

# Package ‘POMA’

May 16, 2024

**Title** Tools for Omics Data Analysis

**Version** 1.14.0

**Description** The POMA package offers a comprehensive toolkit designed for omics data analysis, streamlining the process from initial visualization to final statistical analysis. Its primary goal is to simplify and unify the various steps involved in omics data processing, making it more accessible and manageable within a single, intuitive R package. Emphasizing on reproducibility and user-friendliness, POMA leverages the standardized SummarizedExperiment class from Bioconductor, ensuring seamless integration and compatibility with a wide array of Bioconductor tools. This approach guarantees maximum flexibility and replicability, making POMA an essential asset for researchers handling omics datasets. See <https://github.com/pcastellanoescuder/POMAShiny>. Paper: Castellano-Escuder et al. (2021) <[doi:10.1371/journal.pcbi.1009148](https://doi.org/10.1371/journal.pcbi.1009148)> for more details.

**License** GPL-3

**Encoding** UTF-8

**LazyData** true

**biocViews** BatchEffect, Classification, Clustering, DecisionTree, DimensionReduction, MultidimensionalScaling, Normalization, Preprocessing, PrincipalComponent, Regression, RNASeq, Software, StatisticalMethod, Visualization

**Imports** broom, caret, ComplexHeatmap, dbscan, dplyr, DESeq2, FSA, ggplot2, ggrepel, glmnet, impute, janitor, limma, lme4, magrittr, MASS, mixOmics, randomForest, RankProd (>= 3.14), rlang, SummarizedExperiment, sva, tibble, tidyr, utils, uwot, vegan

**Suggests** BiocStyle, covr, ggraph, ggtext, knitr, patchwork, plotly, tidyverse, testthat (>= 2.3.2)

**Roxygen** list(markdown = TRUE)

**RoxygenNote** 7.2.3

**Depends** R (>= 4.0)

**VignetteBuilder** knitr

**URL** <https://github.com/pcastellanoescuder/POMA>

**BugReports** <https://github.com/pcastellanoescuder/POMA/issues>

**git\_url** <https://git.bioconductor.org/packages/POMA>

**git\_branch** RELEASE\_3\_19

**git\_last\_commit** 1a5a6e4

**git\_last\_commit\_date** 2024-04-30

**Repository** Bioconductor 3.19

**Date/Publication** 2024-05-15

**Author** Pol Castellano-Escuder [aut, cre]  
(<https://orcid.org/0000-0001-6466-877X>)

**Maintainer** Pol Castellano-Escuder <polcaes@gmail.com>

## Contents

box_cox_transformation . . . . .	3
cor_pmat . . . . .	3
detect_decimals . . . . .	4
flattenCorrMatrix . . . . .	4
PomaBatch . . . . .	4
PomaBoxplots . . . . .	5
PomaClust . . . . .	7
PomaCorr . . . . .	8
PomaCreateObject . . . . .	9
PomaDensity . . . . .	10
PomaDESeq . . . . .	11
PomaHeatmap . . . . .	12
PomaImpute . . . . .	13
PomaLasso . . . . .	14
PomaLimma . . . . .	15
PomaLM . . . . .	16
PomaLMM . . . . .	17
PomaNorm . . . . .	19
PomaOddsRatio . . . . .	20
PomaOutliers . . . . .	21
PomaPCA . . . . .	22
PomaPCR . . . . .	23
PomaPLS . . . . .	24
PomaRandForest . . . . .	26
PomaRankProd . . . . .	27
PomaUMAP . . . . .	28
PomaUnivariate . . . . .	30
PomaVolcano . . . . .	31
poma_pal_c . . . . .	33
poma_pal_d . . . . .	33
quantile_norm . . . . .	33
scale_color_poma_c . . . . .	34

<i>box_cox_transformation</i>	3
scale_color_poma_d . . . . .	34
scale_fill_poma_c . . . . .	34
scale_fill_poma_d . . . . .	34
st000284 . . . . .	35
st000336 . . . . .	36
sum_norm . . . . .	36
theme_poma . . . . .	37
%>% . . . . .	38
<b>Index</b>	<b>39</b>

---

*box\_cox\_transformation*  
*Box-Cox Transformation*

---

**Description**

Compute Box-Cox normalization.

**Usage**

`box_cox_transformation(data)`

**Arguments**

`data`            A single variable.

---

*cor\_pmat*            *Correlation P-Values*

---

**Description**

Compute correlation p-values.

**Usage**

`cor_pmat(x, method)`

**Arguments**

`x`                A data matrix.

`method`          Character indicating which correlation coefficient has to be computed. Options are "pearson" (default), "kendall" and "spearman".

---

detect_decimals	<i>Detect decimals</i>
-----------------	------------------------

---

**Description**

Detect decimal variables.

**Usage**

```
detect_decimals(data)
```

**Arguments**

data	A data matrix (samples in rows).
------	----------------------------------

---

flattenCorrMatrix	<i>Flatten Correlation Matrix</i>
-------------------	-----------------------------------

---

**Description**

Flatten Correlation Matrix

**Usage**

```
flattenCorrMatrix(cormat, pmat)
```

**Arguments**

cormat	Output from cor.
pmat	Output from cor_pmat.

---

PomaBatch	<i>Batch Correction</i>
-----------	-------------------------

---

**Description**

PomaBatch performs batch correction on a SummarizedExperiment object given a batch factor variable.

**Usage**

```
PomaBatch(data, batch, mod = NULL)
```

**Arguments**

data	A SummarizedExperiment object.
batch	Character. The name of the column in colData that contains the batch information.
mod	Character vector. Indicates the names of colData columns to be included as covariates. Default is NULL (no covariates).

**Value**

A SummarizedExperiment object with batch-corrected data.

**Author(s)**

Pol Castellano-Escuder

**References**

Leek JT, Johnson WE, Parker HS, Fertig EJ, Jaffe AE, Zhang Y, Storey JD, Torres LC (2023). sva: Surrogate Variable Analysis. doi:10.18129/B9.bioc.sva <https://doi.org/10.18129/B9.bioc.sva>

**Examples**

```
data("st000284")

st000284 %>%
  PomaImpute(method = "knn") %>%
  PomaBatch(batch = "gender")
```

---

PomaBoxplots

*Boxplots and Violin Plots*

---

**Description**

PomaBoxplots generates boxplots and violin plots for samples and features. This function can be used for data exploration (e.g., comparison between pre and post normalized datasets).

