Package ‘DaMiRseq’

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Description The DaMiRseq package offers a tidy pipeline of data mining procedures to identify transcriptional biomarkers and exploit them for both binary and multi-class classification purposes. The package accepts any kind of data presented as a table of raw counts and allows including both continuous and factorial variables that occur with the experimental setting. A series of functions enable the user to clean up the data by filtering genomic features and samples, to adjust data by identifying and removing the unwanted source of variation (i.e. batches and confounding factors) and to select the best predictors for modeling. Finally, a ``stacking'' ensemble learning technique is applied to build a robust classification model. Every step includes a checkpoint that the user may exploit to assess the effects of data management by looking at diagnostic plots, such as clustering and heatmaps, RLE boxplots, MDS or correlation plot.

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Description

This is a helper function to easily draw (1) clustering dendrogram and heatmap of a sample-per-sample correlation matrix, (2) multidimensional scaling plots (MDS), (3) relative log expression (RLE) boxplots of expression data, (4) a sample-by-sample expression value distribution, and (5) a class average expression value distribution.

Usage

```
DaMiR.Allplot(
  data,
  df,
  type = c("spearman", "pearson"),
  what = c("all", "all_w_PCA", "MDS", "PCA", "heatmap", "RLEbox", "distr", "avg_distr")
)
```

Arguments

data A SummarizedExperiment object or a matrix or a data.frame where rows and cols should be, respectively, observations and features.
df A data frame with class and known variables (or a subset of them); at least one column with 'class' label must be included.
type A character string specifying the metric to be applied to correlation analysis. Either "spearman" or "pearson" is allowed; default is "spearman".
what A character string specifying the plots to be shown 'all', 'all_w_PCA', 'MDS', 'PCA', 'heatmap', 'RLEbox', 'distr', 'avg_distr' are allowed; default is "all".

Details

Please be sure that NAs are not present in df’s columns. Plots will not be drawn in the presence of NAs.

Value

A dendrogram and heatmap, MDS plot(s), a RLE boxplot, a sample-by-sample expression value distribution, and a class average expression value distribution.

Author(s)

Mattia Chiesa, Luca Piacentini
Examples

# use example data:
data(data_norm)
data(df)
# Draw clustering dendrogram and heatmap, MDS, RLE boxplot:
DaMiR.Allplot(data=data_norm, df=df[,5,drop=FALSE])

---

### DaMiR.Clustplot
**Expression data clustering and heatmap**

**Description**

The function helps to draw a clustering dendrogram and a heatmap of expression data.

**Usage**

```r
DaMiR.Clustplot(
data, 
df, 
type_row = c("euclidean", "correlation"), 
type_col = c("euclidean", "correlation")
)
```

**Arguments**

- **data**
  A SummarizedExperiment object or a matrix or a data.frame where rows and cols should be, respectively, observations and features

- **df**
  A data frame with class and (optionally) known variables; at least one column with 'class' label must be included

- **type_row**
  The metric to be used to cluster rows. Either "euclidean" or "correlation" is allowed; default is "euclidean"

- **type_col**
  The metric to be used to cluster cols. Either "euclidean" or "correlation" is allowed; default is "euclidean"

**Value**

A clustering dendrogram and heatmap.

**Author(s)**

Mattia Chiesa, Luca Piacentini
DaMiR.corrplot

Examples

# use example data:
data(data_norm)
data(df)
# use the first 100 genes:
data_norm_red<-data_norm[1:100,]
# Draw heatmap: samples (cols) per genes (rows)
# and use variable annotation:
DaMiR.Clustplot(data=data_norm_red,
df=df, type_row="correlation", type_col="correlation")

DaMiR.corrplot  Correlation Plot

Description

This function easily draws the correlation plot of surrogate variables (sv) and variables.

Usage

DaMiR.corrplot(sv, df, type = c("pearson", "spearman"), sig.level = 0.01)

Arguments

sv
The matrix of sv identified by DaMiR.SV function

df
A data frame with class and known variables; at least one column with 'class' label must be included

type
Type of correlation metric to be applied; default is "pearson"
sig.level
The significance level of the correlation; default is 0.0001

Details

Factorial variables are allowed. They will be tranformed as numeric before applying the rcorr function of Hmisc. The corrplot function, which draws the plot, marks with a cross all the correlations that do not reach the significance threshold defined in the sig.level argument. This plot allows the user to identify those sv that present significant correlations with either technical and biological known variables. Notably, none of the sv should present significant correlation with "class" variable.

Value

A correlation plot between sv and known variables.

Author(s)

Mattia Chiesa, Luca Piacentini
Namely:

**DaMiR.EnsembleLearning**

Build Classifier using 'Stacking' Ensemble Learning strategy.

**Description**

This function implements a 'Stacking' ensemble learning strategy. Users can provide heterogeneous features (other than genomic features) which will be taken into account during classification model building.

**Usage**

```
DaMiR.EnsembleLearning(
  data, classes, variables, fSample.tr = 0.7, fSample.tr.w = 0.7, iter = 100,
  cl_type = c("RF", "kNN", "SVM", "LDA", "LR", "NB", "NN", "PLS")
)
```

**Arguments**

- **data**: A transposed data frame of normalized expression data. Rows and Cols should be, respectively, observations and features.
- **classes**: A class vector with `nrow(data)` elements. Each element represents the class label for each observation. More than two different class labels are handled.
- **variables**: An optional data frame containing other variables (but without 'class' column). Each column represents a different covariate to be considered in the model.
- **fSample.tr**: Fraction of samples to be used as training set; default is 0.7.
- **fSample.tr.w**: Fraction of samples of training set to be used during weight estimation; default is 0.7.
- **iter**: Number of iterations to assess classification accuracy; default is 100.

