

Package ‘NetPathMiner’

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Title NetPathMiner for Biological Network Construction, Path Mining and Visualization

Description NetPathMiner is a general framework for network path mining using genome-scale networks. It constructs networks from KGML, SBML and BioPAX files, providing three network representations, metabolic, reaction and gene representations. NetPathMiner finds active paths and applies machine learning methods to summarize found paths for easy interpretation. It also provides static and interactive visualizations of networks and paths to aid manual investigation.

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VignetteBuilder knitr

License GPL (>= 2)

URL <https://github.com/ahmohamed/NetPathMiner>

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NetPathMiner-package *General framework for network extraction, path mining.*

Description

NetPathMiner implements a flexible module-based process flow for network path mining and visualization, which can be fully integrated with user-customized functions. NetPathMiner supports construction of various types of genome scale networks from KGML, SBML and BioPAX formats, enabling its utility to most common pathway databases. NetPathMiner also provides different visualization techniques to facilitate the analysis of even thousands of output paths.

Author(s)

Ahmed Mohamed <mohamed@kuicr.kyoto-u.ac.jp>

assignEdgeWeights *Assigning weights to network edges*

Description

This function computes edge weights based on a gene expression profile.

Usage

```
assignEdgeWeights(microarray, graph, use.attr, y, weight.method = "cor",
  complex.method = "max", missing.method = "median",
  same.gene.penalty = "median", bootstrap = 100, verbose = TRUE)
```

Arguments

- microarray Microarray should be a Dataframe or a matrix, with genes as rownames, and samples as columns.
- graph An annotated igraph object.
- use.attr An attribute name to map microarray rows (genes) to graph vertices. The attribute must be annotated in graph, and the values correspond to rownames of microarray. You can check the coverage and if there are complex vertices using [getAttrStatus](#). You can eliminate complexes using [expandComplexes](#).
- y Sample labels, given as a factor or a character vector. This must be the same size as the columns of microarray

weight.method	A function, or a string indicating the name of the function to be used to compute the edge weights. The function is provided with 2 numerical vectors (2 rows from microarray), and it should return a single numerical value (or NA). The default computes Pearson's correlation.
complex.method	A function, or a string indicating the name of the function to be used in weighting edges connecting complexes. If a vertex has >1 attribute value, all possible pairwise weights are first computed, and given to complex.method. The default function is <code>max</code> .
missing.method	A function, or a string indicating the name of the function to be used in weighting edges when one of the vertices lack expression data. The function is passed all edge weights on the graph. Default is <code>median</code> .
same.gene.penalty	A numerical value to be assigned when 2 adjacent vertices have the same attribute value, since correlation and similarity measure will give perfect scores. Alternatively, same.gene.penalty can be a function, computing the penalty from all edge weights on the graph (excluding same-gene and missing values). The default is to take the <code>median</code>
bootstrap	An integer n, where the weight.method is performed on n permutations of the gene profiles, and taking the median value. Set it to NA to disable bootstrapping.
verbose	Print the progress of the function.

Value

The input graph with edge.weight as an edge attribute. The attribute can be a list of weights if y labels were provided.

Author(s)

Ahmed Mohamed

Examples

```
## Convert a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

# Using Spearman correlation, assigning missing edges to -1
## Not run:
assignEdgeWeights(microarray, graph, use.attr="miriam.affy.probeset",
y=factor(colnames(microarray)),
weight.method = function(x1,x2) cor(x1,x2, method="spearman"),
missing.method = -1)
```

```
## End(Not run)
```

biopax2igraph	<i>Processes BioPAX objects into igraph objects</i>
---------------	---

Description

This function takes BioPAX objects (level 2 or 3) as input, and returns either a metabolic or a signaling network as output.

Usage

```
biopax2igraph(biopax, parse.as = c("metabolic", "signaling"),
  expand.complexes = FALSE, inc.sm.molecules = FALSE, verbose = TRUE)
```

Arguments

biopax	BioPAX object generated by readBiopax .
parse.as	Whether to process file into a metabolic or a signaling network.
expand.complexes	Split protein complexes into individual gene nodes. Ignored if parse.as="metabolic".
inc.sm.molecules	Include small molecules that are participating in signaling events. Ignored if parse.as="metabolic".
verbose	Whether to display the progress of the function.

Details

This function requires `rBiopaxParser` installed.

Users can specify whether files are processes as metabolic or signaling networks.

Metabolic networks are given as bipartite graphs, where metabolites and reactions represent vertex types. Reactions are constructed from `Conversion` classes, connecting them to their corresponding `Lefts` and `Rights`. Each reaction vertex has `genes` attribute, listing all `Catalysis` relationships of this reaction. As a general rule, reactions inherit all annotation attributes of its catalyzing genes.

Signaling network have genes as vertices and edges represent interactions, such as activation / inhibition. Genes participating in successive reactions are also connected. Signaling interactions are constructed from `Control` classes, where edges are drawn from `controller` to `controlled`.

All annotation attributes are extracted from XRefs associated with the vertices, and are stored according to MIRIAM guidelines (`miraim.db`, where `db` is the database name).

Value

An igraph object, representing a metabolic or a signaling network.

Author(s)

Ahmed Mohamed

See Also

Other Database extraction methods: [KGML2igraph](#), [SBML2igraph](#)

Examples

```
if(requireNamespace("rBiopaxParser")){
  data(ex_biopax)
  # Process biopax as a metabolic network
  g <- biopax2igraph(ex_biopax)
  plotNetwork(g)

  # Process SBML file as a signaling network
  g <- biopax2igraph(ex_biopax, parse.as="signaling", expand.complexes=TRUE)
}
```

colorVertexByAttr *Computes colors for vertices according to their attributes.*

Description

This function returns a list of colors for vertices, assigned similar colors if they share a common attribute (ex: in the same pathway, etc).

Usage

```
colorVertexByAttr(graph, attr.name, col.palette = palette())
```

Arguments

graph	An annotated igraph object.
attr.name	The attribute name (ex: "pathway") by which vertices will be colored. Complex attributes, where a vertex belongs to more than one group, are supported.
col.palette	A color palette, or a palette generating function (ex: col.palette=rainbow).

Value

A list of colors (in HEX format) for vertices.

Author(s)

Ahmed Mohamed

See Also

Other Plotting methods: [layoutVertexByAttr](#), [plotAllNetworks](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotCytoscapeGML](#), [plotNetwork](#), [plotPathClassifier](#), [plotPaths](#)

Examples

```
data("ex_kgml_sig")
v.colors <- colorVertexByAttr(ex_kgml_sig, "pathway")
plotNetwork(ex_kgml_sig, vertex.color=v.colors)
```

expandComplexes

Expand reactions / complexes into their gene constituents.

Description

These are general functions to expand vertices by their attributes, i.e. create a separate vertex for each attribute value.

Usage

```
expandComplexes(graph, v.attr, keep.parent.attr = "^pathway",
  expansion.method = c("normal", "duplicate"),
  missing.method = c("keep", "remove", "reconnect"))

makeGeneNetwork(graph, v.attr = "genes", keep.parent.attr = "^pathway",
  expansion.method = "duplicate", missing.method = "remove")
```

Arguments

graph	An annotated igraph object.
v.attr	Name of the attribute which vertices are expanded to.
keep.parent.attr	A (List of) regex expressions representing attributes to be inherited by daughter vertices. If "all" is passed, all parent attributes are inherited.
expansion.method	If "duplicate", attribute values sharing more than one parent vertex are duplicated for each vertex they participate in. For example, if one gene G1 catalyzes reactions R1, R2; then G1##R1, and G1##R2 vertices are created. If "normal" only one vertex (G1) is created, and inherit all R1 and R2 connections and attributes.
missing.method	How to deal with vertices with no attribute values. "keep" retains the parent node, "remove" simply deletes the vertex, and "reconnect" removes the vertex and connect its neighbours to each other (to prevent graph cuts).