**Usage**

```
PomaBoxplots(
  data,
  x = "samples",
  violin = FALSE,
  feature_name = NULL,
  theme_params = list(legend_title = FALSE, axis_x_rotate = TRUE)
)
```

**Arguments**

data	A SummarizedExperiment object.
x	Character. Options are "samples" (to visualize sample boxplots) and "features" (to visualize feature boxplots). Default is "samples".
violin	Logical. Indicates if violin plots should be displayed instead of boxplots. Default is FALSE.
feature_name	Character vector. Indicates the feature/s to display. Default is NULL (all features will be displayed).
theme_params	List. Indicates theme_poma parameters.

**Value**

A ggplot object.

**Author(s)**

Pol Castellano-Escuder

**Examples**

```
data("st000284")

# Sample boxplots
st000284 %>%
PomaNorm() %>%
PomaBoxplots(theme_params = list(axistext = "y"))

# Sample violin plots
st000284 %>%
PomaNorm() %>%
PomaBoxplots(violin = TRUE, theme_params = list(axistext = "y"))

# All feature boxplots
st000284 %>%
PomaNorm() %>%
PomaBoxplots(x = "features", theme_params = list(axis_x_rotate = TRUE))

# Specific feature boxplots
st000284 %>%
PomaNorm() %>%
PomaBoxplots(x = "features",
              feature_name = c("ornithine", "orotate"))

# Specific feature violin plots
st000284 %>%
PomaNorm() %>%
PomaBoxplots(x = "features",
              violin = TRUE,
              feature_name = c("ornithine", "orotate"))
```

**Description**

PomaClust performs a k-means clustering and plots the results in a classical multidimensional scaling (MDS) plot.

**Usage**

```
PomaClust(  
  data,  
  method = "euclidean",  
  k = NA,  
  k_max = floor(min(dim(data))/2),  
  show_clusters = TRUE,  
  labels = FALSE  
)
```

**Arguments**

data	A SummarizedExperiment object.
method	Character. Indicates the distance method to perform MDS. Options are "euclidean", "maximum", "manhattan", "canberra" and "minkowski". See ?dist().
k	Numeric. Indicates the number of clusters (default is NA). The optimal number of clusters will be used by default.
k_max	Numeric. Indicates the number of clusters among which the optimal k will be selected.
show_clusters	Logical. Indicates if clusters should be plotted or not.
labels	Logical. Indicates if sample names should be plotted or not.

**Value**

A list with results including plots and tables.

**Author(s)**

Pol Castellano-Escuder

**Examples**

```
data("st000284")  
  
PomaClust(st000284)
```

---

PomaCorr

*Correlation Analysis*

---

## Description

PomaCorr computes all pairwise correlations in the data.

## Usage

```
PomaCorr(data, method = "pearson", label_size = 8, theme_params = list())
```

## Arguments

data	A SummarizedExperiment object.
method	Character. Indicates which correlation coefficient has to be computed. Options are "pearson" (default), "kendall" and "spearman".
label_size	Numeric. Indicates plot label size.
theme_params	List. Indicating theme_poma parameters.

## Value

A list with the results.

## Author(s)

Pol Castellano-Escuder

## References

Jerome Friedman, Trevor Hastie and Rob Tibshirani (2019). *glasso: Graphical Lasso: Estimation of Gaussian Graphical Models*. R package version 1.11. <https://CRAN.R-project.org/package=glasso>

## Examples

```
data("st000284")

# Pearson correlation
PomaCorr(st000284)$correlations

## Gaussian graphical model
# library(ggraph)
# PomaCorr(st000284, corr_type = "glasso")
```



---

PomaCreateObject      *Create a SummarizedExperiment Object*

---

## Description

PomaCreateObject creates a SummarizedExperiment object from data frames.

## Usage

```
PomaCreateObject(metadata = NULL, features = NULL, factor_levels = 10)
```

## Arguments

metadata	Metadata variables structured in columns. Sample ID must be the first column.
features	Matrix of features. Each feature is a column.
factor_levels	Numeric. Integer variables with more levels than indicated by this parameter will be treated as factors.

## Value

A SummarizedExperiment object.

## Author(s)

Pol Castellano-Escuder

## References

Morgan M, Obenchain V, Hester J, Pagès H (2021). SummarizedExperiment: SummarizedExperiment container. R package version 1.24.0, <https://bioconductor.org/packages/SummarizedExperiment>.

## Examples

```
data(iris)

# Create metadata: Data frame with sample names and a group factor
metadata <- data.frame(ID = 1:150, Group = iris$Species)

# Create features: `p` column data frame with features
features <- iris[, 1:4]

# Create a `SummarizedExperiment` object with `POMA`
object <- PomaCreateObject(metadata = metadata, features = features)
```

PomaDensity

*Density Plots***Description**

PomaDensity generates a density plot for samples and features. This function can be used for data exploration (e.g., comparison between pre and post normalized datasets).

**Usage**

```
PomaDensity(
  data,
  x = "samples",
  feature_name = NULL,
  theme_params = list(legend_title = FALSE)
)
```

**Arguments**

data	A SummarizedExperiment object.
x	Character. Options are "samples" (to visualize sample density plots) and "features" (to visualize feature density plots). Default is "samples".
feature_name	Character vector. Indicates the feature/s to display. Default is NULL (all features will be displayed).
theme_params	List. Indicates theme_poma parameters.

**Value**

A ggplot object.

**Author(s)**

Pol Castellano-Escuder

**Examples**

```
data("st000284")

# Sample density plots
st000284 %>%
PomaNorm() %>%
PomaDensity(theme_params = list(axistext = "y"))

# All feature density plots
st000284 %>%
PomaNorm() %>%
PomaDensity(x = "features", theme_params = list(legend_position = "none"))
```

```
# Specific feature density plots
st000284 %>%
PomaNorm() %>%
PomaDensity(x = "features",
            feature_name = c("ornithine", "orotate"))
```

---

PomaDESeq	<i>Differential Expression Analysis Based on the Negative Binomial Distribution</i>
-----------	---

---

## Description

PomaDESeq estimates variance-mean dependence in count data from high-throughput sequencing assays and test for differential expression based on a model using the negative binomial distribution.

