

Package ‘DExMA’

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Type Package

Title Differential Expression Meta-Analysis

Version 1.0.2

Description performing all the steps of gene expression meta-analysis without eliminating those genes that are presented in at least two datasets. It provides the necessary functions to be able to perform the different methods of gene expression meta-analysis. In addition, it contains functions to apply quality controls, download GEO data sets and show graphical representations of the results.

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Depends R (>= 4.1), DExMAdata

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biocViews DifferentialExpression, GeneExpression, StatisticalMethod, QualityControl

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allSameID	<i>Set all datasets in the same ID</i>
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Description

Set all datasets in the same ID (Official Gene Symbol, Entrez or Ensembl)

Usage

```
allSameID(objectMA, initialIDs, finalID = "GeneSymbol",
           organism="Homo sapiens")
```

Arguments

objectMA	A list of list. Each list contains two elements. The first element is the expression matrix (genes in rows and sample in columns) and the second element is a vector of zeros and ones that represents the state of the different samples of the expression matrix. 0 represents one group (controls) and 1 represents the other group (cases). The result of the CreateobjectMA can be used too.
initialIDs	A vector in which each element indicates the ID of the equivalent element of listExpMatrix. To know the available ids, you can write availableIDs
finalID	A character that indicates the final ID all the different studies will have. To know the available ids, you can write availableIDs.
organism	A character that indicates the organism of the data. To know the available organisms write availableOrganism

Value

The same list with all the datasets in the same selected gene ID.

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

See Also[createObjectMA](#)**Examples**

```
data(DExMAExampleData)
sameData <- allSameID(objectMA = maObjectDif, initialIDs = c("Entrez",
"Entrez", "GeneSymbol", "GeneSymbol"), finalID = "GeneSymbol",
organism = "Homo sapiens")
sameData
```

batchRemove*Elimination of covariates batch effect or bias*

Description

It eliminates the effects of batch or bias of the covariates

Usage

```
batchRemove(expressionMatrix, pheno, formula, mainCov = NULL, nameGroup)
```

Arguments

expressionMatrix	A matrix or data frame with genes in rows and samples in columns. An ExpressionSet object can be used too
pheno	A dataframe with samples in rows and covariates in columns.
formula	Formula of the covariates that are wanted to be corrected
mainCov	Name of the main covariate to be corrected
nameGroup	Name of the column of the Phenodata object in which the reference groups (cases and controls) are

Value

The Expression Matrix with the bias or batch effect corrected. Moreover a plot of the visualization of the association between principal components and covariates is shown.

Note

For more information of the returned visualization see the seeCOV function

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

References

Martin Lauss (2019). *swamp: Visualization, Analysis and Adjustment of High-Dimensional Data in Respect to Sample Annotations*. R package version 1.5.1. <https://CRAN.R-project.org/package=swamp>

See Also

[seeCov](#)

Examples

```
data(DEXMAExampleData)
batchRemove(listMatrixEX$Study2, listPhenodatas$Study2, formula=~gender+race,
nameGroup="condition")
```

calculateES

Calculation of Effects Sizes and their variance

Description

This function uses the Hedges'g estimator to calculate the different Effects size and their variances for each genes and for each dataset.

Usage

```
calculateES(objectMA, missAllow = 0.3)
```

Arguments

objectMA	A list of list. Each list contains two elements. The first element is the expression matrix (genes in rows and sample in columns) and the second element is a vector of zeros and ones that represents the state of the different samples of the expression matrix. 0 represents one group (controls) and 1 represents the other group (cases). The result of the CreateobjectMA can be used too.
missAllow	a number that indicates the maximun proportion of missing values allowed in a sample. If the sample has more proportion of missing values the sample will be eliminated. In the other case the missing values will be imputed using the K-NN algorithm.

Value

A list formed by three elements:

- First element (ES) is a dataframe were columns are each of the studies (datasets) and rows are the genes. Each element of the dataframe represents the Effect Size.
- Second element (Var) is a dataframe were columns are each of the studies (datasets) and rows are the genes. Each element of the dataframe represents the variance of the Effect size.

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

See Also

[createObjectMA](#), [metaAnalysisDE](#)

Examples

```
data(DEXMAExampleData)

resultsEffects <- calculateES(objectMA = maObject, missAllow = 0.3)
resultsEffects
```

createObjectMA	<i>Creation of the object to use in meta-analysis</i>
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Description

It allows the creation of an object to perform meta-analysis.