Details

These functions can be very useful when merging networks constructed from different databases. For example, to match a network created from Reactome to a KEGG network, you can expand metabolite vertices by "miriam.kegg.compound" attribute.

Value

A new graph with vertices expanded.

makeGeneNetwork returns a graph, where nodes are genes, and edges represent participation in successive reactions.

Author(s)

Ahmed Mohamed

See Also

Other Network processing methods: [makeMetaboliteNetwork](#), [makeReactionNetwork](#), [reindexNetwork](#), [rmSmallCompounds](#), [simplifyReactionNetwork](#), [vertexDeleteReconnect](#)

Examples

```
## Make a gene network from a reaction network.
data(ex_sbml) # A bipartite metabolic network.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)
ggraph <- makeGeneNetwork(rgraph)

## Expand vertices into their constituent genes.
data(ex_kgml_sig) # Ras and chemokine signaling pathways in human
ggraph <- expandComplexes(ex_kgml_sig, v.attr = "miriam.ncbigene",
  keep.parent.attr= c("^pathway", "^compartment"))

## Create a separate vertex for each compartment. This is useful in duplicating
## metabolite vertices in a network.
## Not run:
graph <- expandComplexes(graph, v.attr = "compartment",
  keep.parent.attr = "all",
  expansion.method = "duplicate",
  missing.method = "keep")

## End(Not run)
```

extractPathNetwork *Creates a subnetwork from a ranked path list*

Description

Creates a subnetwork from a ranked path list generated by [pathRanker](#).

Usage

```
extractPathNetwork(paths, graph)
```

Arguments

paths	The paths extracted by pathRanker .
graph	A annotated igraph object.

Value

A subnetwork from all paths provided. If paths are computed for several labels (sample categories), a subnetwork is returned for each label.

Author(s)

Ahmed Mohamed

See Also

Other Path ranking methods: [getPathsAsEIDs](#), [pathRanker](#), [samplePaths](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=6)

## Get the subnetwork of paths in reaction graph.
reaction.sub <- getPathsAsEIDs(ranked.p, rgraph)
```

```
## Get the subnetwork of paths in the original metabolic graph.  
metabolic.sub <- getPathsAsEIDs(ranked.p, ex_sbml)
```

ex_biopax *Biopax example data*

Description

A dataset containing Porphyrin metabolism pathway in Biopax Level 3 and parsed with [readBiopax](#).

Examples

```
data(ex_biopax)  
ex_biopax
```

ex_kgml_sig *Signaling network from KGML example*

Description

An example igraph object representing Ras and chemokine signaling pathways in human extracted from KGML files.

Examples

```
data(ex_kgml_sig)  
plotNetwork(ex_kgml_sig, vertex.color="pathway")
```

ex_microarray *An microarray data example.*

Description

An microarray data example. This is part of the ALL dataset, for demonstration purposes.

Examples

```
data(ex_microarray)
```

ex_sbml	<i>Metabolic network from SBML example</i>
---------	--

Description

An example igraph object representing bipartite metabolic network of Carbohydrate metabolism extracted from SBML file from Reactome database.

Examples

```
data(ex_sbml)
plotNetwork(ex_sbml, vertex.color="compartment.name")
```

getAttrStatus	<i>Get / Set vertex attribute names and coverage</i>
---------------	--

Description

These functions report the annotation status of the vertices of a given network, modify or remove certain annotations.

Usage

```
getAttrStatus(graph, pattern = "^miriam.")
getAttrNames(graph, pattern = "")
getAttribute(graph, attr.name)
setAttribute(graph, attr.name, attr.value)
rmAttribute(graph, attr.name)
```

Arguments

graph	An annotated igraph object.
pattern	A regex expression representing attribute name pattern.
attr.name	The attribute name
attr.value	A list of attribute values. This must be the same size as the number of vertices.

Details

NetPathMiner stores all its vertex annotation attributes in a list, and stores them collectively as a single attr. This is not to interfere with [graph_attr_names](#) from igraph package. All functions here target NetPathMiner annotations only.

Value

For `getAttrStatus`, a dataframe summarizing the number of vertices with no (missing), one (single) or more than one (complex) attribute value. The coverage

For `getAttrNames`, a character vector of attribute names matching the pattern.

For `getAttribute`, a list of vertex annotation values for the query attribute.

For `setAttribute`, a graph with the new attribute set.

For `rmAttrNames`, a new igraph object with the attribute removed.

Author(s)

Ahmed Mohamed

See Also

Other Attribute handling methods: [stdAttrNames](#)

Examples

```
data(ex_kgml_sig) # Ras and chemokine signaling pathways in human

# Get status of attribute "pathway" only
getAttrStatus(ex_kgml_sig, "^pathway$")

# Get status of all attributes starting with "pathway" and "miriam" keywords
getAttrStatus(ex_kgml_sig, "(^miriam)|(^pathway)")

# Get all attribute names containing "miriam"
getAttrNames(ex_kgml_sig, "miriam")
# Get all attribute names containing "miriam"
getAttribute(ex_kgml_sig, "miriam.ncbigene")

# Remove an attribute from graph
graph <- rmAttribute(ex_kgml_sig, "miriam.ncbigene")
```

getGeneSetNetworks *Generate geneset networks from an annotated network.*

Description

This function generates geneset networks based on a given network, by grouping vertices sharing common attributes (in the same pathway or compartment).

Usage

```
getGeneSetNetworks(graph, use.attr = "pathway", format = c("list",
  "pathway-class"))
```

Arguments

graph	An annotated igraph object..
use.attr	The attribute by which vertices are grouped (typically pathway, or GO)
format	The output format. If "list" is specified, a list of subgraphs are returned (default). If "pathway-class" is specified, a list of pathway-class objects are returned. Pathway-class is used by graphite package to run several methods of topology-based enrichment analyses.

Value

A list of geneset networks as igraph or Pathway-class objects.

Author(s)

Ahmed Mohamed

See Also

[getGeneSets](#)

Examples

```
data(ex_kgml_sig) # Ras and chemokine signaling pathways in human
genesetnets <- getGeneSetNetworks(ex_kgml_sig, use.attr="pathway")

# Integration with graphite package
## Not run:
if(requireNamespace("graphite") & requireNamespace("clipper") & requireNamespace("ALL")){
genesetnets <- getGeneSetNetworks(ex_kgml_sig,
use.attr="pathway", format="pathway-class")
path <- convertIdentifiers(genesetnets$`Chemokine signaling pathway`,
"entrez")
genes <- nodes(path)
data(ALL)
all <- as.matrix(exprs(ALL[1:length(genes),1:20]))
classes <- c(rep(1,10), rep(2,10))
rownames(all) <- genes

runClipper(path, all, classes, "mean", pathThr=0.1)
}

## End(Not run)
```

getGeneSets	<i>Generate genesets from an annotated network.</i>
-------------	---

Description

This function generates genesets based on a given network, by grouping vertices sharing common attributes (in the same pathway or compartment). Genes associated with each vertex can be specified through `gene.attr` argument.

Usage

```
getGeneSets(graph, use.attr = "pathway", gene.attr = "genes", gmt.file)
```

Arguments

<code>graph</code>	An annotated igraph object..
<code>use.attr</code>	The attribute by which vertices are grouped (typically pathway, or GO)
<code>gene.attr</code>	The attribute listing genes annotated with each vertex (ex: <code>miriam.ncbigene</code> , <code>miriam.uniprot</code> , ...)
<code>gmt.file</code>	Optional. If provided, Results are exported to a GMT file. GMT files are readily used by most gene set analysis packages.

Value

A list of genesets or written to gmt file if provided.

Author(s)

Ahmed Mohamed

See Also

[getGeneSetNetworks](#)

Examples