## Usage

```
PomaDESeq(data, adjust = "fdr")
```

## Arguments

data	A SummarizedExperiment object.
adjust	Character. Multiple comparisons correction method to adjust p-values. Available options are: "fdr" (false discovery rate), "holm", "hochberg", "hommel", "bonferroni", "BH" (Benjamini-Hochberg), and "BY" (Benjamini-Yekutieli).

## Value

A tibble with the results.

## Author(s)

Pol Castellano-Escuder

## References

Love, M.I., Huber, W., Anders, S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2 Genome Biology 15(12):550 (2014)

---

PomaHeatmap

*Heatmap Plot*

---

## Description

PomaHeatmap generates a heatmap.

## Usage

```
PomaHeatmap(  
  data,  
  covs = NULL,  
  sample_names = TRUE,  
  feature_names = FALSE,  
  show_legend = TRUE  
)
```

## Arguments

data	A SummarizedExperiment object.
covs	Character vector. Indicates the names of colData columns to be included as covariates. Default is NULL (no covariates).
sample_names	Logical. Indicates if sample names should be displayed or not. Default is TRUE.
feature_names	Logical. Indicates if feature names should be displayed or not. Default is FALSE.
show_legend	Logical. Indicates if legend should be displayed or not. Default is TRUE.

## Value

A heatmap plot.

## Author(s)

Pol Castellano-Escuder

## Examples

```
data("st000284")  
  
# Basic heatmap  
st000284 %>%  
  PomaNorm() %>%  
  PomaHeatmap()  
  
# Heatmap with one covariate  
st000284 %>%  
  PomaNorm() %>%
```

```

PomaHeatmap(covs = "factors")

# Heatmap with two covariates
st000284 %>%
  PomaNorm() %>%
  PomaHeatmap(covs = c("factors", "smoking_condition"))

```

---

PomaImpute

*Impute Missing Values*


---

## Description

PomaImpute performs missing value imputation on a dataset using various imputation methods.

## Usage

```

PomaImpute(
  data,
  zeros_as_na = FALSE,
  remove_na = TRUE,
  cutoff = 20,
  group_by = TRUE,
  method = "knn"
)

```

## Arguments

data	A SummarizedExperiment object.
zeros_as_na	Logical. Indicates if the zeros in the data are missing values. Default is FALSE.
remove_na	Logical. Indicates if features with a percentage of missing values over the cutoff parameter should be removed. Default is TRUE.
cutoff	Numeric. Percentage of missing values allowed in each feature.
group_by	Logical. If metadata file is present and its first variable is a factor, it can be used to compute missing values per group and drop them accordingly. Features will be removed only if all of the groups contain more missing values than allowed. Default is TRUE.
method	Character. The imputation method to use. Options include "none" (no imputation, replace missing values by zeros), "half_min" (replace missing values with half of the minimum value), "median" (replace missing values with the median), "mean" (replace missing values with the mean), "min" (replace missing values with the minimum value), "knn" (replace missing values using k-nearest neighbors imputation), and "random_forest" (replace missing values using random forest imputation).

## Value

A SummarizedExperiment object without missing values.

**Author(s)**

Pol Castellano-Escuder

**References**

Armitage, E. G., Godzien, J., Alonso-Herranz, V., López-González, Á., & Barbas, C. (2015). Missing value imputation strategies for metabolomics data. *Electrophoresis*, 36(24), 3050-3060.

**Examples**

```
data("st000336")

PomaImpute(st000336, method = "knn")
```

---

PomaLasso	<i>Lasso, Ridge, and Elasticnet Regularized Generalized Linear Models for Binary Outcomes</i>
-----------	---

---

**Description**

PomaLasso performs LASSO, Ridge, and Elasticnet regression for feature selection and prediction purposes for binary outcomes.

**Usage**

```
PomaLasso(
  data,
  alpha = 1,
  ntest = NULL,
  nfolds = 10,
  lambda = NULL,
  labels = FALSE
)
```

**Arguments**

data	A SummarizedExperiment object.
alpha	Numeric. Indicates the elasticnet mixing parameter. $\alpha = 1$ is the LASSO penalty and $\alpha = 0$ is the Ridge penalty.
ntest	Numeric. Indicates the percentage of observations that will be used as test set. Default is NULL (no test set).
nfolds	Numeric. Indicates number of folds for cross-validation (default is 10). Although nfolds can be as large as the sample size (leave-one-out CV), it is not recommended for large datasets. Smallest value allowable is $\text{nfolds} = 3$ .
lambda	Numeric. Indicates the user supplied lambda sequence. Typical usage is to have the program compute its own lambda sequence based on <code>nlambda</code> and <code>lambda.min.ratio</code> . See <code>?glmnet::glmnet()</code> .

labels Logical. Indicates if feature names should be plotted in coefficient plot or not. Default is FALSE.

**Value**

A list with results.

**Author(s)**

Pol Castellano-Escuder

**References**

Jerome Friedman, Trevor Hastie, Robert Tibshirani (2010). Regularization Paths for Generalized Linear Models via Coordinate Descent. *Journal of Statistical Software*, 33(1), 1-22. URL <http://www.jstatsoft.org/v33/i01/>.

**Examples**

```
data("st000336")

# lasso
st000336 %>%
  PomaImpute() %>%
  PomaNorm() %>%
  PomaLasso()

# elasticnet
st000336 %>%
  PomaImpute() %>%
  PomaNorm() %>%
  PomaLasso(alpha = 0.5)

# ridge
st000336 %>%
  PomaImpute() %>%
  PomaNorm() %>%
  PomaLasso(alpha = 0)
```

**Description**

PomaLimma uses the classical limma package to compute differential expression analysis.

**Usage**

```
PomaLimma(data, contrast = NULL, covs = NULL, adjust = "fdr", weights = FALSE)
```

**Arguments**

data	A SummarizedExperiment object.
contrast	Character. Indicates the comparison. For example, "Group1-Group2" or "control-intervention".
covs	Character vector. Indicates the names of colData columns to be included as covariates. Default is NULL (no covariates). If not NULL, a limma model will be fitted using the specified covariates. Note: The order of the covariates is important and should be listed in increasing order of importance in the experimental design.
adjust	Character. Indicates the multiple comparisons correction method. Options are: "fdr", "holm", "hochberg", "hommel", "bonferroni", "BH" and "BY".
weights	Logical. Indicates whether the limma model should estimate the relative quality weights for each group. See ?limma::arrayWeights().