**Examples**

```r
# use example data:
data(df)
data(sv)
# Draw correlation plot:
# DaMiR.corrplot(sv=sv, df=df, type = "pearson", sig.level=0.01)
```
cl_type

List of weak classifiers that will compose the meta-learners. Only "RF", "kNN", "SVM", "LDA", "LR", "NB", "NN", "PLS" are allowed. Default is c("RF", "LR", "kNN", "LDA", "NB", "SVM")

Details

To assess the robustness of a set of predictors, a specific 'Stacking' strategy has been implemented. First, a training set (TR1) and a test set (TS1) are generated by 'bootstrap' sampling. Then, sampling again from TR1 subset, another pair of training (TR2) and test set (TS2) are obtained. TR2 is used to train Random Forest (RF), Naive Bayes (NB), Support Vector Machines (SVM), k-Nearest Neighbour (kNN), Linear Discriminant Analysis (LDA) and Logistic Regression (LR) classifiers, whereas TS2 is used to test their accuracy and to calculate weights. The decision rule of 'Stacking' classifier is made by a linear combination of the product between weights (w) and predictions (Pr) of each classifier; for each sample k, the prediction is computed by:

$$P_{k, \text{Ensemble}} = w_{k, RF} \times Pr_{k, RF} + w_{k, NB} \times Pr_{k, NB} + w_{k, SVM} \times Pr_{k, SVM} + w_{k, kNN} \times Pr_{k, kNN} + w_{k, LDA} \times Pr_{k, LDA} + w_{k, LR} \times Pr_{k, LR}$$

$$P_{k, \text{Ensemble}} = \sum (w[RF] \times Pr[k, i]), i = 1, N$$

Performance of 'Stacking' classifier is evaluated by using TS1. This process is repeated several times (default 100 times).

Value

A list containing:

- A matrix of accuracies of each classifier in each iteration.
- A matrix of weights used for each classifier in each iteration.
- A list of all models generated in each iteration.
- A violin plot of model accuracy obtained for each iteration.

Author(s)

Mattia Chiesa, Luca Piacentini

Examples

```r
# use example data:
data(selected_features)
data(df)
set.seed(1)
# only for the example:
# speed up the process setting a low 'iter' argument value;
# for real data set use default 'iter' value (i.e. 100) or higher:
# Classification_res <- DaMiR.EnsembleLearning(selected_features,
# classes=df$class, fSample.tr=0.6, fSample.tr.w=0.6, iter=3,
# cl_type=c("RF","kNN"))
```
DaMiR.EnsembleLearning2cl

Build a Binary Classifier using 'Stacking' Learning strategy.

Description

This function implements a 'Stacking' ensemble learning strategy. Users can provide heterogeneous features (other than genomic features) which will be taken into account during classification model building. A 'two-classes' classification task is addressed.

Usage

DaMiR.EnsembleLearning2cl(
  data,
  classes,
  variables,
  fSample.tr = 0.7,
  fSample.tr.w = 0.7,
  iter = 100,
  cl_type = c("RF", "kNN", "SVM", "LDA", "LR", "NB", "NN", "PLS")
)

Arguments

data A transposed data frame of normalized expression data. Rows and Cols should be, respectively, observations and features

classes A class vector with nrow(data) elements. Each element represents the class label for each observation. Two different class labels are allowed

variables An optional data frame containing other variables (but without 'class' column). Each column represents a different covariate to be considered in the model

fSample.tr Fraction of samples to be used as training set; default is 0.7

fSample.tr.w Fraction of samples of training set to be used during weight estimation; default is 0.7

iter Number of iterations to assess classification accuracy; default is 100

cl_type List of weak classifiers that will compose the meta-learners. "RF", "kNN", "SVM", "LDA", "LR", "NB", "NN", "PLS" are allowed. Default is c("RF", "LR", "kNN", "LDA", "NB", "SVM")

Details

To assess the robustness of a set of predictors, a specific 'Stacking' strategy has been implemented. First, a training set (TR1) and a test set (TS1) are generated by 'bootstrap' sampling. Then, sampling again from TR1 subset, another pair of training (TR2) and test set (TS2) are obtained. TR2 is used to train Random Forest (RF), Naive Bayes (NB), Support Vector Machines (SVM), k-Nearest Neighbour (kNN), Linear Discriminant Analysis (LDA) and Logistic Regression (LR) classifiers,
whereas TS2 is used to test their accuracy and to calculate weights. The decision rule of 'Stacking' classifier is made by a linear combination of the product between weights (w) and predictions (Pr) of each classifier; for each sample k, the prediction is computed by:

\[
P_{k,\text{Ensemble}} = w_{RF} \times Pr_{k,RF} + w_{NB} \times Pr_{k,NB} + w_{SVM} \times Pr_{k,SVM} + w_{kNN} \times Pr_{k,kNN} + w_{LDA} \times Pr_{k,LDA} + w_{LR} \times Pr_{k,LR}
\]

Performance of 'Stacking' classifier is evaluated by using TS1. This process is repeated several times (default 100 times).

Value

A list containing:

- A matrix of accuracies of each classifier in each iteration.
- A matrix of weights used for each classifier in each iteration.
- A list of all models generated in each iteration.
- A violin plot of model accuracy obtained for each iteration.