```
data(ex_kgml_sig) # Ras and chemokine signaling pathways in human
genesets <- getGeneSets(ex_kgml_sig, use.attr="pathway", gene.attr="miriam.ncbigene")

# Write the genesets in a GMT file, and read it using GSEABase package.
getGeneSets(ex_kgml_sig, use.attr="pathway", gene.attr="miriam.ncbigene", gmt.file="kgml.gmt")
## Not run:
if(requireNamespace("GSEABase"))
  toGmt("kgml.gmt")

## End(Not run)
```

```
# Create genesets using compartment information
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
genesets <- getGeneSets(ex_sbml, use.attr="compartment.name", gene.attr="miriam.uniprot")
```

getPathsAsEIDs *Convert a ranked path list to edge ids of a graph*

Description

Convert a ranked path list to Edge ids of a graph, where paths can come from a different representation (for example matching path from a reaction network to edges on a metabolic network).

Usage

```
getPathsAsEIDs(paths, graph)
```

Arguments

paths	The paths extracted by pathRanker .
graph	A annotated igraph object.

Value

A list of edge ids on the provided graph.

Author(s)

Ahmed Mohamed

See Also

Other Path ranking methods: [extractPathNetwork](#), [pathRanker](#), [samplePaths](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
```

```

ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=6)

## Get the edge ids along paths in the reaction graph.
path.eids <- getPathsAsEIDs(ranked.p, rgraph)

## Get the edge ids along paths in the original metabolic graph.
path.eids <- getPathsAsEIDs(ranked.p, ex_sbml)

```

KGML2igraph

Processes KGML files into igraph objects

Description

This function takes KGML files as input, and returns either a metabolic or a signaling network as output.

Usage

```

KGML2igraph(filename, parse.as = c("metabolic", "signaling"),
expand.complexes = FALSE, verbose = TRUE)

```

Arguments

filename	A character vector containing the KGML files to be processed. If a directory path is provided, all *.xml files in it and its subdirectories are included.
parse.as	Whether to process file into a metabolic or a signaling network.
expand.complexes	Split protein complexes into individual gene nodes. This argument is ignored if parse.as="metabolic"
verbose	Whether to display the progress of the function.

Details

Users can specify whether files are processes as metabolic or signaling networks.

Metabolic networks are given as bipartite graphs, where metabolites and reactions represent vertex types. This is constructed from <reaction> xml node in KGML file, connecting them to their corresponding substrates and products. Each reaction vertex has genes attribute, listing all genes associated with the reaction. As a general rule, reactions inherit all annotation attributes of its catalyzing genes.

Signaling network have genes as vertices and edges represent interactions, such as activation / inhibition. Genes participating in successive reactions are also connected. Signaling parsing method processes <ECrel>, <PPrel> and <PCrel> interactions from KGML files.

To generate a genome scale network, simply provide a list of files to be parsed, or put all file in a directory, as pass the directory path as filename

Value

An igraph object, representing a metabolic or a signaling network.

Author(s)

Ahmed Mohamed

See Also

Other Database extraction methods: [SBML2igraph](#), [biopax2igraph](#)

Examples

```
if(is.loaded("readkgmlfile")){ # This is false if libxml2 wasn't available at installation.
  filename <- system.file("extdata", "hsa00860.xml", package="NetPathMiner")

  # Process KGML file as a metabolic network
  g <- KGML2igraph(filename)
  plotNetwork(g)

  # Process KGML file as a signaling network
  g <- KGML2igraph(filename, parse.as="signaling", expand.complexes=TRUE)
  plotNetwork(g)
}
```

layoutVertexByAttr *A graph layout function, which groups vertices by attribute.*

Description

This function generates a layout for igraph objects, keeping vertices with the same attribute (ex: in the same pathway, etc) close to each other.

Usage

```
layoutVertexByAttr(graph, attr.name, cluster.strength = 1,
  layout = layout.auto)
```

Arguments

graph	An annotated igraph object.
attr.name	The attribute name by which vertices are laid out.
cluster.strength	A number indicating tie strengths between vertices with the same attribute. The larger it is, the closer the vertices will be.
layout	A layout function, ideally a force-directed layout function, such as layout_with_fr and layout_with_kk .

Value

A two-column matrix indicating the x and y positions of vertices.

Author(s)

Ahmed Mohamed

See Also

Other Plotting methods: [colorVertexByAttr](#), [plotAllNetworks](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotCytoscapeGML](#), [plotNetwork](#), [plotPathClassifier](#), [plotPaths](#)

Examples

```
data("ex_kgml_sig")
v.layout <- layoutVertexByAttr(ex_kgml_sig, "pathway")
plotNetwork(ex_kgml_sig, vertex.color="pathway", layout=v.layout)

v.layout <- layoutVertexByAttr(ex_kgml_sig, "pathway", cluster.strength=5)
plotNetwork(ex_kgml_sig, vertex.color="pathway", layout=v.layout)
```

makeMetaboliteNetwork *Convert metabolic network to metabolite network.*

Description

This function removes reaction nodes keeping them as edge attributes. The resulting network contains metabolite nodes only, where edges indicate that reaction conversions.

Usage

```
makeMetaboliteNetwork(graph)
```

Arguments

graph A metabolic network.

Value

A reaction network.

Author(s)

Ahmed Mohamed

See Also

Other Network processing methods: [expandComplexes](#), [makeReactionNetwork](#), [reindexNetwork](#), [rmSmallCompounds](#), [simplifyReactionNetwork](#), [vertexDeleteReconnect](#)

Examples

```
## Conver a metabolic network to a metbolite network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
mgraph <- makeMetaboliteNetwork(ex_sbml)
```

makeReactionNetwork *Convert metabolic network to reaction network.*

Description

This function removes metabolite nodes keeping them as edge attributes. The resulting network contains reaction nodes only, where edges indicate that a metabolite produced by one reaction is consumed by the other.

Usage

```
makeReactionNetwork(graph, simplify = FALSE)
```

Arguments

graph	A metabolic network.
simplify	An option to remove translocation and spontaneous reactions that require no catalyzing genes. Translocation reactions are detected from reaction name (SBML, BioPAX), or by having identical substrates and products.

Value

A reaction network.

Author(s)

Ahmed Mohamed

See Also

Other Network processing methods: [expandComplexes](#), [makeMetaboliteNetwork](#), [reindexNetwork](#), [rmSmallCompounds](#), [simplifyReactionNetwork](#), [vertexDeleteReconnect](#)

Examples

```
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)
```

NPMdefaults

Default values for NetPathMiner

Description

This function gets a NetPathMiner default value for a variable.

Usage

```
NPMdefaults(value)
```

Arguments

value a character string indicating the variable name.

Details

NetPathMiner defines the following defaults:

- `small.comp.ls` Dataframe of ubiquitous metabolites. Used by [rmSmallCompounds](#).
- `bridge` Dataframe of attributes supported by Bridge Database. Used by [fetchAttribute](#).
- `bridge.organisms` A list of bridge supported organisms. Used by [fetchAttribute](#).
- `bridge.web` The base URL for Bridge Database webservice. Used by [fetchAttribute](#).

Value

The default value for the given variable.

Author(s)

Ahmed Mohamed

Examples

```
# Get the default list of small compounds (ubiquitous metabolites).  
NPMdefaults("small.comp.ls")
```

pathClassifier *HME3M Markov pathway classifier.*

Description

HME3M Markov pathway classifier.