**Value**

A tibble with the results.

**Author(s)**

Pol Castellano-Escuder

**References**

Matthew E. Ritchie, Belinda Phipson, Di Wu, Yifang Hu, Charity W. Law, Wei Shi, Gordon K. Smyth, limma powers differential expression analyses for RNA-sequencing and microarray studies, *Nucleic Acids Research*, Volume 43, Issue 7, 20 April 2015, Page e47, <https://doi.org/10.1093/nar/gkv007>

**Examples**

```
data("st000284")

st000284 %>%
  PomaNorm() %>%
  PomaLimma(contrast = "Healthy-CRC", adjust = "fdr")
```

---

PomaLM

*Linear Models*

---

**Description**

PomaLM performs a linear model on a SummarizedExperiment object.

**Usage**

```
PomaLM(data, x = NULL, y = NULL, adjust = "fdr")
```



**Arguments**

data	A SummarizedExperiment object.
x	Character vector. Indicates the names of independent variables. If it's NULL (default), all features will be used.
y	Character. Indicates the name of colData numeric columns to be used as dependent variable. If it's set to NULL, the first numeric variable in colData will be used as the dependent variable.
adjust	Character. Multiple comparisons correction method to adjust p-values. Available options are: "fdr" (false discovery rate), "holm", "hochberg", "hommel", "bonferroni", "BH" (Benjamini-Hochberg), and "BY" (Benjamini-Yekutieli).

**Value**

A list with results including plots and tables.

**Author(s)**

Pol Castellano-Escuder

**Examples**

```
data("st000284")

# Perform linear model with all features
st000284 %>%
PomaLM()

# Perform linear model with two features
st000284 %>%
PomaLM(x = c("x1_methyladenosine", "x2_deoxyuridine"))
```

---

PomaLMM

*Linear Mixed Models*

---

**Description**

PomaLMM performs linear mixed models on a SummarizedExperiment object.

**Usage**

```
PomaLMM(data, x = NULL, y = NULL, adjust = "fdr", clean_plot = FALSE)
```

**Arguments**

<code>data</code>	A SummarizedExperiment object.
<code>x</code>	Character vector. Indicates the names of <code>colData</code> columns to be used as random and fixed effects (independent variables). If it's set to <code>NULL</code> (default), all variables in <code>colData</code> will be used.
<code>y</code>	Character vector. Indicates the names of dependent variables. If it's <code>NULL</code> (default), all features will be used.
<code>adjust</code>	Character. Multiple comparisons correction method to adjust p-values. Available options are: "fdr" (false discovery rate), "holm", "hochberg", "hommel", "bonferroni", "BH" (Benjamini-Hochberg), and "BY" (Benjamini-Yekutieli).
<code>clean_plot</code>	Logical. Indicates if remove intercept and linear mixed model residues boxplots from the plot. Default is <code>FALSE</code> .

**Value**

A list with results including plots and tables. Table values indicate the percentage variance explained per variable.

**Author(s)**

Pol Castellano-Escuder

**Examples**

```
data("st000284")

# Perform linear mixed model with all features
st000284 %>%
PomaLMM()

# Perform linear mixed model with two features
st000284 %>%
PomaLMM(y = c("x1_methyladenosine", "x1_methylhistamine"))

# Perform linear mixed model with one random effect
st000284 %>%
PomaLMM(x = "smoking_condition")

# Perform linear mixed model with two random effects and two features
st000284 %>%
PomaLMM(x = c("smoking_condition", "gender"),
        y = c("x1_methyladenosine", "x1_methylhistamine"))

# Perform linear mixed model with no random effects and two features, therefore, a linear model will be fitted
st000284 %>%
PomaLMM(x = "age_at_consent", # Numerical, i.e., fixed effect
        y = c("x1_methyladenosine", "x1_methylhistamine"))

# Perform linear mixed model with no random effects and all features, therefore, a linear model will be fitted
st000284 %>%
```

```
PomaLMM(x = "age_at_consent") # Numerical i.e., fixed effect
```

---

PomaNorm

*Normalize Data*

---

## Description

PomaNorm performs data normalization using various normalization methods.

## Usage

```
PomaNorm(data, sample_norm = "none", method = "log_pareto")
```

## Arguments

data	A SummarizedExperiment object.
sample_norm	Character. Sample normalization method. Options include "none" (default), "sum", or "quantile".
method	Character. The normalization method to use. Options include "none" (no normalization), "auto_scaling" (autoscaling normalization, i.e., Z-score normalization), "level_scaling" (level scaling normalization), "log_scaling" (log scaling normalization), "log_transform" (log transformation normalization), "vast_scaling" (vast scaling normalization), "log_pareto" (log Pareto scaling normalization), "min_max" (min-max normalization), and "box_cox" (Box-Cox transformation).

## Value

A SummarizedExperiment object with normalized data.

## Author(s)

Pol Castellano-Escuder

## References

Van den Berg, R. A., Hoefsloot, H. C., Westerhuis, J. A., Smilde, A. K., & van der Werf, M. J. (2006). Centering, scaling, and transformations: improving the biological information content of metabolomics data. *BMC genomics*, 7(1), 142.

## Examples

```
data("st000284")  
  
PomaNorm(st000284, method = "log_pareto")
```

---

PomaOddsRatio      *Logistic Regression Model Odds Ratios*

---

### Description

PomaOddsRatio calculates the Odds Ratios for each feature from a logistic regression model using the binary outcome (group/type must be a binary factor) as a dependent variable.

### Usage

```
PomaOddsRatio(data, feature_name = NULL, covs = NULL, show_ci = TRUE)
```

### Arguments

data	A SummarizedExperiment object.
feature_name	Character vector. Indicates the name/s of feature/s that will be used to fit the model. If it's NULL (default), all variables will be included in the model.
covs	Character vector. Indicates the names of colData columns to be included as covariates. Default is NULL (no covariates).
show_ci	Logical. Indicates if the 95% confidence intervals will be plotted. Default is TRUE.

