Package ‘InTAD’

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Type   Package
Title  Search for correlation between epigenetic signals and gene expression in TADs
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Description The package is focused on the detection of correlation between expressed genes and selected epigenomic signals (i.e. enhancers obtained from ChIP-seq data) either within topologically associated domains (TADs) or between chromatin contact loop anchors. Various parameters can be controlled to investigate the influence of external factors and visualization plots are available for each analysis step.
License GPL (>=2)
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combineInTAD

Preparation for correlation analysis

Description

This function combines signals and genes in inside of Topologically Associated Domains (TADs)

Usage

combineInTAD(object, tadGR, selMaxTadOvlp = TRUE, closestGene = TRUE)

Arguments

- object: InTADSig object
- tadGR: TAD genomic regions
- selMaxTadOvlp: If a signal overlaps 2 or more TADs by default only single TAD with max overlap is selected. All overlaps can be included by deactivating this option.
- closestGene: By default closest to TAD genes are selected based on TSS location. Deactivate this option to use genes only lying within TAD.
**combineWithLoops**

**Description**

This function combines signals and genes based on the usage of loops obtained from HiC data analysis.

**Usage**

`combineWithLoops(object, loopsInitDf, fragmentLength = 0, tssWidth = 2000, extSize = 0)`

**Arguments**

- **object**: InTADSig object
- **loopsInitDf**: Data frame with loops. By default 6-column format (chr1,start1,end1,chr2,start2,pos2) is expected.
- **fragmentLength**: In case the input format is 4-column (chr1,middlePos1, chr2, middlePos2) fragment length should be provided to extend the corresponding loci for loop start and end positions.
- **tssWidth**: The transcription start site width is used to control overlaps with loop anchor. Default is 2000 base pairs.
- **extSize**: The loop endings can be extended upstream and downstream with provided corresponding increase size in base pairs.
Details

The expected input is the loops data.frame applied to find connections of signals to genes. This data.frame could be in two formats: either (chr1,start1,end1,chr2,start2,end2) or (chr1,middlePos1,chr2,middlePos2) with fragment size.

Value

Updated InTADSig object containing genes connected to signals via loops

<table>
<thead>
<tr>
<th>enhSel</th>
<th>Enhancer signals subset detected from medulloblastoma samples</th>
</tr>
</thead>
</table>

Description

This data.frame contains 65 selected in chr15 normalized enhancers signals subset from 25 medulloblastoma samples.

Usage

enhSel

Format

a data.frame instance

Value

NULL, but makes available the dataframe

<table>
<thead>
<tr>
<th>enhSelGR</th>
<th>Genomic coordinates of enhancer signals subset</th>
</tr>
</thead>
</table>

Description

This GRanges object contains the coordinates of 65 medulloblastoma enhancer signals in chr15 target region

Usage

enhSelGR

Format

a GRanges object

Value

NULL, but makes available the dataset
**exprs.InTADSig-method**

*Gene expression counts table*

**Description**

This function returns gene expression counts table

**Usage**

```r
## S4 method for signature 'InTADSig'
exprs(object)
```

**Arguments**

- `object`:
  - InTADSig object with signals and genes

**Value**

Gene expression table

**Examples**

```r
inTadSig <- newSigInTAD(enhSel, enhSelGR, rpkmCountsSel, txsSel)
head(exprs(inTadSig))
```

---

**filterGeneExpr**

*Function to filter gene expression*

**Description**

This function performs filtering of gene expression counts based on various parameters

**Usage**

```r
filterGeneExpr(obj, cutVal = 0, geneType = NA, checkExprDistr = FALSE, plotExprDistr = FALSE)
```

**Arguments**

- `obj`:
  - InTADSig object
- `cutVal`:
  - Exclude genes that have max expression less or equal to this value in all samples. Default: 0
- `geneType`:
  - Type of gene to select for filtering i.e. "protein_coding". Default: NA
- `checkExprDistr`:
  - Adjust cutVal based on gene expression distribution
- `plotExprDistr`:
  - Perform visualization of the distribution
**findCorFromLoops**

*Function to perform correlation analysis via loops.*

**Details**

The function allows to stabilize the functional activity of the genes. By default all not expressed genes are filtered. It is also possible to set type of gene to take into account i.e. "protein_coding" only. This option requires additional metadata column "transcript_type". Also, special filtering option based on mclust library allows to analyze distribution of counts and adjust the cut value to exclude low expressed genes.