Usage

```
pathClassifier(paths, target.class, M, alpha = 1, lambda = 2,
  hme3miter = 100, plriter = 1, init = "random")
```

Arguments

paths	The training paths computed by pathsToBinary
target.class	he label of the targe class to be classified. This label must be present as a label within the paths\y object
M	Number of components within the paths to be extracted.
alpha	The PLR learning rate. (between 0 and 1).
lambda	The PLR regularization parameter. (between 0 and 2)
hme3miter	Maximum number of HME3M iterations. It will stop when likelihood change is < 0.001.
plriter	Maximum number of PLR interactions. It will stop when likelihood change is < 0.001.
init	Specify whether to initialize the HME3M responsibilities with the 3M model - random is recommended.

Details

Take care with selection of lambda and alpha - make sure you check that the likelihood is always increasing.

Value

A list with the following elements. A list with the following values

h	A dataframe with the EM responsibilities.
theta	A dataframe with the Markov parameters for each component.
beta	A dataframe with the PLR coefficients for each component.
proportions	The probability of each HME3M component.
posterior.probs	The HME3M posterior probability.
likelihood	The likelihood convergence history.
plrplr	The posterior predictions from each components PLR model.

path.probabilities	The 3M probabilities for each path belonging to each component.
params	The parameters used to build the model.
y	The binary response variable used by HME3M. A 1 indicates the location of the target.class labels in paths\\$.y
perf	The training set ROC curve AUC.
label	The HME3M predicted label for each path.
component	The HME3M component assignment for each path.

Author(s)

Timothy Hancock and Ichigaku Takigawa

References

Hancock, Timothy, and Mamitsuka, Hiroshi: A Markov Classification Model for Metabolic Pathways, Workshop on Algorithms in Bioinformatics (WABI), 2009

Hancock, Timothy, and Mamitsuka, Hiroshi: A Markov Classification Model for Metabolic Pathways, Algorithms for Molecular Biology 2010

See Also

Other Path clustering & classification methods: [pathCluster](#), [pathsToBinary](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotPathClassifier](#), [plotPathCluster](#), [predictPathClassifier](#), [predictPathCluster](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=6)

## Convert paths to binary matrix.
ybinpaths <- pathsToBinary(ranked.p)
p.class <- pathClassifier(ybinpaths, target.class = "BCR/ABL", M = 3)

## Contingency table of classification performance
table(ybinpaths$y,p.class$label)
```

```
## Plotting the classifier results.  
plotClassifierROC(p.class)  
plotClusters(ybinpaths, p.class)
```

pathCluster	<i>3M Markov mixture model for clustering pathways</i>
-------------	--

Description

3M Markov mixture model for clustering pathways

Usage

```
pathCluster(ybinpaths, M, iter = 1000)
```

Arguments

ybinpaths	The training paths computed by pathsToBinary .
M	The number of clusters.
iter	The maximum number of EM iterations.

Value

A list with the following items:

h	The posterior probabilities that each path belongs to each cluster.
labels	The cluster membership labels.
theta	The probabilities of each gene for each cluster.
proportions	The mixing proportions of each path.
likelihood	The likelihood convergence history.
params	The specific parameters used.

Author(s)

Ichigaku Takigawa
Timothy Hancock

References

Mamitsuka, H., Okuno, Y., and Yamaguchi, A. 2003. Mining biologically active patterns in metabolic pathways using microarray expression profiles. *SIGKDD Explor. News* 1, 5, 2 (Dec. 2003), 113-121.

See Also

Other Path clustering & classification methods: [pathClassifier](#), [pathsToBinary](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotPathClassifier](#), [plotPathCluster](#), [predictPathClassifier](#), [predictPathCluster](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot", bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=8)

## Convert paths to binary matrix.
ybinpaths <- pathsToBinary(ranked.p)
p.cluster <- pathCluster(ybinpaths, M=2)
plotClusters(ybinpaths, p.cluster)
```

pathRanker

Extracting and ranking paths from a network

Description

Given a weighted igraph object, path ranking finds a set of node/edge sequences (paths) to maximize the sum of edge weights. `pathRanker(method="prob.shortest.path")` extracts the K most probable paths within a weighted network. `pathRanker(method="pvalue")` extracts a list of paths whose sum of edge weights are significantly higher than random paths of the same length.

Usage

```
pathRanker(graph, method = c("prob.shortest.path", "pvalue"), start, end,
verbose = TRUE, ...)
```

Arguments

graph	A weighted igraph object. Weights must be in <code>edge.weights</code> or <code>weight edge</code> attributes.
method	Which path ranking method to use.
start	A list of start vertices, given by their vertex id.

end	A list of terminal vertices, given by their vertex id.
verbose	Whether to display the progress of the function.
...	Method-specific parameters. See Details section.

Details

The input here is graph. A weight must be assigned to each edge. Bootstrapped Pearson correlation edge weights can be assigned to each edge by [assignEdgeWeights](#). However the specification of the edge weight is flexible with the condition that increasing values indicate stronger relationships between vertices.

Probabilistic Shortest Paths: `pathRanker(method="prob.shortest.path")` finds the K most probable loopless paths given a weighted network. Before the paths are ranked the edge weights are converted into probabilistic edge weights using the Empirical Cumulative Distribution (ECDF) over all edge weights. This is called ECDF edge weight. The ECDF edge weight serves as a probabilistic rank of the most important gene-gene interactions. The probabilistic nature of the ECDF edge weights allow for a significance test to determine if a path contains any functional structure or is simply a random walk. The probability of a path is simply the product of all ECDF weights along the path. This is computed as a sum of the logs of the ECDF edge weights.

The following arguments can be passed to `pathRanker(method="prob.shortest.path")`:

`K` Maximum number of paths to extract. Defaults to 10.

`minPathSize` The minimum number of edges for each extracted path. Defaults to 1.

`normalize` Specify if you want to normalize the probabilistic edge weights (across different labels) before extracting the paths. Defaults to TRUE.

P-value method: `pathRanker(method="pvalue")` searches all paths between the specified start and end vertices, and if a significant path is found it returns it. However, it doesn't search for the best path between the start and terminal vertices, as there could be many paths which lead to the same terminal vertex, and searching through all of them is time consuming. We just stop when the first significant path is found.

All provided edge weights are recaled from 0-1. Path significance is calculated based on the empirical distribution of random paths of the same length. This can be estimated using [samplePaths](#) and passed as an argument.

The following arguments can be passed to `pathRanker(method="pvalue")`:

`sampledpaths` The empirical results from [samplePaths](#).

`alpha` The P value cut-off. Defaults to 0.01

Value

A list of paths where each path has the following items:

gene	The ordered sequence of genes visited along the path.
compounds	The ordered sequence of compounds visited along the path.
weights	The ordered sequence of the log(ECDF edge weights) along the path.
distance	The sum of the log(ECDF edge weights) along each path. (a sum of logs is a product)

Author(s)

Timothy Hancock, Ichigaku Takigawa, Nicolas Wicker and Ahmed Mohamed

See Also

getPathAsEIDs, extractPathNetwork

Other Path ranking methods: [extractPathNetwork](#), [getPathAsEIDs](#), [samplePaths](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=6)

## Get significantly correlated paths using "p-value" method.
## First, establish path score distribution by calling "samplePaths"
pathsample <- samplePaths(rgraph, max.path.length=10,
num.samples=100, num.warmup=10)

## Get all significant paths with p<0.1
significant.p <- pathRanker(rgraph, method = "pvalue",
sampledpaths = pathsample ,alpha=0.1)
```

pathsToBinary

Converts the result from pathRanker into something suitable for pathClassifier or pathCluster.

Description

Converts the result from pathRanker into something suitable for pathClassifier or pathCluster.

Usage