Usage

```
createObjectMA(
  listEX,
  listPheno = NULL,
  namePheno = c(rep(1, length(listEX))),
  expGroups = c(rep(1, length(listEX))),
  refGroups = c(rep(2, length(listEX)))
)
```

Arguments

listEX	A list of dataframes or matrix (genes in rows and sample in columns). A list of ExpressionSets can be used too
listPheno	A list of phenodatas (dataframes or matrix). If the object listEX is a list of ExpressionSets this element can be null.
namePheno	A list or vector of the different column names or positions from the phenodatas where the experimental and reference groups are identified. Each element of namePheno correspond to its equivalent element in the listPheno (default a vector of 1, all the first columns of each elements of listPheno are selected).
expGroups	A list of vectors or a vector containing the names or the positions with which we identify the elements of the experiment groups (cases) of the namePheno element (default a vector of 1, all the first groups are selected)
refGroups	A list of vectors or a vector containing the names or the positions with which we identify the elements of the reference groups (control) of the namePheno elements (default a vector of 1, all the first groups are selected)

Value

The object needed to perform meta-analysis. This object is list of nested lists. Each list contains two elements:

- The first element is the expression matrix (genes in rows and sample in columns)
- The second element is a vector of zeros and ones that represents the state of the different samples of the expression matrix. 0 represents reference group (controls) and 1 represents experimental group (cases).

Author(s)

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See Also

[elementObjectMA](#)

Examples

```
data(DEXMAExampleData)

phenoGroups = c("condition", "condition", "state", "state")
phenoCases = list(Study1 = "Diseased", Study2 = c("Diseased", "ill"),
                  Study3 = "Diseased", Study4 = "ill")
phenoControls = list(Study1 = "Healthy", Study2 = c("Healthy", "control"),
                    Study3 = "Healthy", Study4 = "control")

newObjectMA <- createObjectMA(listEX=listMatrixEX, listPheno = listPhenodatas,
                             namePheno=phenoGroups, expGroups=phenoCases,
                             refGroups = phenoControls)

newObjectMA
```

dataLog	<i>Auxiliary function to check if data are log transformed and transformed if it are not log-transformed</i>
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Description

Auxiliary function to check if data are log transformed and transformed if it are not log-transformed

Usage

```
dataLog(objectMA)
```

Arguments

`objectMA` A list of list. Each list contains two elements. The first element is the expression matrix (genes in rows and sample in columns) and the second element is a vector of zeros and ones that represents the state of the different samples of the expression matrix. 0 represents one group (controls) and 1 represents the other group (cases). The result of the `CreateObjectMA` should be used.

Value

The same object with log-transformed expression matrix

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

See Also

[createObjectMA](#)

Examples

```
data(DEXMAExampleData)
dataLog(maObject)
```

`downloadGEOData`

Download datasets from GEO database

Description

Download different ExpressionSets objects from the GEO database

Usage

```
downloadGEOData(GEOobject, directory = getwd())
```

Arguments

`GEOobject` a vector of character where each element represents a GEO object for downloading.

`directory` The directory where the different downloaded GSE Series Matrix files from GEO will be stored. By default they are downloaded to the working directory

Details

This function internally uses `getGEO` function of `GEOquery` package. However, `downloadGEO` allows you to download multiple files at the same time.

Value

A list of the different ExpressionSets

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

References

Davis, S. and Meltzer, P. S. GEOquery: a bridge between the Gene Expression Omnibus (GEO) and BioConductor. *Bioinformatics*, 2007, 14, 1846-1847

Examples

```
## Not run:
GEOobjects<- c("GSE4588", "GSE10325")
dataGEO<-downloadGEOData(GEOobjects)
dataGEO

## End(Not run)
```

elementObjectMA

Creation of the object to use in meta-analysis

Description

It allows the creation of a element of the object needed to perform meta-analysis

Usage

```
elementObjectMA(
  expressionMatrix,
  pheno = NULL,
  groupPheno,
  expGroup = 1,
  refGroup = 2
)
```

Arguments

expressionMatrix	A dataframe or matrix that containing genes in rows and samples if columns. An ExpressionSet object can be used too.
pheno	A data frame or a matrix containing samples in rows and covariates in columns. If NULL (default), pheno is extracted from the ExpressionSet object
groupPheno	The column name or position from pheno where experimental group (cases) and reference group (control) are identified

expGroup	The group name or position from groupPheno variable used as experimental group (cases). By default the first group (character) is taken
refGroup	The group name or position from groupPheno variable used as reference group (control). By default the second group (character) is taken

Value

An element that can be included in meta-analysis object.