Author(s)

Mattia Chiesa, Luca Piacentini

Examples

```r
# use example data:
data(selected_features)
data(df)
set.seed(1)
# only for the example:
# speed up the process setting a low 'iter' argument value;
# for real data set use default 'iter' value (i.e. 100) or higher:
# Classification_res <- DaMiR.EnsembleLearning(selected_features,
# classes=df$class, fSample.tr=0.6, fSample.tr.w=0.6, iter=3,
# cl_type=c("RF","kNN"))
```

---

**DaMiR.EnsembleLearningNcl**

*Build a Multi-Class Classifier using 'Stacking' Learning strategy.*

**Description**

This function implements a 'Stacking' ensemble learning strategy. Users can provide heterogeneous features (other than genomic features) which will be taken into account during classification model building. A 'multi-classes' classification task is addressed.
Usage

DaMiR.EnsembleLearningNcl(
  data,
  classes,
  variables,
  fSample.tr = 0.7,
  fSample.tr.w = 0.7,
  iter = 100,
  cl_type = c("RF", "kNN", "SVM", "LDA", "LR", "NB", "NN", "PLS")
)

Arguments

data A transposed data frame of normalized expression data. Rows and Cols should be, respectively, observations and features

classes A class vector with nrow(data) elements. Each element represents the class label for each observation. More than two different class labels are allowed

variables An optional data frame containing other variables (but without 'class' column). Each column represents a different covariate to be considered in the model

fSample.tr Fraction of samples to be used as training set; default is 0.7

fSample.tr.w Fraction of samples of training set to be used during weight estimation; default is 0.7

iter Number of iterations to assess classification accuracy; default is 100

cl_type List of weak classifiers that will compose the meta-learners. "RF", "kNN", "SVM", "LDA", "LR", "NB", "NN", "PLS" are allowed. Default is c("RF", "LR", "kNN", "LDA", "NB", "SVM")

Details

To assess the robustness of a set of predictors, a specific ‘Stacking’ strategy has been implemented. First, a training set (TR1) and a test set (TS1) are generated by 'bootstrap' sampling. Then, sampling again from TR1 subset, another pair of training (TR2) and test set (TS2) are obtained. TR2 is used to train Random Forest (RF), Naive Bayes (NB), Support Vector Machines (SVM), k-Nearest Neighbour (kNN), Linear Discriminant Analysis (LDA) and Logistic Regression (LR) classifiers, whereas TS2 is used to test their accuracy and to calculate weights. The decision rule of 'Stacking' classifier is made by a linear combination of the product between weights (w) and predictions (Pr) of each classifier; for each sample k, the prediction is computed by:

\[ Pr_{k, Ensemble} = w_{RF} \times Pr_{k,RF} + w_{NB} \times Pr_{k,NB} + w_{SVM} \times Pr_{k,SVM} + w_{k,kNN} \times Pr_{k,kNN} + w_{k,LDA} \times Pr_{k,LDA} + w_{k,LR} \times Pr_{k,LR} \]

Performance of 'Stacking' classifier is evaluated by using TS1. This process is repeated several times (default 100 times).

Value

A matrix of accuracies of each classifier in each iteration.
**Author(s)**
Mattia Chiesa, Luca Piacentini

**Examples**

```r
# use example data:
data(selected_features)
data(df)
set.seed(1)
# only for the example:
# speed up the process setting a low 'iter' argument value;
# for real data set use default 'iter' value (i.e. 100) or higher:
# Classification_res <- DaMiR.EnsembleLearning(selected_features,
# classes=df$class, fSample.tr=0.6, fSample.tr.w=0.6, iter=3,
# cl_type=c("RF","kNN"))
```

---

**DaMiR.EnsL_Predict**  
Predict new samples class

**Description**
The best model learned by the *DaMiR.EnsL_Train* function is tested on a new dataset, in order to predict the samples class.

**Usage**

`DaMiR.EnsL_Predict(data, bestModel)`

**Arguments**

data A SummarizedExperiment object or a data frame/matrix of normalized expression data. Rows and Cols should be observations and features, respectively.

bestModel The best model, selected between those trained by the *DaMiR.EnsL_Train* function.

**Details**
This function implements the prediction step on new data, given a model learned by *DaMiR.EnsL_Train*.

**Value**
A matrix containing the predictions

**Author(s)**
Mattia Chiesa, Luca Piacentini
Examples

# use example data:
data(selected_features)
data(df)

Description

This function tests the models learned by the DaMiR.EnsL_Train function, on a test set

Usage

DaMiR.EnsL_Test(data, classes, EnsL_model)

Arguments

data A SummarizedExperiment object or a data frame/matrix of normalized expression data. Rows and Cols should be observations and features, respectively.
classes A class vector with nrow(data) elements. Each element represents the class label for each observation. Two different class labels are allowed. Note. this argument should not be set when 'data' is a SummarizedExperiment object
EnsL_model A list with the models trained by DaMiR.EnsL_Train function.

Details

This function implements the test step of DaMiR.EnsembleLearning2cl function

Value

A dataframe containing the predictions on the testset

Author(s)

Mattia Chiesa, Luca Piacentini

Examples

# use example data:
data(selected_features)
data(df)
set.seed(1)
# only for the example:
# speed up the process setting a low 'iter' argument value;  
# for real data set use default 'iter' value (i.e. 100) or higher:
# Tr_res <- DaMiR.EnsL_Train(
Train a Binary Classifier using 'Stacking' Learning strategy.