```
pathsToBinary(ypaths)
```

Arguments

ypaths The result of [pathRanker](#).

Details

Converts a set of pathways from [pathRanker](#) into a list of binary pathway matrices. If the pathways are grouped by a response label then the *pathsToBinary* returns a list labeled by response class where each element is the binary pathway matrix for each class. If the pathways are from [pathRanker](#) then a list with a single element containing the binary pathway matrix is returned. To look up the structure of a specific binary path in the corresponding ypaths object simply use matrix index by calling `ypaths[[ybinpaths$pidx[i,]]]`, where *i* is the row in the binary paths object you wish to reference.

Value

A list with the following elements.

paths	All paths within ypaths converted to a binary string and concatenated into the one matrix.
y	The response variable.
pidx	An matrix where each row specifies the location of that path within the ypaths object.

Author(s)

Timothy Hancock and Ichigaku Takigawa

See Also

Other Path clustering & classification methods: [pathClassifier](#), [pathCluster](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotPathClassifier](#), [plotPathCluster](#), [predictPathClassifier](#), [predictPathCluster](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=6)
```

```
## Convert paths to binary matrix.
ybinpaths <- pathsToBinary(ranked.p)
p.cluster <- pathCluster(ybinpaths, M=3)
plotClusters(ybinpaths, p.cluster, col=c("red", "green", "blue") )
```

plotAllNetworks *Highlighting ranked paths over multiple network representations.*

Description

This function highlighting ranked paths over different network representations, metabolic, reaction and gene networks. The functions finds equivalent paths across different networks and marks them.

Usage

```
plotAllNetworks(paths, metabolic.net = NULL, reaction.net = NULL,
  gene.net = NULL, path.clusters = NULL, plot.clusters = TRUE,
  col.palette = palette(), layout = layout.auto, ...)
```

Arguments

paths	The result of pathRanker .
metabolic.net	A bipartite metabolic network.
reaction.net	A reaction network, resulting from makeReactionNetwork .
gene.net	A gene network, resulting from makeGeneNetwork .
path.clusters	The result from pathCluster or pathClassifier .
plot.clusters	Whether to plot clustering information, as generated by plotClusters
col.palette	A color palette, or a palette generating function (ex: col.palette=rainbow).
layout	Either a graph layout function, or a two-column matrix specifying vertex coordinates.
...	Additional arguments passed to plotNetwork .

Value

Highlights the path list over all provided networks.

Author(s)

Ahmed Mohamed

See Also

Other Plotting methods: [colorVertexByAttr](#), [layoutVertexByAttr](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotCytoscapeGML](#), [plotNetwork](#), [plotPathClassifier](#), [plotPaths](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=6)

plotAllNetworks(ranked.p, metabolic.net = ex_sbml, reaction.net = rgraph,
vertex.label = "", vertex.size = 4)
```

plotClassifierROC *Diagnostic plots for pathClassifier.*

Description

Diagnostic plots for [pathClassifier](#).

Usage

```
plotClassifierROC(mix)
```

Arguments

mix The result from [pathClassifier](#).

Value

Diagnostic plots of the result from [pathClassifier](#). `itemTop` ROC curves for the posterior probabilities (`mix$posterior.probs`) and for each HME3M component (`mix$h`). This gives information about what response label each relates to. A ROC curve with an $AUC < 0.5$ relates to $y = 0$. Conversely ROC curves with $AUC > 0.5$ relate to $y = 1$. `itemBottom` The likelihood convergence history for the HME3M model. If the parameters `alpha` or `lambda` are set too large then the likelihood may decrease.

Author(s)

Timothy Hancock and Ichigaku Takigawa

See Also

Other Path clustering & classification methods: [pathClassifier](#), [pathCluster](#), [pathsToBinary](#), [plotClusterMatrix](#), [plotPathClassifier](#), [plotPathCluster](#), [predictPathClassifier](#), [predictPathCluster](#)

Other Plotting methods: [colorVertexByAttr](#), [layoutVertexByAttr](#), [plotAllNetworks](#), [plotClusterMatrix](#), [plotCytoscapeGML](#), [plotNetwork](#), [plotPathClassifier](#), [plotPaths](#)

plotClusterMatrix *Plots the structure of all path clusters*

Description

Plots the structure of all path clusters

Usage

```
plotClusterMatrix(ybinpaths, clusters, col = rainbow(clusters$params$M),
  grid = TRUE)
```

```
plotClusterProbs(clusters, col = rainbow(clusters$params$M))
```

```
plotClusters(ybinpaths, clusters, col, ...)
```

Arguments

ybinpaths	The training paths computed by pathsToBinary .
clusters	The pathway cluster model trained by pathCluster or pathClassifier .
col	Colors for each path cluster.
grid	A logical, whether to add a grid to the plot
...	Extra paramaters passed to plotClusterMatrix

Value

[plotClusterMatrix](#) plots an image of all paths the training dataset. Rows are the paths and columns are the genes (features) included within each path. Paths are colored according to cluster membership.

[plotClusterProbs](#) The training set posterior probabilities for each path belonging to a 3M component.

[plotClusters](#): combines the two plots produced by [plotClusterProbs](#) and [plotClusterMatrix](#).

Author(s)

Ahmed Mohamed

See Also

Other Path clustering & classification methods: [pathClassifier](#), [pathCluster](#), [pathsToBinary](#), [plotClassifierROC](#), [plotPathClassifier](#), [plotPathCluster](#), [predictPathClassifier](#), [predictPathCluster](#)

Other Plotting methods: [colorVertexByAttr](#), [layoutVertexByAttr](#), [plotAllNetworks](#), [plotClassifierROC](#), [plotCytoscapeGML](#), [plotNetwork](#), [plotPathClassifier](#), [plotPaths](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=8)

## Convert paths to binary matrix.
ybinpaths <- pathsToBinary(ranked.p)
p.cluster <- pathCluster(ybinpaths, M=2)
plotClusters(ybinpaths, p.cluster, col=c("red", "blue") )
```

`plotCytoscapeGML`*Plots an annotated igraph object in Cytoscape.*

Description

`plotCytoscape` function has been removed because RCytoscape is no longer present in Bioconductor. Future plans will use RCy3 for Cytoscape plotting, once RCy3 is supported on MacOS and Windows. [plotCytoscapeGML](#) exports the network plot in GML format, that can be later imported into Cytoscape (using "import network from file" option). This function is compatible with all Cytoscape versions.

Usage

```
plotCytoscapeGML(graph, file, layout = layout.auto, vertex.size,
vertex.label, vertex.shape, vertex.color, edge.color)
```

Arguments

graph	An annotated igraph object.
file	Output GML file name to which the network plot is exported.
layout	Either a graph layout function, or a two-column matrix specifying vertex coordinates.
vertex.size	Vertex size. If missing, the vertex attribute "size" ($V(g)$size$) will be used.
vertex.label	Vertex labels. If missing, the vertex attribute "label" ($V(g)$label$) will be used. If missing, vertices are labeled by their name.
vertex.shape	Vertex shape in one of igraph shapes. If missing, the vertex attribute "shape" ($V(g)$shape$) will be used. Shapes are converted from igraph convention to Cytoscape convention. "square", "rectangle" and "vrectangle" are converted to "RECT", "csquare" and "crectangle" are converted to "ROUND_RECT", all other shapes are considered "ELLIPSE"
vertex.color	A color or a list of colors for vertices. Vertices with multiple colors are not supported. If missing, the vertex attribute "color" ($V(g)$color$) will be used.
edge.color	A color or a list of colors for edges. If missing, the edge attribute "color" ($E(g)$color$) will be used.

Value

For plotCytoscapeGML, results are written to file.