### Value

A list with results including plots and tables.

### Author(s)

Pol Castellano-Escuder

### Examples

```
data("st000336")

st000336 %>%
  PomaImpute() %>%
  PomaNorm() %>%
  PomaOddsRatio(feature_name = c("glutamic_acid", "glutamine", "glycine", "histidine"))
```

**Description**

PomaOutliers analyses and removes statistical outliers from the data.

**Usage**

```
PomaOutliers(  
  data,  
  method = "euclidean",  
  type = "median",  
  coef = 2,  
  labels = FALSE  
)
```

**Arguments**

data	A SummarizedExperiment object.
method	Character. Indicates the distance measure method to perform MDS.
type	Character. Indicates the type of outlier analysis to perform. Options are "median" (default) and "centroid". See <code>vegan::betadisper</code> .
coef	Numeric. Indicates the outlier coefficient. Lower values are more sensitive to outliers while higher values are less restrictive about outliers.
labels	Logical. Indicates if sample names should to be plotted.

**Value**

A list with the results.

**Author(s)**

Pol Castellano-Escuder

**Examples**

```
data("st000336")  
  
# clean outliers  
st000336 %>%  
  PomaImpute() %>%  
  PomaNorm() %>%  
  PomaOutliers()
```

---

PomaPCA

*Principal Components Analysis*

---

### Description

PomaPCA performs a principal components analysis on the given SummarizedExperiment object.

### Usage

```
PomaPCA(  
  data,  
  center = TRUE,  
  scale = TRUE,  
  ncomp = 4,  
  labels = FALSE,  
  ellipse = FALSE,  
  load_length = 1  
)
```

### Arguments

data	A SummarizedExperiment object.
center	Logical. Indicates whether the variables should be shifted to be zero centered. Default is TRUE.
scale	Logical. Indicates whether the variables should be scaled to have unit variance before the analysis takes place. Default is TRUE.
ncomp	Numeric. Number of components to be included in the results. Default is 4.
labels	Logical. Indicates if sample names should be displayed.
ellipse	Logical. Indicates whether a 95 percent confidence interval ellipse should be displayed in score plot and biplot. Default is FALSE.
load_length	Numeric. Indicates the length of biplot loading arrows. Value between 1 and 2. Default is 1.

### Value

A list with results including plots and tables.

### Author(s)

Pol Castellano-Escuder

**Examples**

```

data("st000336")

st000336 %>%
  PomaImpute() %>%
  PomaNorm() %>%
  PomaPCA()

```

---

PomaPCR	<i>Principal Components Regression</i>
---------	--

---

**Description**

PomaPCR performs Principal Components Regression.

**Usage**

```
PomaPCR(data, center = TRUE, scale = TRUE, ncomp = 2, y = NULL, adjust = "fdr")
```

**Arguments**

data	A SummarizedExperiment object.
center	Logical. Indicates whether the variables should be shifted to be zero centered. Default is TRUE.
scale	Logical. Indicates whether the variables should be scaled to have unit variance before the analysis takes place. Default is TRUE.
ncomp	Numeric. Indicates the number of principal components used as predictors in the model. Default is 2.
y	Character. Indicates the name of colData columns to be used as dependent variable. If it's set to NULL, the first numeric variable in colData will be used as the dependent variable.
adjust	Character. Multiple comparisons correction method to adjust p-values. Available options are: "fdr" (false discovery rate), "holm", "hochberg", "hommel", "bonferroni", "BH" (Benjamini-Hochberg), and "BY" (Benjamini-Yekutieli).

**Value**

A tibble with the results.

**Author(s)**

Pol Castellano-Escuder

## Examples

```
data("st000284")

# PCR with 2 components
st000284 %>%
  PomaPCR(y = "age_at_consent")

# PCR with 20 components
st000284 %>%
  PomaPCR(ncomp = 20)
```

---

PomaPLS

*Partial Least Squares Methods*

---

## Description

PomaPLS performs Partial Least Squares (PLS) regression, Partial Least Squares Discriminant Analysis (PLS-DA) to classify samples, and Sparse Partial Least Squares Discriminant Analysis (sPLS-DA) to classify samples (supervised analysis) and select variables.

## Usage

```
PomaPLS(  
  data,  
  method = "pls",  
  y = NULL,  
  ncomp = 5,  
  labels = FALSE,  
  ellipse = TRUE,  
  cross_validation = FALSE,  
  validation = "Mfold",  
  folds = 5,  
  nrepeat = 10,  
  vip = 1,  
  num_features = 10,  
  theme_params = list()  
)
```

## Arguments

<code>data</code>	A SummarizedExperiment object.
<code>method</code>	Character. PLS method. Options include "pls", "plsda", and "splda".
<code>y</code>	Character. Indicates the name of colData columns to be used as dependent variable. If it's set to NULL, the first variable in colData will be used as the dependent variable.
<code>ncomp</code>	Numeric. Number of components in the model. Default is 5.



labels	Logical. Indicates if sample names should be displayed.
ellipse	Logical. Indicates whether a 95 percent confidence interval ellipse should be displayed. Default is TRUE.
cross_validation	Logical. Indicates if cross-validation should be performed for PLS-DA ("plsda") and sPLS-DA ("splstda") methods. Default is FALSE.
validation	Character. (Only for "plsda" and "splstda" methods). Indicates the cross-validation method. Options are "Mfold" and "loo" (Leave-One-Out).
folds	Numeric. (Only for "plsda" and "splstda" methods). Number of folds for "Mfold" cross-validation method (default is 5). If the validation method is "loo", this value is set to 1.
nrepeat	Numeric. (Only for "plsda" and "splstda" methods). Number of times the cross-validation process is repeated.
vip	Numeric. (Only for "plsda" method). Indicates the variable importance in the projection (VIP) cutoff.
num_features	Numeric. (Only for "splstda" method). Number of features to discriminate groups.
theme_params	List. Indicates theme_poma parameters.

**Value**

A list with results including plots and tables.