Description

This function learns a meta learner by a 'Stacking' strategy. Users can provide heterogeneous features (other than genomic features) which will be taken into account during classification model building. A 'two-classes' classification task is addressed.

Usage

DaMiR.EnsL_Train(
  data,
  classes,
  variables,
  fSample.tr.w = 0.7,
  cl_type = c("RF", "SVM", "LDA", "LR", "NB", "NN", "PLS")
)

Arguments

data A SummarizedExperiment object or a data frame/matrix of normalized expression data. Rows and Cols should be observations and features, respectively.

classes A class vector with nrow(data) elements. Each element represents the class label for each observation. Two different class labels are allowed. Note: this argument should not be set when 'data' is a SummarizedExperiment object.

variables An optional data frame containing other variables (but without 'class' column). Each column represents a different covariate to be considered in the model.

fSample.tr.w Fraction of samples of training set to be used during weight estimation; default is 0.7.

cl_type List of weak classifiers that will compose the meta-learners. "RF", "SVM", "LDA", "LR", "NB", "NN", "PLS" are allowed. Default is c("RF", "LR", "LDA", "NB", "SVM")

Details

This function implements the training step of DaMiR.EnsembleLearning2cl function.
Value

A list containing:

- The models of each classifier used to build the Ensemble meta-learner with the median or the best accuracy (over the iteration) for the Ensemble classifier;
- the weights associated to each weak classifier;

Author(s)

Mattia Chiesa, Luca Piacentini

Examples

```r
# use example data:
data(selected_features)
data(df)
set.seed(1)
# For the example:
# speed up the process setting a low 'iter' argument value;
# for real data set use default 'iter' value (i.e. 100) or higher:
# Classification_res <- DaMiR.EnsL_Train(
# selected_features,classes=df$class, fSample.tr.w=0.6, iter=3,
# cl_type=c("RF","LR"))
```

---

DaMiR.FBest

Select best predictors to build Classification Model

Description

This function allows the user to select a subset of predictors; the number of predictors can be defined by user or selected automatically.

Usage

```r
DaMiR.FBest(
  data,
  ranking,
  autoselect = c("no", "yes"),
  n.pred = 10,
  th.zscore = 2
)
```
Arguments

data A transposed data frame of expression data. Rows and Cols should be, respectively, observations and features

rankings A data frame with importance score for each feature, generated by DaMiR.FSort

autoselect A flag to specify how to select predictors:
  * "no" (default) - Manually: users can specify the number of best predictors, setting n.pred argument
  * "yes" - Automatically: users have to specify the importance threshold defined by the th.zscore argument; features will be accordingly selected

n.pred If autoselect="no" then the user have to specify the number of predictors; default is 10

th.zscore Threshold of scaled importance score (Z-score); default value is 2

Value

A list containing:

  * A data frame of normalized expression data of the most important selected predictors.
  * A vector with predictors name.

Author(s)

Mattia Chiesa, Luca Piacentini

See Also

DaMiR.FSort

Examples

# use example data:
data(data_reduced)
data(data_relief)
# select the first 8 predictors ranked by importance:
selected_features <- DaMiR.FBest(data_reduced, data_relief, n.pred = 8)
# select predictors by importance but automatically:
selected_features <- DaMiR.FBest(data_reduced, data_relief, autoselect = "yes", th.zscore = 1.5)
DaMiR.FReduct  

Remove highly correlated features, based on feature-per-feature correlation.

Description

This function allows the user to remove highly correlated features.

Usage

DaMiR.FReduct(data, th.corr = 0.85, type = c("spearman", "pearson"))

Arguments

data  A transposed data frame or matrix of normalized expression data. Rows and Cols should be, respectively, observations and features
th.corr  Feature-per-feature correlation threshold; default is 0.85
type  Type of correlation metric to be applied; default is "spearman"

Details

This function produces an absolute correlation matrix that it is then used to reduce pair-wise correlations. When two features present a correlation higher than that defined by the user in th.corr argument, the function, first, calculates the mean absolute correlation of each feature and, then, removes the feature with the largest mean absolute correlation.

Value

An expression matrix without highly correlated features.

Author(s)

Mattia Chiesa, Luca Piacentini

See Also

rcorr, findCorrelation

Examples

# use example data:
data(data_reduced)
# reduce the number of features:
data_Reduced <- DaMiR.FReduct(data_reduced, th.corr = 0.75, type = "pearson")
**Description**

This function identifies the class-correlated principal components (PCs) which are then used to implement a backward variable elimination procedure for the removal of non-informative features.

**Usage**

```r
DaMiR.FSelect(
  data,
  df,
  th.corr = 0.6,
  type = c("spearman", "pearson"),
  th.VIP = 3,
  nPlsIter = 1
)
```

**Arguments**

- `data`: A transposed data frame or a matrix of normalized expression data. Rows and Cols should be, respectively, observations and features.
- `df`: A data frame with known variables; at least one column with 'class' label must be included.
- `th.corr`: Minimum threshold of correlation between class and PCs; default is 0.6. Note. If df$class has more than two levels, this option is disable and the number of PCs is set to 3.
- `type`: Type of correlation metric; default is "spearman".
- `th.VIP`: Threshold for bve_pls function, to remove non-important variables; default is 3.
- `nPlsIter`: Number of times that bve_pls has to run. Each iteration produces a set of selected features, usually similar to each other but not exactly the same! When nPlsIter > 1, the intersection between each set of selected features is performed; so that, only the most robust features are selected. Default is 1.