Author(s)

Ahmed Mohamed

See Also

Other Plotting methods: [colorVertexByAttr](#), [layoutVertexByAttr](#), [plotAllNetworks](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotNetwork](#), [plotPathClassifier](#), [plotPaths](#)

Examples

```

data("ex_sbml")
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)
v.layout <- layoutVertexByAttr(rgraph, "compartment")
v.color <- colorVertexByAttr(rgraph, "compartment")

# Export network plot to GML file
plotCytoscapeGML(rgraph, file="example.gml", layout=v.layout,
vertex.color=v.color, vertex.size=10)

```

plotNetwork

Plots an annotated igraph object.

Description

This function is a wrapper function for `plot.igraph`, with 2 main additions. 1. Add the ability to color vertices by their attributes (see examples), accompanied by an informative legend. 2. Resize `vertex.size`, `edge.arrow.size`, `label.cex` according to the plot size and the size of the network.

Usage

```

plotNetwork(graph, vertex.color, col.palette = palette(),
  layout = layout.auto, legend = TRUE, ...)

```

Arguments

<code>graph</code>	An annotated igraph object.
<code>vertex.color</code>	A list of colors for vertices, or an attribute names (ex: "pathway") by which vertices will be colored. Complex attributes, where a vertex belongs to more than one group, are supported. This can also be the output of <code>colorVertexByAttr</code> .
<code>col.palette</code>	A color palette, or a palette generating function (ex: <code>col.palette=rainbow</code>).
<code>layout</code>	Either a graph layout function, or a two-column matrix specifying vertex coordinates.
<code>legend</code>	Whether to plot a legend. The legend is only plotted if vertices are colored by attribute values.
<code>...</code>	Additional arguments passed to <code>plot.igraph</code> .

Value

Produces a plot of the network.

Author(s)

Ahmed Mohamed

See Also

Other Plotting methods: [colorVertexByAttr](#), [layoutVertexByAttr](#), [plotAllNetworks](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotCytoscapeGML](#), [plotPathClassifier](#), [plotPaths](#)

Examples

```
data("ex_kgml_sig")
plotNetwork(ex_kgml_sig, vertex.color="pathway")
plotNetwork(ex_kgml_sig, vertex.color="pathway", col.palette=heat.colors)
plotNetwork(ex_kgml_sig, vertex.color="pathway",
            col.palette=c("red", "green", "blue", "grey"))
```

`plotPathClassifier` *Plots the structure of specified path found by pathClassifier.*

Description

Plots the structure of specified path found by pathClassifier.

Usage

```
plotPathClassifier(ybinpaths, obj, m, tol = NULL)
```

Arguments

<code>ybinpaths</code>	The training paths computed by pathsToBinary
<code>obj</code>	The pathClassifier pathClassifier .
<code>m</code>	The path component to view.
<code>tol</code>	A tolerance for 3M parameter theta which is the probability for each edge within each cluster. If the tolerance is set all edges with a theta below that tolerance will be removed from the plot.

Value

Produces a plot of the paths with the path probabilities and prediction probabilities and ROC curve overlaid.

Center Plot An image of all paths the training dataset. Rows are the paths and columns are the genes (vertices) included within each pathway. A colour within image indicates if a particular gene (vertex) is included within a specific path. Colours flag whether a path belongs to the current HME3M component ($P > 0.5$).

Center Right	The training set posterior probabilities for each path belonging to the current 3M component.
Center Top	The ROC curve for this HME3M component.
Top Bar Plots	Theta: The 3M component probabilities - indicates the importance of each edge is to a path. Beta: The PLR coefficient - the magnitude indicates the importance of the edge to the classify the response.

Author(s)

Timothy Hancock and Ichigaku Takigawa

See Also

Other Path clustering & classification methods: [pathClassifier](#), [pathCluster](#), [pathsToBinary](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotPathCluster](#), [predictPathClassifier](#), [predictPathCluster](#)

Other Plotting methods: [colorVertexByAttr](#), [layoutVertexByAttr](#), [plotAllNetworks](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotCytoscapeGML](#), [plotNetwork](#), [plotPaths](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=6)

## Convert paths to binary matrix.
ybinpaths <- pathsToBinary(ranked.p)
p.class <- pathClassifier(ybinpaths, target.class = "BCR/ABL", M = 3)

## Plotting the classifier results.
plotClassifierROC(p.class)
plotClusters(ybinpaths, p.class)
```

plotPathCluster	<i>Plots the structure of specified path cluster</i>
-----------------	--

Description

Plots the structure of specified path found by pathCluster.

Usage

```
plotPathCluster(ybinpaths, clusters, m, tol = NULL)
```

Arguments

ybinpaths	The training paths computed by pathsToBinary .
clusters	The pathway cluster model trained by pathCluster or pathClassifier .
m	The path cluster to view.
tol	A tolerance for 3M parameter theta which is the probability for each edge within each cluster. If the tolerance is set all edges with a theta below that tolerance will be removed from the plot.

Value

Produces a plot of the paths with the path probabilities and cluster membership probabilities.

Center Plot	An image of all paths the training dataset. Rows are the paths and columns are the genes (features) included within each path.
Right	The training set posterior probabilities for each path belonging to the current 3M component.
Top Bar Plots	Theta, The 3M component probabilities - indicates the importance of each edge to a pathway.

Author(s)

Timothy Hancock and Ichigaku Takigawa

See Also

Other Path clustering & classification methods: [pathClassifier](#), [pathCluster](#), [pathsToBinary](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotPathClassifier](#), [predictPathClassifier](#), [predictPathCluster](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot", bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=8)

## Convert paths to binary matrix.
ybinpaths <- pathsToBinary(ranked.p)
p.cluster <- pathCluster(ybinpaths, M=2)
plotPathCluster(ybinpaths, p.cluster, m=2, tol=0.05)
```

plotPaths

Plots an annotated igraph object highlighting ranked paths.

Description

This function plots a network highlighting ranked paths. If `path.clusters` are provided, paths in the same cluster are assigned similar colors.

Usage

```
plotPaths(paths, graph, path.clusters = NULL, col.palette = palette(),
layout = layout.auto, ...)
```

Arguments

<code>paths</code>	The result of pathRanker .
<code>graph</code>	An annotated igraph object.
<code>path.clusters</code>	The result from pathCluster or pathClassifier .
<code>col.palette</code>	A color palette, or a palette generating function (ex: <code>col.palette=rainbow</code>).
<code>layout</code>	Either a graph layout function, or a two-column matrix specifying vertex coordinates.
<code>...</code>	Additional arguments passed to plotNetwork .

Value

Produces a plot of the network with paths highlighted. If paths are computed for several labels (sample categories), a plot is created for each label.

Author(s)

Ahmed Mohamed

See Also

Other Plotting methods: [colorVertexByAttr](#), [layoutVertexByAttr](#), [plotAllNetworks](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotCytoscapeGML](#), [plotNetwork](#), [plotPathClassifier](#)

Examples

```
## Prepare a weighted reaction network.
## Convert a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray))), bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=6)

## Plot paths.
plotPaths(ranked.p, rgraph)

## Convert paths to binary matrix, build a classifier.
ybinpaths <- pathsToBinary(ranked.p)
p.class <- pathClassifier(ybinpaths, target.class = "BCR/ABL", M = 3)

## Plotting with clusters, on a metabolic graph.
plotPaths(ranked.p, ex_sbml, path.clusters=p.class)
```

predictPathClassifier *Predicts new paths given a pathClassifier model.*

Description

Predicts new paths given a pathClassifier model.

Usage

```
predictPathClassifier(mix, newdata)
```

Arguments

mix	The result from pathClassifier .
newdata	A data.frame containing the new paths to be classified.

Value

A list with the following elements.

h	The posterior probabilities for each HME3M component.
posterior.probs	The posterior probabilities for HME3M model to classify the response.
label	A vector indicating the HME3M cluster membership.
component	The HME3M component membership for each pathway.
path.probabilities	The 3M path probabilities.
plr.probabilities	The PLR predictions for each component.