**Value**

InTADSig object with filtered counts table

**Examples**

```r
## perform analysis on test data
inTadSig <- newSigInTAD(enhSel, enhSelGR, rpkmCountsSel, txsSel)
## default filtering
inTadSig <- filterGeneExpr(inTadSig)
## filter based on gene type
inTadSig <- filterGeneExpr(inTadSig, geneType = "protein_coding")
```

---

**Description**

This function combines genes and signals using obtained loop connections.

**Usage**

```r
findCorFromLoops(object, method = "pearson", adj.pval = FALSE)
```

**Arguments**

- `object`: InTADSig object with signals and genes combined via loops
- `method`: Correlation method: "pearson" (default), "kendall", "spearman"
- `adj.pval`: Perform p-value adjustment and include q-values in result

**Value**

A table with correlation values for signal-gene pairs including correlation p-value and euclidian distance.
findCorrelation

Function to perform correlation analysis in TADs

Description

This function combines genes and signals in inside of TADs

Usage

```r
findCorrelation(object, method = "pearson", adj.pval = FALSE,
               plot.proportions = FALSE)
```

Arguments

- `object`: InTADSig object with signals and genes combined in TADs
- `method`: Correlation method: "pearson" (default), "kendall", "spearman"
- `adj.pval`: Perform p-value adjustment and include q-values in result
- `plot.proportions`: Plot proportions of signals and genes in correlation

Value

A table with correlation values for signal-gene pairs including correlation p-value, euclidian distance and rank.

Examples

```r
## perform analysis on test data
inTadSig <- newSigInTAD(enhSel, enhSelGR, rpkmCountsSel, txsSel)
inTadSig <- filterGeneExpr(inTadSig, geneType = "protein_coding")
inTadSig <- combineInTAD(inTadSig, tadGR)
corData <- findCorrelation(inTadSig, method="pearson")
```

fnSE

Preparation for correlation analysis for a signal

Description

This function collects all genes for signal genomic region inside of Topologically Associated Domains (TADs)

Usage

```r
fnSE(id, sigList, tadGR, tss, pickMaxOvlp, nearestTad)
```
Arguments

- id: Id of signal from the list
- sigList: List of signal GRs and their names
- tadGR: TAD genomic regions
- tss: Gene transcription start sites
- pickMaxOvlp: Use TAD with max overlap
- nearestTad: The table listing TADs nearest to each TSS

Details

The signal is checked if it is lying inside of TAD. Then all genes in this TAD are collected.

Value

Data.frame containing genes connected to signal

geneCoords <- function(object)
  {  # S4 method for signature 'InTADSig'
    geneCoords(object)
  }

Arguments

- object: InTADSig object with signals and genes

Value

Gene GRanges

Examples

inTadSig <- newSigInTAD(enhSel, enhSelGR, rpkmCountsSel, txsSel)
head(geneCoords(inTadSig))
**get.enr.bg.normfit**

*Function to estimate gene expression*

---

**Description**

This function uses mclust package to analyze gene expression distribution.

**Usage**

```r
get.enr.bg.normfit(x)
```

**Arguments**

- `x` Full gene expression vector

**Details**

The function adjusts filtering cut value based on mclust library to exclude low-expressed genes. It is a part of filtering procedure.

**Value**

Distribution properties: mean and std

---

**InTADSig**

*The InTADSig Class*

---

**Description**

The InTADSig object stores signals and gene expression data for the samples.

**Details**

It uses MultiAssayExperiment object to store information. Key slots to access are listed below.

**Slots**

- `sigMAE`: "MultiAssayExperiment", MultiAssayExperiment object containing signals and gene counts
- `signalConnections`: "list", The list of signals representing gene data frames in the same TAD
- `loopsDf`: "data.frame", The data.frame containing details of provided input loops
- `loopConnections`: "list", The list of connections between signals and genes via loops
- `ncore`: "numeric", Number of cores to use for parallel computing"
loadSigInTAD

Load InTADSig object from text files

Description

The function loads the data tables to create an object that contains the signals and gene expression data frames along with their genomic coordinates for further processing.