**Details**

The function aims to reduce the number of features to obtain the most informative variables for classification purpose. First, PCs obtained by principal component analysis (PCA) are correlated with "class". The correlation threshold is defined by the user in `th.corr` argument. The higher is the correlation, the lower is the number of PCs returned. Importantly, if df$class has more than two levels, the number of PCs is automatically set to 3. In a binary experimental setting, users should pay attention to appropriately set the `th.corr` argument because it will also affect the total number of selected features that ultimately depend on the number of PCs. The `bve_pls` function of plsVarSel package is, then, applied. This function exploits a backward variable elimination...
procedure coupled to a partial least squares approach to remove those variable which are less informative with respect to class. The returned vector of variables is further reduced by the following `DaMiR.FReduct` function in order to obtain a subset of non correlated putative predictors.

### Value
A list containing:

- An expression matrix with only informative features.
- A data frame with class and optional variables information.

### Author(s)
Mattia Chiesa, Luca Piacentini

### References

### See Also
- `bve_pls`
- `DaMiR.FReduct`

### Examples
```r
# use example data:
data(data_norm)
data(df)
# extract expression data from SummarizedExperiment object # and transpose the matrix:
t_data<-t(assay(data_norm))
t_data <- t_data[, seq_len(100)]
# select class-related features
data_reduced <- DaMiR.FSelect(t_data, df,
th.corr = 0.7, type = "spearman", th.VIP = 1)
```

---

**DaMiR.FSort** *Order features by importance, using RReliefF filter*

### Description
This function implements a procedure in order to rank features by their importance evaluated by RReliefF score.
Usage

DaMiR.FSort(data, df, fSample = 1)

Arguments

data
A transposed data frame of expression data, i.e. transformed counts by vst or rlog. A log2 transformed expression matrix is also accepted. Rows and Cols should be, respectively, observations and features

df
A data frame with class and known variables; at least one column with ‘class’ label must be included

fSample
Fraction of sample to be used for the implementation of RReliefF algorithm; default is 1

Details

This function is very time-consuming when the number of features is high. We observed there is a quadratic relationship between execution time and the number of features. Thus, we have also implemented a formula which allows the users to estimate the time to perform this step, given the number of features. The formula is:

\[ T = 0.0011 \times N^2 - 0.1822 \times N + 27.092 \]

where \( T \) = Time and \( N \) = Number of genes. We strongly suggest to filter out non informative features before performing this step.

Value

A data frame with two culmuns, where features are sorted by importance scores:

- RReliefF score - Calculated by relief function, implemented in FSelector package;
- scaled.RReliefF score - Z-score value, computed for each RReliefF score.

A plot with the first 50 features ordered by their importance.

Author(s)

Mattia Chiesa, Luca Piacentini

References


See Also

relief, DaMiR.FSelect, DaMiR.FReduct
Examples

# use example data:
data(data_reduced)
data(df)

# rank features by importance:
df.importance <- DaMiR.FSort(data_reduced[,1:10],
                               df, fSample = 0.75)

---

DaMiR.goldenDice Generate a Number to Set Seed

Description

This function implements a formula based on current date and time.

Usage

DaMiR.goldenDice()

Details

The number is generated by combining current seconds (S), minutes (Mi), hours (H), days (D), months (Mo), years (Y) and golden ratio ($\phi$), in the form:

$$Num = (S \times Mi + H \times D \times Mo/D)^\phi$$

Value

An integer number.

Author(s)

Mattia Chiesa, Luca Piacentini

Examples

gen_num <- DaMiR.goldenDice()
set.seed(gen_num)
**Description**

This function aims to perform a batch correction on a normalized independent test set, exploiting the ComBat function of the sva package.

**Usage**

```
DaMiR.iTSadjust(adj_Learning_set, norm_Ind_Test_set, iTS_batch)
```

**Arguments**

- `adj_Learning_set`  
  A SummarizedExperiment object or a data frame/matrix of adjusted and normalized data, obtained by the `DaMiR.SVadjust` function.

- `norm_Ind_Test_set`  
  A data frame or a matrix of normalized data. The independent test set is supposed to be already normalized by the `DaMiR.iTSnorm` function.

- `iTS_batch`  
  (Optional). A factor or a data.frame, containing information regarding experimental batches of the independent test set. Users can ignore this argument, if the independent test set is deemed a single experimental batch.

**Details**

The function applied a batch correction procedure to the independent test set, normalized by `DaMiR.iTSnorm`.

**Value**

A matrix containing a normalized and adjusted expression matrix (log2 scale).

**Author(s)**

Mattia Chiesa, Luca Piacentini

**References**


**See Also**

ComBat
Examples

# use example data:
data(SE)

---

**Description**

This function aims to normalize properly an actual independent test set by taking information from the Learning set that will be used to transform the new sample(s).