**Author(s)**

Pol Castellano-Escuder

**Examples**

```
data("st000284")

# PLS
st000284 %>%
  PomaNorm() %>%
  PomaPLS(method = "pls")

data("st000336")

# PLSDA
st000336 %>%
  PomaImpute() %>%
  PomaNorm() %>%
  PomaPLS(method = "plsda")

# PLSDA with Cross-Validation
st000336 %>%
  PomaImpute() %>%
  PomaNorm() %>%
```

```

PomaPLS(method = "plsda", cross_validation = TRUE)

# sPLSDA
st000336 %>%
  PomaImpute() %>%
  PomaNorm() %>%
  PomaPLS(method = "splstda")

# sPLSDA with Cross-Validation
st000336 %>%
  PomaImpute() %>%
  PomaNorm() %>%
  PomaPLS(method = "splstda", ncomp = 3, cross_validation = TRUE)

```

---

PomaRandForest

*Classification Random Forest*


---

## Description

PomaRandForest performs classification random forest. This method can be used both for prediction and variable selection.

## Usage

```

PomaRandForest(
  data,
  ntest = NULL,
  ntree = 500,
  mtry = floor(sqrt(ncol(t(SummarizedExperiment::assay(data))))),
  nodesize = 1,
  nvar = 20
)

```

## Arguments

data	A SummarizedExperiment object.
ntest	Numeric. Indicates the percentage of observations that will be used as test set. Default is NULL (no test set).
ntree	Numeric. Indicates the number of trees to grow.
mtry	Numeric. Indicates the number of variables randomly sampled as candidates at each split. This value is set $\sqrt{p}$ (where $p$ is number of variables in data) by default.
nodesize	Numeric. Indicates the minimum size of terminal nodes. Default is 1.
nvar	Numeric. Indicates the number of variables to show in the Gini Index plot.

## Value

A list with results including plots and tables.

**Author(s)**

Pol Castellano-Escuder

**References**

A. Liaw and M. Wiener (2002). Classification and Regression by randomForest. R News 2(3), 18–22.

**Examples**

```
data("st000336")

st000336 %>%
  PomaImpute() %>%
  PomaRandForest()
```

---

PomaRankProd

*Rank Product/Rank Sum Analysis*

---

**Description**

PomaRankProd performs the Rank Product (or Rank Sum) method to identify differentially expressed genes.

**Usage**

```
PomaRankProd(data, logged = TRUE, paired = NA, cutoff = 0.05, method = "pfp")
```

**Arguments**

data	A SummarizedExperiment object.
logged	Logical. Indicates if data should be log transformed first.
paired	Numeric. Indicates the number of random pairs generated in the function, if set to NA (default), the odd integer closer to the square of the number of replicates is used.
cutoff	Numeric. Indicates the pfp/pvalue threshold value used to select features.
method	Character. Indicates the method to identify features. "pfp" uses percentage of false prediction, which is a default setting. "pval" uses p-values which is less stringent than pfp.

**Value**

A list with results including plots and tables.

**Author(s)**

Pol Castellano-Escuder

## References

Breitling, R., Armengaud, P., Amtmann, A., and Herzyk, P.(2004) Rank Products: A simple, yet powerful, new method to detect differentially regulated genes in replicated microarray experiments, FEBS Letter, 57383-92

Hong, F., Breitling, R., McEntee, W.C., Wittner, B.S., Nemhauser, J.L., Chory, J. (2006). RankProd: a bioconductor package for detecting differentially expressed genes in meta-analysis Bioinformatics. 22(22):2825-2827

Del Carratore, F., Jankevics, A., Eisinga, R., Heskes, T., Hong, F. & Breitling, R. (2017). RankProd 2.0: a refactored Bioconductor package for detecting differentially expressed features in molecular profiling datasets. Bioinformatics. 33(17):2774-2775

## Examples

```
data("st000336")

st000336 %>%
  PomaImpute() %>%
  PomaRankProd()
```

---

PomaUMAP

*Dimensionality Reduction with UMAP*

---

## Description

PomaUMAP performs a dimension reduction of the data using the Uniform Manifold Approximation and Projection (UMAP) method. See `?uwot::umap()` for more.

## Usage

```
PomaUMAP(
  data,
  n_neighbors = floor(sqrt(nrow(data))),
  n_components = 2,
  metric = "euclidean",
  pca = NULL,
  min_dist = 0.01,
  spread = 1,
  hdbSCAN_minpts = floor(nrow(data) * 0.05),
  show_clusters = TRUE,
  hide_noise = TRUE,
  labels = FALSE,
  theme_params = list(legend_title = TRUE, legend_position = "bottom")
)
```

**Arguments**

<code>data</code>	A SummarizedExperiment object.
<code>n_neighbors</code>	Numeric. Indicates the size of local neighborhood (sample points) used for manifold approximation.
<code>n_components</code>	Numeric. Indicates the dimension of the space to embed into.
<code>metric</code>	Character. Indicates the distance measure method to find nearest neighbors. Options are "euclidean", "cosine", "manhattan", "hamming" and "correlation". See <code>?uwot::umap()</code> .
<code>pca</code>	If not NULL (default), reduce data to this number of columns using PCA before UMAP.
<code>min_dist</code>	Numeric. Indicates the effective minimum distance between embedded points.
<code>spread</code>	Numeric. Indicates the effective scale of embedded points.
<code>hdbscan_minpts</code>	Numeric. Indicates the minimum size of clusters. See <code>?hdbscan::hdbscan()</code> .
<code>show_clusters</code>	Logical. Indicates if clusters computed with HDBSCAN method should be plotted or not.
<code>hide_noise</code>	Logical. Specifies whether to hide Cluster 0 in the plot. In HDBSCAN, Cluster 0 is typically regarded as "noise."
<code>labels</code>	Logical. Indicates if sample names should be plotted or not.
<code>theme_params</code>	List. Indicates <code>theme_poma</code> parameters.

**Value**

A list with results including plots and tables.

**Author(s)**

Pol Castellano-Escuder

**References**

McInnes, L., Healy, J., & Melville, J. (2018). Umap: Uniform manifold approximation and projection for dimension reduction. arXiv preprint arXiv:1802.03426.

Campello, R. J., Moulavi, D., & Sander, J. (2013, April). Density-based clustering based on hierarchical density estimates. In Pacific-Asia conference on knowledge discovery and data mining (pp. 160-172). Springer, Berlin, Heidelberg.