Usage

loadSigInTAD(signalsFile, countsFile, gtfFile, annFile = "", performLog = TRUE, logExprsOffset = 1, ncores = 1)

Arguments

- `signalsFile`: Tab-separated data table containing signals and their coordinates as row names.
- `countsFile`: Tab-separated counts table.
- `gtfFile`: GTF file containing all gene coordinates.
- `annFile`: Tab-delimited phenotype annotation of samples.
- `performLog`: Perform log2 conversion of expression values. Default: TRUE.
- `logExprsOffset`: Offset x for log2 gene expression i.e. log2(value + x). Default: 1
- `ncores`: Number of cores to use for parallel computing.

Details

The function loads data from input files and creates an object that stores matrices of signals and gene expression values along with coordinates. The samples order and names of columns should match in both tables. It is expected that gene ids are applied in the validation of counts table.

Value

Novel InTADSig object

Examples

# create sigInTAD object
inTadSig <- newSigInTAD(enhSel, enhSelGR, rpkmCountsSel, txsSel)
**loopsDfSel**

Data frame containing coordinates of loops

**Description**

The table contains genomic coordinates of chromatin loops in 6-column format derived from IMR90 cell line (focus : chr15)

**Usage**

loopsDfSel

**Format**

a data.frame object

**Value**

NULL, but makes available the dataset

---

**mbAnnData**

Data frame containing information about samples

**Description**

The table includes additional information about MB tumour samples (subgroup, gender, age, histology and M.Stage)

**Usage**

mbAnnData

**Format**

a data.frame object

**Value**

NULL, but makes available the dataset
newSigInTAD

Create InTADSig object

Description

The function generates an object that contains the signals and gene expression dataframes along with their genomic coordinates for further processing.

Usage

newSigInTAD(signalData = NULL, signalRegions = NULL, countsData = NULL, geneRegions = NULL, sampleInfo = NULL, performLog = TRUE, logExprsOffset = 1, ncores = 1)

Arguments

- signalData: data frame containing signals
- signalRegions: genomic regions of the signals
- countsData: data matrix containing count expression values
- geneRegions: gene coordiantes
- sampleInfo: data frame containing additional sample info
- performLog: Perform log2 conversion of expression values. Default: TRUE.
- logExprsOffset: Offset x for log2 gene expression i.e. log2(value + x). Default: 1
- ncores: Number of cores to use for parallel computing

Details

InTADSig object stores matrices of signals and gene expression values along with coordinates. The order of samples and names of columns should match in both datasets. For gene coordinates GRanges "gene_id" and "gene_name" are required in metadata. These are typical markers of genes in GTF annotation format.

Value

Novel InTADSig object

Examples

```r
## create sigInTAD object
inTadSig <- newSigInTAD(enhSel, enhSelGR, rpkmCountsSel, txsSel)
```
plotCorAcrossRef  

Function to plot correlation across genome

Description

This function creates a plot of correlation strength in target genomic region from the result table. The X-coordinates represent signals, Y-coords represent genes, while each dot represents -log10(P-value) from correlation test. Additionally all TAD boundaries can be visualized.

Usage

plotCorAcrossRef(obj, corRes, targetRegion, showCorVals = FALSE, symmetric = FALSE, tads = NULL)

Arguments

- **obj**: InTADSig object with signals and genes combined in TADS
- **corRes**: Correlation result table created by function findCorrelation()
- **targetRegion**: Target genomic region visualise.
- **showCorVals**: Use this option to visualize postive correlation values instead of correlation strength
- **symmetric**: Activate mirror symmetry for gene-signal connections
- **tads**: TAD regions to visualize. By default only TADs present in correlation result table are applied (NULL value).

Value

A ggplot object for visualization or customization.