**Usage**

```r
damiriTSnorm(
  Learning_set, 
  Ind_Test_set, 
  normtype = c("vst", "rlog", "logcpm"), 
  method = c("precise", "quick")
)
```

**Arguments**

- **Learning_set**: A SummarizedExperiment object or a data frame/matrix of raw count data. The learning set is supposed to be a raw counts dataset of the expressed features (not all features). Rows and Cols should be features and samples, respectively.
- **Ind_Test_set**: A SummarizedExperiment object or a data frame/matrix of raw count data. The independent test set is supposed to be a raw counts dataset with the same features of 'Learning_set'. Rows and Cols should be features and samples, respectively.
- **normtype**: Type of normalization to be applied: varianceStabilizingTransformation (vst), rlog or logcpm are allowed; default is "vst".
- **method**: Type of method to estimate the dispersion, applied to the independent test set to normalize data. Only 'precise' and 'quick' are allowed. In the first case, the dispersion is estimated by the Learning set and applied to the independent test set. In the second case, is estimated from the independent test set. Default is "precise". See details in dispersionFunction

**Details**

The Learning_set is supposed to be a raw counts dataset of the expressed features. Moreover, the independent test set is supposed to be a raw counts dataset with the same features of 'Learning_set'. The independent test set is normalized, taking into account the dispersion parameter, estimated by the Learning set ('precise' method) or by the independent test set itself ('quick' method).
Value

A matrix containing a normalized expression matrix (log2 scale)

Author(s)

Mattia Chiesa, Luca Piacentini

References


See Also

varianceStabilizingTransformation, rlog cpm

Examples

# use example data:
data(SE)

DaMiR.makeSE(x, y)

Arguments

x  A tab-delimited file which contains RNA-Seq count data. Each row is a feature (i.e. gene, transcript, exon etc.) and each column is a sample

y  A tab-delimited file which contains experiment information. Each row is a sample and each column is a variable. This file must contain at least one column which represent 'class' information for data adjustment and classification; the class column must be labeled as 'class'

Details

Before creating a SummarizedExperiment object, the function performs some checks on input data to ensure that only a matrix of raw counts is accordingly loaded. Other checks allows the identification of missing data (NA) in the data frame of the variables of interest.
Value

A SummarizedExperiment object containing raw counts, class and (optionally) variables of interest.

Author(s)

Mattia Chiesa, Luca Piacentini

References


See Also

SummarizedExperiment

Examples

rawdata.path <- system.file(package = "DaMiRseq","extdata")
# import tab-delimited files:
# sample data are a small subset of Genotype-Tissue Expression (GTEx)
# RNA-Seq database (dbGap Study Accession: phs000424.v6.p1):
count_data <- read.delim(file.path(rawdata.path, "counts_import.txt"))
variables_data <- read.delim(file.path(rawdata.path, "annotation_import.txt"))
# create a SummarizedExperiment object:
SE <- DaMiR.makeSE(count_data, variables_data)
print(SE)

Description

A MDS plot is drawn in order to visualize class clustering.

Usage

DaMiR.MDSplot(data, df, type = c("spearman", "pearson"))

Arguments

data A SummarizedExperiment object or a matrix or a data.frame where rows and cols should be, respectively, observations and features
df A data frame with class; it can be directly subset from data
type A character string specifying the metric to be applied to correlation analysis. Either "spearman" or "pearson" is allowed; default is "spearman"
**Details**

The MDS plot is drawn taking as input a dissimilarity matrix produced by either a sample-per-sample Pearson’s or Spearman’s correlation of normalized expression data.

**Value**

A MDS plot, using only 'class' information

**Author(s)**

Mattia Chiesa, Luca Piacentini

**Examples**

```r
# use example data:
data(data_reduced)
data(df)
# Draw MDS:
DaMiR.MDSplot(data=data_reduced, df=df, type="pearson")
```

---

**DaMiR.ModelSelect**  
*Select the best classification model*

**Description**

This function selects the best models learned by the `DaMiR.EnsL_Train` and tested by `DaMiR.EnsL_Test`.

**Usage**

```r
DaMiR.ModelSelect(
  df,
  type.sel = c("mode", "median", "greater"),
  th.sel = 0.85,
  npred.sel = c("min", "rnd"),
  metric.idx = 1,
  npred.idx = 2
)
```

**Arguments**

- `df`  
  A data frame of performance metrics. At least two columns representing a specific classification metrics (e.g., Accuracy) and the number of predictors must be provided. Additionally, other classification metrics (e.g., MCC, Sensitivity, Specificity, PPV, NPV, AUC, ...) can be appended (from the third column onwards) and used for the evaluation, by correctly setting the 'metric.idx' parameter.
DaMiR.normalization

**type.sel**
The method to select the best models. Only "mode","median" and "greater" values are allowed. For a specific classification metrics, "mode" selects all models whose score is the mode of all scores; "median" selects all models whose score is the median of all scores; and, "greater" selects all models whose score is greater than the value specified in "th.sel". Default: "mode".

**th.sel**
Threshold for the evaluation of the performance when "type.sel" is equal to "greater". Default: 0.85

**npred.sel**
The method to select the best model. Only "min" and "rnd" values are allowed. Taking into account the subset of models found by 'type.sel', this parameter selects one single model with the minimum number of predictors ("min") or randomly ("rnd"). Default: "min".

**metric.idx**
The index of the 'df' column (i.e., classification metrics) to be considered for the models evaluation. Default: 1.