**Examples**

```
data("st000284")

st000284 %>%
  PomaNorm() %>%
  PomaUMAP()
```

**Description**

PomaUnivariate performs parametric and non-parametric univariate statistical tests on a SummarizedExperiment object to compare groups or conditions. Available methods include T-test, ANOVA, ANCOVA, Mann Whitney U Test (Wilcoxon Rank Sum Test), and Kruskal-Wallis.

**Usage**

```
PomaUnivariate(
  data,
  method = "ttest",
  covs = NULL,
  error = NULL,
  paired = FALSE,
  var_equal = FALSE,
  adjust = "fdr",
  run_post_hoc = TRUE
)
```

**Arguments**

data	A SummarizedExperiment object.
method	Character. The univariate statistical test to be performed. Available options include "ttest" (T-test), "anova" (analysis of variance), "mann" (Wilcoxon rank-sum test), and "kruskal" (Kruskal-Wallis test).
covs	Character vector. Indicates the names of colData columns to be included as covariates. Default is NULL (no covariates). If not NULL, an ANCOVA model will be fitted using the specified covariates. Note: The order of the covariates is important and should be listed in increasing order of importance in the experimental design.
error	Character vector. Indicates the name of a colData column to be included as an error term (e.g. replicates). Default is NULL (no error term).
paired	Logical. Indicates if the data is paired or not. Default is FALSE.
var_equal	Logical. Indicates if the data variances are assumed to be equal or not. Default is FALSE.
adjust	Character. Multiple comparisons correction method to adjust p-values. Available options are: "fdr" (false discovery rate), "holm", "hochberg", "hommel", "bonferroni", "BH" (Benjamini-Hochberg), and "BY" (Benjamini-Yekutieli).
run_post_hoc	Logical. Indicates if computing post-hoc tests or not. Setting this parameter to FALSE can save time for large datasets.

**Value**

A list with the results.

**Author(s)**

Pol Castellano-Escuder

**Examples**

```
data("st000336")

# Perform T-test
st000336 %>%
PomaImpute() %>%
PomaUnivariate(method = "ttest")

# Perform Mann-Whitney U test
st000336 %>%
PomaImpute() %>%
PomaUnivariate(method = "mann", adjust = "fdr")

data("st000284")
# Perform Two-Way ANOVA
st000284 %>%
PomaUnivariate(method = "anova", covs = c("gender"))

# Perform Three-Way ANOVA
st000284 %>%
PomaUnivariate(method = "anova", covs = c("gender", "smoking_condition"))

# Perform ANCOVA with one numeric covariate and one factor covariate
# st000284 %>%
# PomaUnivariate(method = "anova", covs = c("age_at_consent", "smoking_condition"))

# Perform Kruskal-Wallis test
st000284 %>%
PomaUnivariate(method = "kruskal", adjust = "holm")
```

---

PomaVolcano

*Volcano Plot*

---

**Description**

PomaVolcano creates a volcano plot from a given dataset. This function is designed to visualize the statistical significance (p-value) against the magnitude of change (log2 fold change) for features.

**Usage**

```
PomaVolcano(
  data,
  pval_cutoff = 0.05,
  log2fc_cutoff = NULL,
  labels = FALSE,
  x_label = "log2 (Fold Change)",
  y_label = "-log10 (P-value)"
)
```

**Arguments**

<code>data</code>	A data frame with at least three columns: feature names, statistical significance values, and magnitude of change values. These should be the first three columns of the data, in this exact order.
<code>pval_cutoff</code>	Numeric. Specifies the p-value threshold for significance in the volcano plot. The default is set to 0.05. This parameter determines the horizontal line in the plot indicating the threshold for statistical significance.
<code>log2fc_cutoff</code>	Numeric. Specifies the log2 fold change cutoff for the volcano plot. If not provided, the cutoff is set to the 75th percentile of the absolute log2 fold changes in the data. This parameter determines the vertical lines in the plot indicating the magnitude of change threshold.
<code>labels</code>	Logical. Indicates whether to plot labels for significant features.
<code>x_label</code>	Character. Custom label for the x-axis.
<code>y_label</code>	Character. Custom label for the y-axis.

**Value**

A ggplot object representing the volcano plot.

**Author(s)**

Pol Castellano-Escuder

**Examples**

```
st000336 %>%
  PomaImpute() %>%
  PomaUnivariate() %>%
  magrittr::extract2("result") %>%
  dplyr::select(feature, fold_change, pvalue) %>%
  PomaVolcano()
```



---

poma_pal_c	<i>Return function to interpolate a continuous POMA color palette</i>
------------	---

---

**Description**

Return function to interpolate a continuous POMA color palette

**Usage**

```
poma_pal_c(palette = "nature")
```

**Arguments**

palette	Character name of palette in poma_palettes
---------	--

---

poma_pal_d	<i>Return function to interpolate a discrete POMA color palette</i>
------------	---

---

**Description**

Return function to interpolate a discrete POMA color palette

**Usage**

```
poma_pal_d(palette = "nature")
```

**Arguments**

palette	Character name of palette in poma_palettes
---------	--

---

quantile_norm	<i>Sample Quantile Normalization</i>
---------------	--------------------------------------

---

**Description**

Compute quantile normalization.

**Usage**

```
quantile_norm(data)
```

**Arguments**

data	A data matrix (samples in rows).
------	----------------------------------

---

scale\_color\_poma\_c     *Color scale constructor for continuous viridis "plasma" palette*

---

**Description**

Color scale constructor for continuous viridis "plasma" palette

**Usage**

```
scale_color_poma_c()
```

---

scale\_color\_poma\_d     *Color scale constructor for discrete viridis "plasma" palette*

---

**Description**

Color scale constructor for discrete viridis "plasma" palette

**Usage**

```
scale_color_poma_d()
```

---

scale\_fill\_poma\_c     *Fill scale constructor for continuous viridis "plasma" palette*

---

**Description**

Fill scale constructor for continuous viridis "plasma" palette

**Usage**

```
scale_fill_poma_c()
```

---

scale\_fill\_poma\_d     *Fill scale constructor for discrete viridis "plasma" palette*