Author(s)

Timothy Hancock and Ichigaku Takigawa

See Also

Other Path clustering & classification methods: [pathClassifier](#), [pathCluster](#), [pathsToBinary](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotPathClassifier](#), [plotPathCluster](#), [predictPathCluster](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=6)
```

```
## Convert paths to binary matrix.
ybinpaths <- pathsToBinary(ranked.p)
p.class <- pathClassifier(ybinpaths, target.class = "BCR/ABL", M = 3)

## Just an example of how to predict cluster membership
pclass.pred <- predictPathCluster(p.class, ybinpaths$paths)
```

predictPathCluster *Predicts new paths given a pathCluster model*

Description

Predicts new paths given a pathCluster model.

Usage

```
predictPathCluster(pfit, newdata)
```

Arguments

pfit The pathway cluster model trained by [pathCluster](#) or [pathClassifier](#).
newdata The binary pathway dataset to be assigned a cluster label.

Value

A list with the following elements:

labels a vector indicating the 3M cluster membership.
posterior.probs a matrix of posterior probabilities for each path belonging to each cluster.

Author(s)

Ichigaku Takigawa
Timothy Hancock

See Also

Other Path clustering & classification methods: [pathClassifier](#), [pathCluster](#), [pathsToBinary](#), [plotClassifierROC](#), [plotClusterMatrix](#), [plotPathClassifier](#), [plotPathCluster](#), [predictPathClassifier](#)

Examples

```
## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot", bootstrap = FALSE)

## Get ranked paths using probabilistic shortest paths.
ranked.p <- pathRanker(rgraph, method="prob.shortest.path",
K=20, minPathSize=8)

## Convert paths to binary matrix.
ybinpaths <- pathsToBinary(ranked.p)
p.cluster <- pathCluster(ybinpaths, M=2)

## just an example of how to predict cluster membership.
pclust.pred <- predictPathCluster(p.cluster,ybinpaths$paths)
```

registerMemoryErr *Internal method to register memery errors.*

Description

Internal method to register memery errors, caused by compiled code. This method is used only by the package, and should not be invoked by users.

Usage

```
registerMemoryErr(method)
```

Arguments

method The method which generated the error.

Author(s)

Ahmed Mohamed

reindexNetwork	<i>Replaces current vertex ids with chosen attribute.</i>
----------------	---

Description

This function allows users to replace vertex ids with another attribute, calculating connectivities based on the new attribute.

Usage

```
reindexNetwork(graph, v.attr)
```

Arguments

graph	An annotated igraph object.
v.attr	Name of the attribute to use as vertex ids.

Details

This functions can be very useful when merging networks constructed from different databases. For example, to match a network created from Reactome to a KEGG network, you can reindex the vertices by "miriam.kegg.compound" attribute. Another usage is to remove duplicated vertices (in case of different subcellular compartments, for example). if a network has ATP_membrane & ATP_cytoplasm vertices, reindexing by chemical name will collapse them into one 'ATP' vertex.

Value

A new graph with vertices expanded.

Author(s)

Ahmed Mohamed

See Also

Other Network processing methods: [expandComplexes](#), [makeMetaboliteNetwork](#), [makeReactionNetwork](#), [rmSmallCompounds](#), [simplifyReactionNetwork](#), [vertexDeleteReconnect](#)

Examples

```
## Make a gene network from a reaction network.
data(ex_sbml) # A bipartite metbaolic network.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)
ggraph <- makeGeneNetwork(rgraph)

## Expand vertices into their contituent genes.
data(ex_kgml_sig) # Ras and chemokine signaling pathways in human
ggraph <- expandComplexes(ex_kgml_sig, v.attr = "miriam.ncbigene",
```

```
keep.parent.attr= c("^pathway", "^compartment"))

## Create a separate vertex for each compartment. This is useful in duplicating
## metabolite vertices in a network.
## Not run:
graph <- expandComplexes(graph, v.attr = "compartment",
  keep.parent.attr = "all",
  expansion.method = "duplicate",
  missing.method = "keep")

## End(Not run)
```

rmSmallCompounds *Remove ubiquitous compounds from a metabolic network*

Description

This function removes ubiquitous compounds (metabolites connected to numerous reactions) from a metabolic network. These compounds are reaction cofactors and currency compounds, such as ATP, CO₂, etc. A path through these metabolites may not be biologically meaningful. The default small compound list is derived from Reactome, containing kegg.compound, pubchem.compound, ChEBI and CAS identifiers.

Usage

```
rmSmallCompounds(graph, method = c("remove", "duplicate"),
  small.comp.ls = NPMdefaults("small.comp.ls"))
```

Arguments

graph	A metabolic network.
method	How to handle small compounds. Either simply delete these vertices "remove" (default), or make a separate vertex for each reaction they participate in "duplicate".
small.comp.ls	A list of small compounds to be used.

Value

A modified graph, with the small compounds removed or duplicated.

Author(s)

Ahmed Mohamed

See Also

Other Network processing methods: [expandComplexes](#), [makeMetaboliteNetwork](#), [makeReactionNetwork](#), [reindexNetwork](#), [simplifyReactionNetwork](#), [vertexDeleteReconnect](#)

Examples

```
data(ex_sbml)

sbml.removed <- rmSmallCompounds(ex_sbml, method="remove")
```

samplePaths	<i>Creates a set of sample path p-values for each length given a weighted network</i>
-------------	---

Description

Randomly traverses paths of increasing lengths within a set network to create an empirical pathway distribution for more accurate determination of path significance.

Usage

```
samplePaths(graph, max.path.length, num.samples = 1000,
  num.warmup = 10, verbose = TRUE)
```

Arguments

graph	A weighted igraph object. Weights must be in edge.weights or weight edge attributes.
max.path.length	The maximum path length.
num.samples	The number of paths to sample
num.warmup	The number of warm up paths to sample.
verbose	Whether to display the progress of the function.

Details

Can take a bit of time.

Value

A matrix where each row is a path length and each column is the number of paths sampled.

Author(s)

Timothy Hancock
Ahmed Mohamed

See Also

Other Path ranking methods: [extractPathNetwork](#), [getPathsAsEIDs](#), [pathRanker](#)

Examples

```

## Prepare a weighted reaction network.
## Conver a metabolic network to a reaction network.
data(ex_sbml) # bipartite metabolic network of Carbohydrate metabolism.
rgraph <- makeReactionNetwork(ex_sbml, simplify=TRUE)

## Assign edge weights based on Affymetrix attributes and microarray dataset.
# Calculate Pearson's correlation.
data(ex_microarray) # Part of ALL dataset.
rgraph <- assignEdgeWeights(microarray = ex_microarray, graph = rgraph,
weight.method = "cor", use.attr="miriam.uniprot",
y=factor(colnames(ex_microarray)), bootstrap = FALSE)

## Get significantly correlated paths using "p-value" method.
## First, establish path score distribution by calling "samplePaths"
pathsample <- samplePaths(rgraph, max.path.length=10,
num.samples=100, num.warmup=10)

## Get all significant paths with p<0.1
significant.p <- pathRanker(rgraph, method = "pvalue",
sampledpaths = pathsample ,alpha=0.1)

```

SBML2igraph

Processes SBML files into igraph objects

Description

This function takes SBML files as input, and returns either a metabolic or a signaling network as output.

Usage