Examples

```r
inTadSig <- newSigInTAD(enhSel, enhSelGR, rpkmCountsSel, txsSel)
inTadSig <- combineInTAD(inTadSig, tadGR)
corData <- findCorrelation(inTadSig, method="pearson")
plotCorAcrossRef(inTadSig,corData,GRanges("chr15:25000000-28000000"))
```
plotCorrelation

Function to plot correlation

Description

This function creates a plot of selected pair signal-gene

Usage

plotCorrelation(obj, sId, geneName, xLabel = "Gene expression", yLabel = "Signal enrichment", colByPhenotype = "", corMethod = "pearson")

Arguments

obj InTADSig object with signals and genes combined in TADS
sId Signal id based on genomic coordinates i.e. "chr:start-end"
geneName Gene name to select. Based on "gene_name" attribute.
xLabel The label to mark signal X-axis. Default: "Gene expression"
yLabel The label to mark signal Y-axis. Default: "Signal enrichment"
colByPhenotype The pheno data column i.e. tumour type that can be use for colour
corMethod Correlation method. Default: Pearson

Value

A ggplot object for visualization or customization.

Examples

inTadSig <- newSigInTAD(enhSel, enhSelGR, rpkmCountsSel, txsSel)
inTadSig <- combineInTAD(inTadSig, tadGR)
plotCorrelation(inTadSig, "chr15:26372163-26398073", "GABRA5")

rpkmCountsSel

Gene expression subset from medulloblastoma samples

Description

This data.frame contains RPKM gene expression values from chr15 for subset from 25 medulloblastoma samples.

Usage

rpkmCountsSel
**sigCoords**

**Format**
- a data.frame instance

**Value**
- NULL, but makes available the dataframe

---

<table>
<thead>
<tr>
<th>sigCoords</th>
<th>Signal coords GRanges</th>
</tr>
</thead>
</table>

**Description**
- This function returns the signal GRanges

**Usage**

```
sigCoords(object)
```

```mermaid
## S4 method for signature 'InTADSig'
sigCoords(object)
```

**Arguments**
- object: InTADSig object with signals and genes

**Value**
- Signal GRanges

**Examples**

```r
inTadSig <- newSigInTAD(enhSel, enhSelGR, rpkmCountsSel, txsSel)
head(sigCoords(inTadSig))
```

---

<table>
<thead>
<tr>
<th>signals</th>
<th>Signal values table</th>
</tr>
</thead>
</table>

**Description**
- This function returns the signal values table

**Usage**

```
signals(object)
```

```mermaid
## S4 method for signature 'InTADSig'
signals(object)
```
Arguments

object | InTADSig object with signals and genes

Value

Signals table

Examples

```r
inTadSig <- newSigInTAD(enhSel, enhSelGR, rpkmCountsSel, txsSel)
head(signals(inTadSig))
```

<table>
<thead>
<tr>
<th>tadGR</th>
<th>Genomic coordinates of topologically associated domains</th>
</tr>
</thead>
</table>

Description

This GRanges object contains the coordinates of TADs revealed from IMR90 cell line (extracted from 0-indexed .bed file)

Usage

```r
tadGR
```

Format

a GRanges object

Value

NULL, but makes available the dataset

<table>
<thead>
<tr>
<th>txsSel</th>
<th>Genomic coordinates of genes subset</th>
</tr>
</thead>
</table>

Description

This GRanges object contains the coordinates of genes subset from chr15

Usage

```r
txsSel
```

Format

a GRanges object
**Value**

NULL, but makes available the dataset
Index

combineInTAD, 2
combineWithLoops, 3

enhSel, 4
enhSelGR, 4
exprs, InTADSig-method, 5

filterGeneExpr, 5
findCorFromLoops, 6
findCorrelation, 7
fnSE, 7

geneCoords, 8
geneCoords, InTADSig-method
geneCoords, InTADSig-method (geneCoords), 8
get.enr.bg.normfit, 9

InTADSig, 9
InTADSig-class (InTADSig), 9

loadSigInTAD, 10
loopsDFSel, 11

mbAnnData, 11

newSigInTAD, 12

plotCorAcrossRef, 13
plotCorrelation, 14

rpmCountsSel, 14

sigCoords, 15
sigCoords, InTADSig-method (sigCoords), 15
signals, 15
signals, InTADSig-method (signals), 15

tadGR, 16
txsSel, 16