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

See Also

[createObjectMA](#)

Examples

```
data(DEXMAExampleData)

ExpressionSetStudy5
newElem <-elementObjectMA(expressionMatrix = ExpressionSetStudy5,
                           groupPheno = "condition",
                           expGroup = c("Diseased", "ill"),
                           refGroup = c("Healthy", "control"))
```

heterogeneityTest *It allows checking the heterogeneity of the different studies*

Description

Shows a QQ-plot of the Cochran's test and the quantiles of I^2 statistic values to measure heterogeneity

Usage

```
heterogeneityTest(objectMA)
```

Arguments

objectMA A list of list. Each list contains two elements. The first element is the expression matrix (genes in rows and sample in columns) and the second element is a vector of zeros and ones that represents the state of the different samples of the expression matrix. 0 represents one group (controls) and 1 represents the other group (cases). The result of the CreateobjectMA can be used too.

Details

If in the QQ-plot of the Cochran's test most of the values are close to the central line (most of the Cochran's test values are close to the expected distribution), it can be said that there is homogeneity. In the case that these values deviate greatly from the expected distribution, it must be assumed that there is heterogeneity. I^2 measures the percentage of variation across studies due to heterogeneity. To assume homogeneity in the gene expression meta-analysis, almost all I^2 values (quantiles) must be 0 or at least less than 0.25.

Value

Deciles of the I^2 values

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

References

Higgins JPT, Thompson SG. Quantifying heterogeneity in a meta-analysis. *Stat Med* 2002;21:1539–58.
Higgins JPT, Thompson SG, Deeks JJ, et al. Measuring inconsistency in meta-analyses. *BMJ* 2003;327:557–60.

See Also

[createObjectMA](#)

Examples

```
data(DEXMAExampleData)
heterogeneityTest(maObject)
```

makeHeatmap

Visualization of the meta-analysis results

Description

It allows to see how the different significant genes are expressed in the different samples. Missing genes appear in gray

Usage

```
makeHeatmap(
  objectMA,
  resMA,
  typeMethod = c("FEM", "REM", "maxP", "minP", "Fisher", "Stouffer"),
  scaling = c("zscor", "rscale", "swr", "none"),
  regulation = c("all", "up", "down"),
  breaks = c(-2, 2),
  fdrSig = 0.05,
  numSig = 50
)
```

Arguments

objectMA	A list of list. Each list contains two elements. The first element is the expression matrix (genes in rows and sample in columns) and the second element is a vector of zeros and ones that represents the state of the different samples of the expression matrix. 0 represents one group (controls) and 1 represents the other group (cases). The result of the CreateobjectMA can be used too.
resMA	Output generated by the differents functions that performs meta-analysis (metaES, metaPvalue, metaRank or metaAnalysisDE)
typeMethod	A character that indicates the method that have been used to perform meta-analysis. See metaAnalysisDE function for more information.
scaling	Character variable to choose between different scaling approaches. See "Details" for more information.
regulation	Character variable that indicates whether we want the heatmap to show all significant genes ("all"), only the up-regulated genes ("up") or only the down-regulated genes("down")
breaks	Numeric vector of length 2 that contains the extreme values (minimum and maximum) of the range of values in which the heatmap color scale will be distributed. Default a vector By default a vector of -2 and 2 as extreme values.
fdrSig	Adjusted p-value from which a gene is considered significant. Default 0.05
numSig	The number of most significant genes to be represented

Details

Scaling approaches that can be used are:

- "rscale": it applies rescale function of *scales* package. Values will be between -1 and 1)
- "zscor": It calculates a z-score value for each gene, that is, the mean gene expression from each gene is subtracted from each gene expression value and then it is divided by the standard deviation
- "swr": it applys scaling relative to reference dataset approach
- "none": any scaling approach it is applied.

Value

'NULL'

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

References

Hadley Wickham and Dana Seidel (2020). scales: Scale Functions for Visualization. R package version 1.1.1. <https://CRAN.R-project.org/package=scales>

Lazar, C, Meganck, S, Taminau, J, and et al. 2013. "Batch Effect Removal Methods for Microarray Gene Expression Data Integration: A Survey," 469–90.

Raivo Kolde 2019. pheatmap: Pretty Heatmaps. R package version 1.0.12. <https://CRAN.R-project.org/package=pheatmap>

See Also

[createObjectMA](#), [metaAnalysisDE](#)

Examples

```
data(DExMAExampleData)

resultsMA <- metaAnalysisDE(maObject, typeMethod="REM")
makeHeatmap(objectMA=maObject, resMA=resultsMA, typeMethod="REM",
  scaling = "zscor", regulation = "all", breaks=c(-2,2),
  fdrSig = 0.05,numSig=40)
```

metaAnalysisDE

Performing Meta-analysis

Description

It performs meta-analysis using eight different methods.

