = "PloGO2")
# run PloPathway with list of id's, data file and reference
res <- PloPathway( zipFile=paste(path, "PWFiles.zip", sep="/"),
reference="Alldata",
data.file.name = paste(path, "Abundance_data.csv", sep="/"),
datafile.ignore.cols = 1)

png("AbundanceBarplot.png", 2500, 2000, res=300)
par(mar=c(4,10,4,14))

plot.res <- plotAbundanceBar(res$aggregatedAbundance, res$Counts)

dev.off()
```

---

printSummary

*Function to print the summary file of PloGO2 results.*

---

**Description**

Each tab in the summary spreadsheet of the

**Usage**

```
printSummary(results, file="PloGO2Results.xlsx")
```

**Arguments**

results	A list of results from PloGO2 analysis
file	The output file name

**Value**

TRUE if at least some annotations were found, FALSE otherwise.

**Author(s)**

J. Wu

## Examples

```
## Not run:
path <- system.file("files", package = "PloG02")
# run PloPathway with list of id's, data file and reference
res <- PloPathway( zipFile=paste(path, "PWFiles.zip", sep="/"),
reference="Alldata",
data.file.name = paste(path, "Abundance_data.csv", sep="/"),
datafile.ignore.cols = 1)

printSummary(res)

## End(Not run)
```

---

processAnnotation	<i>Function to process a set of annotation files given a list of GO/pathway identifiers of interest</i>
-------------------	---

---

## Description

For each file in the list extract all identifiers that belong to each GO or pathway category in the list of identifiers. An identifier “belongs” to a GO or pathway category if it is annotated at the category itself or any of its children in the GO graph.

## Usage

```
processAnnotation(file.list, AnnotIDlist, data.file.name = NULL, printFiles = FALSE,
format = c("compact", "long"), datafile.ignore.cols = 1, aggregateFun="sum")
```

## Arguments

file.list	A list of files
AnnotIDlist	A list of GO/pathway identifiers
data.file.name	A list containing additional data such as abundance information
printFiles	TRUE/FALSE If true an annotation summary file is generated for each input file
format	One of “compact” by default or “long”, the format of the annotation files. See details.
datafile.ignore.cols	How many columns in the abundance file to ignore. By default assume the first only, containing identifiers.
aggregateFun	Either "sum" or "product"; the aggregation operation for abundance data

**Details**

The format is for the GO/pathway files is “compact” by default, meaning a text file containing ID’s in the first column, and GO/pathway identifiers in the second, separated by spaces or semicolons. This is the same as the “Wego native format”. A “long” format is also accepted, meaning a text file with two or more columns separated by spaces, containing an identifier, followed by a GO/pathway id, followed optionally by other columns which are ignored. The GO/pathway id’s will first be aggregated for each identifier.

**Value**

A list of the same length as the list of files submitted. Each list element is the result of [processGoFile](#) or [processPathFile](#).

**Author(s)**

D. Pascovici, J.Wu

**See Also**

[processGoFile](#), [processPathFile](#)

**Examples**

```
# For GO analysis
# choose two simple GO categories
termList <- c("response to stimulus", "transport")
GOIDmap <- getGoID(termList)
GOIDlist <- names(GOIDmap)

path <- system.file("files", package = "PloGO2")
file.names <- file.path(path, c("00100.txt", "01111.txt", "10000.txt",
"11111.txt", "Control.txt"))

# summarize annotation
res.list <- processAnnotation(file.names, GOIDlist)

# files in long format
longFormat <- file.path(path, "goRetOutput.txt")
processAnnotation(longFormat, GOIDlist, format = "long")

# For KEGG pathway analysis
fname <- file.path(path, "PWFiles", "red.txt")
datafile <- file.path(path, "Abundance_data.csv")

AnnotIDlist <- c("osa01100", "osa01110", "osa01230", "osa00300", "osa00860")

res.list <- processAnnotation(fname, AnnotIDlist, data.file.name = datafile)
```

---

processGoFile	<i>Function to process a single file in wego-like native format</i>
---------------	---

---

**Description**

For each GO category of interest extract all ID's from the file that are annotated at either the category or its GO children. If abundance data is present extract and merge.

**Usage**

```
processGoFile(fname, GOIDlist, datafile = NULL, datafile.ignore.cols = 1, format = c("compact", "long"))
```

**Arguments**

fname	The GO file name, in either Wego native format or long format
GOIDlist	The list of GO id's of interest
datafile	The file containing abundance or NULL if none.
datafile.ignore.cols	How many columns in the abundance file to ignore. By default assume the first only, containing identifiers.
format	Either "compact" or "long"; see details
aggregateFun	Either "sum" or "product"; the aggregation operation for abundance data

**Details**

The format is "compact" by default, meaning a text file containing ID's in the first column, and GO identifiers in the second, separated by spaces or semicolons. This is the same as the "Wego native format". A "long" format is also accepted, meaning a text file with two or more columns separated by spaces, containing an identifier, followed by a GO id, followed optionally by other columns which are ignored. The GO id's will first be aggregated for each identifier. The output of GOretriever can be used as "long" format.

**Value**

A list with the following components

counts	A vector of the same length as the list of GO id's of interest giving the number of ID's in each category
ID.list	The list of ID's for each GO category
datafile	The abundance datafile provided passed through
abundance	A matrix with as many rows as the GO list provided, and as many columns as the abundance data file
N	The number of protein (gene etc) identifiers in each file
fname	The filename without the file path

**Author(s)**

D. Pascovici

**Examples**