```

SBML2igraph(filename, parse.as = c("metabolic", "signaling"),
miriam.attr = "all", gene.attr, expand.complexes, verbose = TRUE)

```

Arguments

filename	A character vector containing the SBML files to be processed. If a directory path is provided, all *.xml and *.sbml files in it and its subdirectories are included.
parse.as	Whether to process file into a metabolic or a signaling network.
miriam.attr	A list of annotation attributes to be extracted. If "all", then all attributes written in MIRIAM guidelines (see Details) are extracted (Default). If "none", then no attributes are extracted. Otherwise, only attributes matching those specified are extracted.
gene.attr	An attribute to distinguish species representing genes from those representing small molecules (see Details). Ignored if parse.as="metabolic".

expand.complexes	Split protein complexes into individual gene nodes. Ignored if <code>parse.as="metabolic"</code> , or when <code>gene.attr</code> is not provided.
verbose	Whether to display the progress of the function.

Details

Users can specify whether files are processes as metabolic or signaling networks.

Metabolic networks are given as bipartite graphs, where metabolites and reactions represent vertex types. This is constructed from `ListOfReactions` in SBML file, connecting them to their corresponding substrates and products (`ListOfSpecies`). Each reaction vertex has `genes` attribute, listing all modifiers of this reaction. As a general rule, reactions inherit all annotation attributes of its catalyzing genes.

Signaling network have genes as vertices and edges represent interactions. Since SBML format may represent signaling events as reaction, all species are assumed to be genes (rather than small molecules). For a simple path $S_0 \rightarrow R_1 \rightarrow S_1$, in signaling network, the path will be $S_0 \rightarrow M(R_1) \rightarrow S_1$ where $M(R_1)$ is R_1 modifier(s). To distinguish gene species from small molecules, user can provide `gene.attr` (for example: `miriam.uniprot` or `miriam.ncbigene`) where only annotated species are considered genes.

All annotation attributes written according to MIRIAM guidelines (either `urn:miriam:xxx:xxx` or `http://identifiers.org/xxx/xxx`) are extracted by default. Non-conforming attributes can be extracted by specifying `miriam.attr`.

To generate a genome scale network, simply provide a list of files to be parsed, or put all file in a directory, as pass the directory path as filename

Note: This function requires libSBML installed (Please see the installation instructions in the Vignette). Some SBML level-3 files may requires additional libraries also (An informative error will be displayed when parsing such files). Please visit http://sbml.org/Documents/Specifications/SBML_Level_3/Packages for more information.

Value

An igraph object, representing a metabolic or a signaling network.

Author(s)

Ahmed Mohamed

See Also

Other Database extraction methods: [KGML2igraph](#), [biopax2igraph](#)

Examples

```
if(is.loaded("readsbmfile")){ # This is false if libSBML wasn't available at installation.
  filename <- system.file("extdata", "porphyrin.sbml", package="NetPathMiner")

  # Process SBML file as a metabolic network
  g <- SBML2igraph(filename)
```

```
plotNetwork(g)

# Process SBML file as a signaling network
g <- SBML2igraph(filename, parse.as="signaling",
                 gene.attr="miriam.uniprot", expand.complexes=TRUE)
dev.new()
plotNetwork(g)
}
```

simplifyReactionNetwork

Removes reactions with no gene annotations

Description

This function removes reaction vertices with no gene annotations as indicated by the parameter `gene.attr`, and connect their neighbour vertices to preserve graph connectivity. This is particularly meaningful when reactions are translocation or spontaneous reactions, which are not catalysed by genes.

Usage

```
simplifyReactionNetwork(reaction.graph, gene.attr = "genes",
                        remove.missing.genes = TRUE,
                        reconnect.threshold = vcount(reaction.graph))
```

Arguments

`reaction.graph` A reaction network.

`gene.attr` The attribute to be considered as "genes". Reactions missing this annotation, will be removed.

`remove.missing.genes`
If FALSE, only translocation and spontaneous reactions are removed, otherwise all reactions with no gene annotations are removed.

`reconnect.threshold`
An argument passed to [vertexDeleteReconnect](#)

Value

A simplified reaction network.

Author(s)

Ahmed Mohamed

See Also

Other Network processing methods: [expandComplexes](#), [makeMetaboliteNetwork](#), [makeReactionNetwork](#), [reindexNetwork](#), [rmSmallCompounds](#), [vertexDeleteReconnect](#)

Examples

```

data(ex_sbml)
rgraph <- makeReactionNetwork(ex_sbml, simplify=FALSE)

## Removes all reaction nodes with no annotated genes.
rgraph <- simplifyReactionNetwork(rgraph, remove.missing.genes=TRUE)

```

stdAttrNames

MIRIAM annotation attributes

Description

These functions deals with conforming with MIRIAM annotation guidelines, conversion and mapping between MIRIAM identifiers.

Usage

```

stdAttrNames(graph, return.value = c("matches", "graph"))

fetchAttribute(graph, organism = "Homo sapiens", target.attr,
  source.attr, bridge.web = NPMdefaults("bridge.web"))

```

Arguments

graph	An annotated igraph object.
return.value	Specify whether to return the names of matched standard annotations, or modify the graph attribute names to match the standards.
organism	The latin name of the organism (Case-sensitive).
target.attr	The target annotation, given as MIRIAM standard in the format miriam.xxx
source.attr	The source annotation attribute from graph
bridge.web	The base URL for Brigde Database webservices.

Value

For stdAttrNames, matches gives the original attribute names and their MIRIAM version. Since this is done by simple text matching, mismatches may occur for ambiguous annotations (such as GO, EC number). graph returns the input graph with attribute names standardized.

For fetchAttribute, the input graph with the fetched attribute mapped to vertices.

Author(s)

Ahmed Mohamed

See Also

Other Attribute handling methods: [getAttrStatus](#)

Examples

```
data(ex_kgml_sig) # Ras and chemokine signaling pathways in human
## Modify attribute names to match MIRIAM standard annotations.
graph <- stdAttrNames(ex_kgml_sig, "graph")

# Use Attribute fetcher to get affymetrix probeset IDs for network vertices.
## Not run:
graph <- fetchAttribute(graph, organism="Homo sapiens",
                        target.attr="miriam.affy.probeset")

## End(Not run)
```

toGraphNEL

Converts an annotated igraph object to graphNEL

Description

Converts an annotated igraph object to graphNEL

Usage

```
toGraphNEL(graph, export.attr = "")
```

Arguments

graph	An annotated igraph object..
export.attr	A regex expression representing vertex attributes to be exported to the new graphNEL object. Supplying an empty string "" (default) will export all attributes.

Value

A graphNEL object.

Author(s)

Ahmed Mohamed

Examples

```
data(ex_kgml_sig) # Ras and chemokine signaling pathways in human
graphNEL <- toGraphNEL(ex_kgml_sig, export.attr="^miriam.")
```

vertexDeleteReconnect *Network editing: removing vertices and connecting their neighbours*

Description

This function removes vertices given as `vids` and connects their neighbours as long as the shortest path between the neighbours are below the `reconnect.threshold`.

Usage

```
vertexDeleteReconnect(graph, vids, reconnect.threshold = vcount(graph),  
  copy.attr = NULL)
```

Arguments

<code>graph</code>	A reaction network.
<code>vids</code>	Vertex ids to be removed.
<code>reconnect.threshold</code>	If the shortest path between vertices is larger than this threshold, they are not reconnected.
<code>copy.attr</code>	A function, or a list of functions, combine edge attributes. Edge attributes of new edges (between reconnected neighbours) are obtained by combining original edges attributes along the shortest path between reconnected neighbors.

Value

A modified graph.

Author(s)

Ahmed Mohamed

See Also

Other Network processing methods: [expandComplexes](#), [makeMetaboliteNetwork](#), [makeReactionNetwork](#), [reindexNetwork](#), [rmSmallCompounds](#), [simplifyReactionNetwork](#)

Examples

```
## Remove all reaction vertices from a bipartite metabolic network  
## keeping only metabolite vertices.  
data(ex_sbml)  
graph <- vertexDeleteReconnect(ex_sbml, vids=which(V(ex_sbml)$reactions))
```

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