Usage

```
metaAnalysisDE(
  objectMA,
  typeMethod = c("FEM", "REM", "maxP", "minP", "Fisher",
    "Stouffer"),
  missAllow = 0.3,
  proportionData = 0.5
)
```

Arguments

objectMA	A list of list. Each list contains two elements. The first element is the expression matrix (genes in rows and sample in columns) and the second element is a vector of zeros and ones that represents the state of the different samples of the expression matrix. 0 represents one group (controls) and 1 represents the other group (cases). The result of the CreateobjectMA can be used too.
typeMethod	A character that indicates the method to be performed. See "Details" for more information
missAllow	a number that indicates the maximum proportion of missing values allowed in a sample. If the sample has more proportion of missing values the sample will be eliminated. In the other case the missing values will be imputed using the K-NN algorithm.
proportionData	The minimum proportion of datasets in which a gene must be contained to be included. By default, the gene must be contained in at least half of the datasets

Details

The different meta-analysis methods that can be applied are:

1. Effects sizes methods:

- "FEM": Fixed Effects model
- "REM": Random Effects model

2. P-value combination methods

- "Fisher": Fisher's methods
- "Stouffer": Stouffer's method
- "maxP": maximum p-value method (Wilkinson's method)
- "minP": minimum p-value method (Tippet's method)

Value

A dataframe with the meta-analysis results. Depending on the applied method, a different dataframe is obtained. For more information see the package vignette.

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

References

Daniel Toro-Domínguez, Juan Antonio Villatoro-García, Jordi Martorell-Marugán, Yolanda Román-Montoya, Marta E Alarcón-Riquelme, Pedro Carmona-Sáez, A survey of gene expression meta-analysis: methods and applications, *Briefings in Bioinformatics*, 2020, bbaa019, <https://doi.org/10.1093/bib/bbaa019>

Michael Dewey (2020). *metap: meta-analysis of significance values*.

Examples

```
data(DExMAExampleData)
ResultsMA <- metaAnalysisDE(objectMA=maObject, typeMethod="REM",
                           missAllow=0.3, proportionData=0.5)
ResultsMA
```

pvalueIndAnalysis *Calculation p-value for each gene and study*

Description

This function uses t-test based on limma package in order to obtain the individual p-values for each study and gene

Usage

```
pvalueIndAnalysis(objectMA, missAllow = 0.3)
```

Arguments

objectMA	A list of list. Each list contains two elements. The first element is the expression matrix (genes in rows and sample in columns) and the second element is a vector of zeros and ones that represents the state of the different samples of the expression matrix. 0 represents one group (controls) and 1 represents the other group (cases). The result of the CreateObjectMA can be used too.
missAllow	a number that indicates the maximum proportion of missing values allowed in a sample. If the sample has more proportion of missing values the sample will be eliminated. In the other case the missing values will be imputed using the K-NN algorithm.

Value

A list formed by two elements:

- First element (p) is a dataframe where columns are each of the studies (datasets) and rows are the genes. Each element of the dataframe represents the p-value.
- Second element (FC) is a dataframe where columns are each of the studies (datasets) and rows are the genes. Each element of the dataframe is the logFC.
- Third element (weights_z) is a dataframe where columns are each of the studies (datasets) and rows are the genes. Each element of the dataframe represents the necessary weights for Stouffer's method.

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

See Also

[createObjectMA](#), [metaAnalysisDE](#)

Examples

```
data(DEXMAexampleData)

pvalues <- pvalueIndAnalysis(objectMA=maObject, missAllow=0.3)
pvalues
```

seeCov

Visualization the effect of covariates in data variability

Description

It uses the `prince` and `prince.plot` function from the `swamp` package to visualize the effect of covariates in data variability

Usage

```
seeCov(expressionMatrix, pheno)
```

Arguments

<code>expressionMatrix</code>	A matrix or data frame with genes in rows and samples in columns. An <code>ExpressionSet</code> object can be used too
<code>pheno</code>	A dataframe with samples in rows and covariates in columns. It should contain only the most important covariates

Value

A visualization (heatmap) in which it can be seen how the data variability is affected by the covariates. The plot represents the p-values of each principal component associated with the covariates.

Note

Requires the package `swamp`

Author(s)

Juan Antonio Villatoro Garcia, <juanantoniovillatorogarcia@gmail.com>

References

Martin Lauss (2019). `swamp`: Visualization, Analysis and Adjustment of High-Dimensional Data in Respect to Sample Annotations. R package version 1.5.1. <https://CRAN.R-project.org/package=swamp>

See Also[batchRemove](#)**Examples**

```
data(DEXMAExampleData)
seeCov(listMatrixEX$Study2, listPhenodatas$Study2)
```


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