```

termList <- c("response to stimulus", "transport")
GOIDmap <- getGoID(termList)
GOIDlist <- names(GOIDmap)

# use one of the stored files
dir <- system.file("files", package="PloG02")
fname <- paste(dir, "00100.txt", sep="/")
datafile <- paste(dir, "NSAF.csv", sep="/")

# or if abundance is present aggregate that by category
processGoFile(fname, GOIDlist, datafile=datafile)

```

---

processPathFile	<i>Function to process a single pathway file in wego-like native format</i>
-----------------	---

---

**Description**

For each pathway extract all ID's from the file. If abundance data is present extract and merge.

**Usage**

```

processPathFile(fname, AnnotIDlist, datafile=NULL, datafile.ignore.cols=1,
format=c("compact", "long"), aggregateFun="sum")

```

**Arguments**

fname	The pathway file name, in either Wego native format or long format
AnnotIDlist	The list of pathway annotation ID
datafile	The file containing abundance or NULL if none.
datafile.ignore.cols	How many columns in the abundance file to ignore. By default assume the first only, containing identifiers.
format	Either "compact" or "long"; see details
aggregateFun	The aggregation function for abundance data

**Details**

The format is "compact" by default, meaning a text file containing ID's in the first column, and pathway identifiers in the second, separated by spaces or semicolons. This is the same as the "Wego native format". A "long" format is also accepted, meaning a text file with two or more columns separated by spaces, containing an identifier, followed by a pathway id, followed optionally by other columns which are ignored. The pathway id's will first be aggregated for each identifier.

**Value**

A list with the following components

counts	A vector of the same length as the list of pathway id's of interest giving the number of ID's in each category
ID.list	The list of ID's for each pathway category
datafile	The abundance datafile provided passed through
abundance	A matrix with as many rows as the pathway list provided, and as many columns as the abundance data file
N	The number of protein (gene etc) identifiers in each file
fname	The filename without the file path

**Author(s)**

J. Wu

**Examples**

```
# use one of the stored files
dir <- system.file("files", package="PloG02")
fname <- paste(dir, "PWFiles/AllData.txt", sep="/")
datafile <- paste(dir, "Abundance_data.csv", sep="/")
AnnotIDlist <- unique(unlist(sapply(read.delim(fname, stringsAsFactors=FALSE)[,2], function(x) strsplit(x, split

# or if abundance is present aggregate that by category
processPathFile(fname, AnnotIDlist, datafile=datafile)
```

---

read.annot.file      *Function to read an annotation file.*

---

**Description**

Accepts GO id's separated by space or semicolon

**Usage**

```
read.annot.file(fname, format = c("compact", "long"))
```

**Arguments**

fname            The file name containing GO annotation  
format            Either "compact" or "long"; see details

**Details**

The format is "compact" by default, meaning a text file containing ID's in the first column, and GO identifiers in the second, separated by spaces or semicolons. This is the same as the "Wego native format". A "long" format is also accepted, meaning a text file with two or more columns separated by tabs, containing an identifier, followed by a GO id, followed optionally by other columns which are ignored. The GO id's will first be aggregated for each identifier. GO files in long format can be obtained using for instance biomart, or GoRetriever.

**Value**

A data frame with two columns, ID's and GO separated by spaces

**Author(s)**

T. Keighley, D.Pascovici

**Examples**

```
# use one of the stored files
dir <- system.file("files", package="PloG02")
fname <- paste(dir,"00100.txt", sep="/")

# Example with GoRetriever download
longFormat <- paste(dir,"goRetOutput.txt", sep="/")
read.annot.file(fname)
read.annot.file(longFormat, format="long")

# Example with biomart download
biomartDownload <- paste(dir,"mart_export.txt", sep="/")
read.annot.file(biomartDownload, format="long")
```

---

writeAnnotation

*Function to print GO/pathway annotation to files*

---

**Description**

Prints available GO or pathway annotation and abundance (if existing) in a long format or an adjacency matrix type format.

**Usage**

```
writeAnnotation(res.list, datafile = NULL, datafile.ignore.cols = 1, format = c("list", "matrix"), outFolder)
```

**Arguments**

res.list	The result of processAnnotation
datafile	A CSV file with additional experimental information, if any
datafile.ignore.cols	The number of columns to ignore in the data file
format	Either "matrix" or "list".
outFolder	The output files folder

**Details**

The GO/pathway information and abundance will be printed to files. If the format is "list", then the files will be text files, and each category will be printed in turn, with all the identifiers and data underneath. If the format is "matrix", then the data will be printed in matrix format, identifiers (rows) by GO categories (columns), with the abundance data appended.

**Value**

The path of the annotation folder.

**Author(s)**

D. Pascovici

**See Also**

[processAnnotation](#)

**Examples**

```
# choose two simple GO categories
termList <- c("response to stimulus", "transport", "signaling")
GOIDmap <- getGoID(termList)
GOIDlist <- names(GOIDmap)

dir <- system.file("files", package="PloG02")
file.names <- paste(dir, c("00100.txt", "01111.txt", "10000.txt",
"11111.txt", "Control.txt"), sep="/")

# summarize annotation
res.list <- processAnnotation(file.names, GOIDlist)

# write to "matrix" or alternatively "list" format
writeAnnotation(res.list, format="matrix")
```

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