**npred.idx**
The index of the 'df' column representing the number of predictors. Default: 2.

**Details**
This function finds the best model, taking into account specific classification metrics.

**Value**
The index of df (row), representing the model selected and a bubble chart

**Author(s)**
Mattia Chiesa, Luca Piacentini

**Examples**

```r
# use example data:
set.seed(1)
```

---

DaMiR.normalization  
*Filter non Expressed and 'Hypervariant' features and Data Normalization*

**Description**
Features will be firstly filtered based on their expression value and/or by their variability across samples; features will be then normalized.
**Usage**

```r
DaMiR.normalization(
  data,
  minCounts = 10,
  fSample = 0.5,
  hyper = c("yes", "no"),
  th.cv = 3,
  type = c("vst", "rlog", "logcpm"),
  nFitType = c("parametric", "local", "mean")
)
```

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td><code>data</code></td>
<td>A SummarizedExperiment object</td>
</tr>
<tr>
<td><code>minCounts</code></td>
<td>Minimum reads counts; default is 10</td>
</tr>
<tr>
<td><code>fSample</code></td>
<td>Fraction of samples with <code>minCounts</code> counts; default is 0.5</td>
</tr>
<tr>
<td><code>hyper</code></td>
<td>Flag to enable gene filtering by Coefficient of Variation (CV); default is &quot;yes&quot;</td>
</tr>
<tr>
<td><code>th.cv</code></td>
<td>Threshold of minimum CV to consider a feature 'Hypervariant' across samples; default is 3</td>
</tr>
<tr>
<td><code>type</code></td>
<td>Type of normalization to be applied: varianceStabilizingTransformation (vst), rlog or logcpm are allowed; default is &quot;vst&quot;</td>
</tr>
<tr>
<td><code>nFitType</code></td>
<td>Type of method to estimate the dispersion by vst or rlog. Default is &quot;parametric&quot;.</td>
</tr>
</tbody>
</table>

**Details**

Before normalization step, this function allows the user to filter features by:

- **Expression** - Features will be filtered out whether their reads count do not reach a `minCounts` in at least `fSample` of samples;
- **CV** - The CV of each feature is individually calculated for each sample class. Features with both class CV greater than `th.cv` will be discarded. Computing a class restricted CV may prevent the removal of hypervariant features that may be specifically associated with a certain class. This could be important, for example, for immune genes whose expression under definite conditions may unveil peculiar class-gene association.

Finally, expressed features will be normalized by `varianceStabilizingTransformation` (default) or `rlog`, both implemented in DESeq2 package. We suggest to use `varianceStabilizingTransformation` to speed up the normalization process because `rlog` is very time-consuming despite the two methods produce quite similar results.

**Value**

A SummarizedExperiment object which contains a normalized expression matrix (log2 scale) and the data frame with 'class' and (optionally) variables.

**Author(s)**

Mattia Chiesa, Luca Piacentini
DaMiR.sampleFilt

Filter Samples by Mean Correlation Distance Metric

Description
This function implements a sample-per-sample correlation. Samples with a mean correlation lower than a user's defined threshold will be filtered out.

Usage
DaMiR.sampleFilt(data, th.corr = 0.9, type = c("spearman", "pearson"))

Arguments
- data: A SummarizedExpression object
- th.corr: Threshold of mean correlation; default is 0.9
- type: Type of correlation metric; default is "spearman"

Details
This step introduces a sample quality checkpoint. Global gene expression should, in fact, exhibit a high correlation among biological replicates; conversely, low correlated samples may be suspected to bear some technical artifact (e.g. poor RNA or library preparation quality), despite they may have passed sequencing quality checks. If not assessed, these samples may, thus, negatively affect all the downstream analysis. This function looks at the mean absolute correlation of each sample and removes those samples with a mean correlation lower than the value set in th.corr argument. This threshold may be specific for different experimental setting but should be as high as possible. For sequencing data we suggest to set th.corr greater than 0.85.
**Value**

A SummarizedExperiment object which contains a normalized and filtered expression matrix (log2 scale) and a filtered data frame with 'class' and (optionally) variables.

**Author(s)**

Mattia Chiesa, Luca Piacentini

**Examples**

```r
# use example data:
data(data_norm)
# filter out samples with Pearson's correlation <0.92:
data_filt<- DaMiR.sampleFilt(data_norm, th.corr=0.92, type="pearson")
```

---

**DaMiR.SV**

*Identification of Surrogate Variables*

**Description**

This function returns a matrix of surrogate variables (sv) using the implementation by Chiesa-Piacentini or the sva method by Leek et al.

**Usage**

```r
DaMiR.SV(
data,
method = c("fve", "leek", "be"),
th.fve = 0.95,
second.var = NULL
)
```

**Arguments**

- `data`: A SummarizedExpression object
- `method`: The method used to identify sv. If missing, the "fve" method will be selected. Otherwise the method "leek" or "be" should be choosen
- `th.fve`: This argument sets the threshold of maximum fraction of variance explained (fve) to be used in conjunction with "fve" method; default is 0.95
- `second.var`: A factor or a numeric vector corresponding to an additional variable to take into account during the sv identification. This variable together with 'class' in the data object will be used to design the model matrix (~ class + second.var)
Details

This function helps the user to identify the appropriate number of sv: it is possible to select a different strategy to be used by changing the option in method argument. Three methods are available:

- "be" - this option uses the num.sv function of sva package with default parameters;
- "leek" - The same of before but with asymptotic approach proposed by Leek;
- "fve" - This method is introduced in DaMiRseq package, and integrates part of sva function with custom code. Briefly, we computed eigenvalues of data using code already implemented in sva function and then, we calculated the squared of each eigenvalues. Thus, the ratio between each “squared eigenvalue” and the sum of them were calculated. These values represent a surrogate measure of the "Percentage of Explained Variance" (pve) obtained by principal component analysis (PCA), and their cumulative sum can be used to select sv.