---

**Description**

Fill scale constructor for discrete viridis "plasma" palette

**Usage**

```
scale_fill_poma_d()
```

---

st000284

*Colorectal Cancer Detection Using Targeted Serum Metabolic Profiling*

---

## Description

Colorectal cancer (CRC) is one of the most prevalent and deadly cancers in the world. Despite an expanding knowledge of its molecular pathogenesis during the past two decades, robust biomarkers to enable screening, surveillance, and therapy monitoring of CRC are still lacking. In this study, we present a targeted liquid chromatography-tandem mass spectrometry-based metabolic profiling approach for identifying biomarker candidates that could enable highly sensitive and specific CRC detection using human serum samples. In this targeted approach, 158 metabolites from 25 metabolic pathways of potential significance were monitored in 234 serum samples from three groups of patients (66 CRC patients, 76 polyp patients, and 92 healthy controls). Partial least squares-discriminant analysis (PLS-DA) models were established, which proved to be powerful for distinguishing CRC patients from both healthy controls and polyp patients. Receiver operating characteristic curves generated based on these PLS-DA models showed high sensitivities (0.96 and 0.89, respectively, for differentiating CRC patients from healthy controls or polyp patients); good specificities (0.80 and 0.88), and excellent areas under the curve (0.93 and 0.95) were also obtained. Monte Carlo cross validation (MCCV) was also applied, demonstrating the robust diagnostic power of this metabolic profiling approach.

## Usage

st000284

## Format

A SummarizedExperiment object: 224 samples, 113 metabolites, 4 covariables and 3 groups (CRC, Healthy and Polyp).

**metabolites** 113 serum metabolites.

**covariables** Age at consent, Gender, Smoking Condition and Alcohol Consumption.

## Source

[https://www.metabolomicsworkbench.org/data/DRCCMetadata.php?Mode=Study&StudyID=ST000284&StudyType=MS&ResultType=1%20target=\\_blank](https://www.metabolomicsworkbench.org/data/DRCCMetadata.php?Mode=Study&StudyID=ST000284&StudyType=MS&ResultType=1%20target=_blank)

## References

Colorectal Cancer Detection Using Targeted Serum Metabolic Profiling, J. Proteome. Res., 2014, 13, 4120-4130.

---

`st000336`*Targeted LC/MS of urine from boys with DMD and controls*

---

**Description**

Duchenne Muscular Dystrophy (DMD) is an X-linked recessive form of muscular dystrophy that affects males via a mutation in the gene for the muscle protein, dystrophin. Progression of the disease results in severe muscle loss, ultimately leading to paralysis and death. Steroid therapy has been a commonly employed method for reducing the severity of symptoms. This study aims to quantify the urine levels of amino acids and organic acids in patients with DMD both with and without steroid treatment. Track the progression of DMD in patients who have provided multiple urine samples.

**Usage**`st000336`**Format**

A SummarizedExperiment object: 57 samples, 31 metabolites, 1 covariable and 2 groups (Controls and DMD).

**metabolites** 31 urine metabolites.

**covariables** Steroid status.

**Source**

<https://www.metabolomicsworkbench.org/data/DRCCMetadata.php?Mode=Study&DataMode=AllData&StudyID=ST000336&StudyType=MS&ResultType=1#DataTabs>

---

`sum_norm`*Sample Sum Normalization*

---

**Description**

Compute sum normalization. Final unit is a percentage.

**Usage**`sum_norm(data)`**Arguments**

`data` A data matrix (samples in rows).

---

theme_poma	<i>A ggplot theme which allow custom yet consistent styling of plots in the POMA package and web app.</i>
------------	---

---

### Description

A ggplot theme which allow custom yet consistent styling of plots in the POMA package and web app.

### Usage

```
theme_poma(  
  base_size = 15,  
  axistitle = "xy",  
  axistext = "xy",  
  legend_position = "bottom",  
  legend_title = TRUE,  
  axis_x_rotate = FALSE,  
  margin = 2  
)
```

### Arguments

base_size	(integer) Base point size
axistitle	(string) Axis titles. Options include "none" or any combination of "X", "Y", "x" and "y".
axistext	(string) Axis text labels for values or groups. Options include "none" or any combination of "X", "Y", "x" and "y".
legend_position	Character. Legend position. See ggplot2 documentation.
legend_title	Logical. Include legend title.
axis_x_rotate	Logical. Rotate x-axis 45 degrees.
margin	(numeric) Should a margin of x be added to the plot? Defaults to 0 (no margin by default).

### Examples

```
## Not run:  
library(ggplot2)  
ggplot(diamonds, aes(cut)) + geom_bar() + theme_poma()  
  
## End(Not run)
```

---

%>%

*Pipe operator*

---

**Description**

See `magrittr::%>%` for details.

**Usage**

lhs %>% rhs

**Value**

Nothing. Just allow the use of magrittr pipe "%>%"

# Index

- \* **datasets**
  - st000284, [35](#)
  - st000336, [36](#)
- \* **internal**
  - %>%, [38](#)
- %>%, [38](#), [38](#)
- box\_cox\_transformation, [3](#)
- cor\_pmat, [3](#)
- detect\_decimals, [4](#)
- flattenCorrMatrix, [4](#)
  
- poma\_pal\_c, [33](#)
- poma\_pal\_d, [33](#)
- PomaBatch, [4](#)
- PomaBoxplots, [5](#)
- PomaClust, [7](#)
- PomaCorr, [8](#)
- PomaCreateObject, [9](#)
- PomaDensity, [10](#)
- PomaDESeq, [11](#)
- PomaHeatmap, [12](#)
- PomaImpute, [13](#)
- PomaLasso, [14](#)
- PomaLimma, [15](#)
- PomaLM, [16](#)
- PomaLMM, [17](#)
- PomaNorm, [19](#)
- PomaOddsRatio, [20](#)
- PomaOutliers, [21](#)
- PomaPCA, [22](#)
- PomaPCR, [23](#)
- PomaPLS, [24](#)
- PomaRandForest, [26](#)
- PomaRankProd, [27](#)
- PomaUMAP, [28](#)
- PomaUnivariate, [30](#)
- PomaVolcano, [31](#)
  
- quantile\_norm, [33](#)
  
- scale\_color\_poma\_c, [34](#)
- scale\_color\_poma\_d, [34](#)
- scale\_fill\_poma\_c, [34](#)
- scale\_fill\_poma\_d, [34](#)
- st000284, [35](#)
- st000336, [36](#)
- sum\_norm, [36](#)
  
- theme\_poma, [37](#)