Value

A matrix of sv. A plot with the sv identified by "fve" method is also returned. A red dot shows the maximum number of variables to be included for a specific "fve".

Author(s)

Mattia Chiesa, Luca Piacentini

References


See Also

sva

Examples

# use example data:
data(data_norm)
sv <- DaMiR.SV(data_norm, method = "fve", th.fve=0.95)

DaMiR.SVadjust Remove variable effects from expression data

Description

This function removes surrogate or other confounding variable effects from normalized expression data by the usage of removeBatchEffect function of limma package.
DaMiR.transpose

Usage

DaMiR.SVadjust(data, sv, n.sv)

Arguments

- data: A SummarizedExpression object
- sv: The matrix of surrogate variables identified by DaMiR.SV function
- n.sv: The number of surrogate variables to be used to adjust the data

Value

A SummarizedExpression object containing a matrix of log-expression values with sv effects removed and the data frame of the variables.

Author(s)

Mattia Chiesa, Luca Piacentini

See Also

removeBatchEffect, DaMiR.SV

Examples

# use example data:
data(data_norm)
data(sv)
data_adjust <- DaMiR.SVadjust(data_norm, sv = sv, n.sv = 3)

---

DaMiR.transpose  Matrix transposition and replacement of '.' and '-' special characters

Description

This function transposes matrix and replaces '.' and '-' special characters.

Usage

DaMiR.transpose(data)

Arguments

- data: Matrix of normalized expression data, i.e. transformed counts by vst or rlog. A log2 transformed expression matrix is also accepted
### data_norm

**Value**

Normalized matrix in which each row is a sample and each column is a feature

**Author(s)**

Mattia Chiesa, Luca Piacentini

**Examples**

```r
data(data_norm)
data.transposed <- DaMiR.transpose(assay(data_norm))
```

### data_min

**Description**

A dataset with a small dimension of normalized expression data in DaMiRseq package

**Usage**

```r
data_min
```

**Format**

A data frame with 40 samples (rows) and 87 genes (columns)

**Value**

An example dataset for DaMiRseq package

### data_norm

**Description**

A dataset with a normalized matrix to test several DaMiRseq functions: sample data are a subset of Genotype-Tissue Expression (GTex) RNA-Seq database (dbGap Study Accession: phs000424.v6.p1)

**Usage**

```r
data_norm
```
Format
A SummarizedExperiment object containing an assay of 4897 genes (rows) and 40 samples (columns) and a colData with 5 variables

Value
An example dataset for DaMiRseq package

data_reduced

Example gene-expression dataset for DaMiRseq package

Description
A dataset with a small dimension of normalized expression data in DaMiRseq package

Usage
data_reduced

Format
A list with:
data reduced expression matrix
variables a data frame with variables

Value
An example dataset for DaMiRseq package

data_relief

Example ranking dataset for DaMiRseq package

Description
A data frame with relieF and scaled relieF scores for each gene

Usage
data_relief

Format
A dataframe with 87 genes (rows) and 2 variables (columns):
reliefF Score relieF score for each gene
scaled relieF Score scaled relieF score for each gene, by z-score
Example dataset for DaMiRseq package

**df**

Example gene-expression dataset for DaMiRseq package

**Description**

A data frame with class and covariates information

**Usage**

df

**Format**

A dataframe with 40 samples (rows) and 5 variables (columns):

- **center**: center where sample has been collected
- **sex**: sample’s gender
- **age**: sample’s age
- **death**: kind of sample’s death, based on Hardy scale
- **class**: sample’s class

**Value**

An example dataset for DaMiRseq package

**Description**

A dataset with count matrix to test several DaMiRseq functions. To show package functionality in a reasonably execution time, sample data are a subset of Genotype-Tissue Expression (GTex) RNA-Seq database (dbGap Study Accession: phs000424.v6.p1). Samples include 20 Anterior Cingulate Cortex (ACC) tissues and 20 Frontal Cortex (FC) tissues. 21363 genes have been previously selected to have 5 read counts in at least 60

**Usage**

SE
**selected_features**

**Format**
A SummarizedExperiment object containing an assay of 21363 randomly selected genes (rows) and 40 samples (columns) and a colData with 5 variables

**Value**
An example dataset for DaMiRseq package

<table>
<thead>
<tr>
<th>selected_features</th>
<th>Example gene-expression dataset for DaMiRseq package</th>
</tr>
</thead>
</table>

**Description**
A dataset with normalized expression data to build classification models in DaMiRseq package

**Usage**
```r
selected_features
```

**Format**
A dataframe with 40 samples (rows) and 7 variables (genes):

**Value**
An example dataset for DaMiRseq package

<table>
<thead>
<tr>
<th>SEtest_norm</th>
<th>A sample dataset with a normalized count matrix for &quot;testthat&quot; functions.</th>
</tr>
</thead>
</table>

**Description**
A sample dataset with a normalized count matrix for "testthat" functions.

**Usage**
```r
SEtest_norm
```

**Format**
A SummarizedExperiment object containing an assay of 100 genes (rows) and 11 samples (columns) and a colData with 5 variables

**Value**
An example dataset for DaMiRseq package
Example Surrogate Variables dataset for DaMiRseq package

Description

A dataset with surrogate variables to test DaMiRseq functions

Usage

sv

Format

A matrix with 40 samples (rows) and 4 surrogate variables (columns):

Value

An example dataset for DaMiRseq package
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