Package ‘AllelicImbalance’

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License GPL-3
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'GvizTrack-methods.R' 'LinkVariantAlmlof-class.R'
'RegionSummary-class.R' 'RiskVariant-class.R'
'auxillary-functions-annotation.R'
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Description

Package AllelicImbalance has functions for importing, filtering and plotting high-throughput data to make an allele specific expression analysis. A major aim of this package is to provide functions to collect as much information as possible from regions of choice, and to be able to explore the allelic expression of that region in detail.

Details

Package: AllelicImbalance
Type: Package
Version: 1.2.0
Date: 2014-08-24
License: GPL-3

Overview - standard procedure

Start out creating a GRrange object defining the region of interest. This can also be done using getAreaFromGeneNames providing gene names as arguments. Then use BamImpGAList to import reads from that region and find potential SNPs using scanForHeterozygotes. Then retrieve the allele counts of heterozygote sites by the function getAlleleCount. With this data create an ASEset. At this point all pre-requisites for a 'basic' allele specific expression analysis is available. Two ways to go on could be to apply chisq.test or barplot on this ASEset object.

Author(s)

Author: Jesper Robert Gadin Author: Lasse Folkersen
Maintainer: Jesper Robert Gadin <j.r.gadin@gmail.com>

References

Reference to published application note (work in progress)

See Also

• code?ASEset
AnnotationWrappers

Description

These functions acts as wrappers to retrieve information from annotation database objects (AnnotationDb objects) or (transcriptDb objects).

Usage

getGenesFromAnnotation(
    OrgDb,
    GR,
    leftFlank = 0,
    rightFlank = 0,
    getUCSC = FALSE,
    verbose = FALSE
)

getGenesVector(OrgDb, GR, leftFlank = 0, rightFlank = 0, verbose = FALSE)

getExonsFromAnnotation(
    TxDb,
    GR,
    leftFlank = 0,
    rightFlank = 0,
    verbose = FALSE
)

getExonsVector(TxDb, GR, leftFlank = 0, rightFlank = 0, verbose = FALSE)

getTranscriptsFromAnnotation(
    TxDb,
    GR,
    leftFlank = 0,
    rightFlank = 0,
    verbose = FALSE
)

getTranscriptsVector(TxDb, GR, leftFlank = 0, rightFlank = 0, verbose = FALSE)

getCDSFromAnnotation(TxDb, GR, leftFlank = 0, rightFlank = 0, verbose = FALSE)

getCDSVector(TxDb, GR, leftFlank = 0, rightFlank = 0, verbose = FALSE)

getAnnotationDataFrame(
    GR,
strand = "+",
annotationType = NULL,
OrgDb = NULL,
TxDb = NULL,
verbose = FALSE
)

Arguments

OrgDb An OrgDb object
GR A GenomicRanges object with sample area
leftFlank An integer specifying number of additional nucleotides around the SNPs for the leftFlank
rightFlank An integer specifying number of additional nucleotides around the SNPs for the rightFlank
getUCSC A logical indicating if UCSC transcript IDs should also be retrieved
verbose A logical making the functions more talkative
TxDb A transcriptDb object
strand Two options; ‘+’ or ‘-’
annotationType select one or more from 'gene', 'exon', 'transcript', 'cds'.

Details

These functions retrieve regional annotation from OrgDb or TxDb objects, when given GRanges objects.

Value

GRanges object with ranges over the genes in the region.
The getGenesVector function will return a character vector where each element are gene symbols separated by comma
GRanges object with ranges over the exons in the region.
The getTranscriptsFromAnnotation function will return a GRanges object with ranges over the transcripts in the region.
The getCDSFromAnnotation function will return a GRanges object with ranges over the CDSFs in the region.
The getExonsVector function will return a character vector where each element are exons separated by comma
The getTranscriptsVector function will return a character vector where each element are transcripts separated by comma
The getCDSVector function will return a character vector where each element are CDSs separated by comma
The getAnnotationDataFrame function will return a data.frame with annotations. This function is used internally by i.e. the barplot-function
Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

```r
data(ASEset)
require(org.Hs.eg.db)
require(TxDb.Hsapiens.UCSC.hg19.knownGene)
OrgDb <- org.Hs.eg.db
TxDb <- TxDb.Hsapiens.UCSC.hg19.knownGene

# use for example Bcflles as the source for SNPs of interest
GR <- rowRanges(ASEset)
# get annotation
g <- getGenesFromAnnotation(OrgDb,GR)
e <- getExonsFromAnnotation(TxDb,GR)
t <- getTranscriptsFromAnnotation(TxDb,GR)
c <- getCDSFromAnnotation(TxDb,GR)
```

---

**annotationBarplot**

*add annotation to AllelicImbalance barplot*

Description

adds a customizable annotation functionality for AllelicImbalance barplots.

Usage

```r
annotationBarplot(
  strand, 
  snp, 
  lowerLeftCorner, 
  annDfPlus, 
  annDfMinus, 
  cex = 0.7, 
  ypos = 0, 
  interspace = 1
)
```

Arguments

- **strand**: `"+", ",-", "*" or "both"
- **snp**: integer for the described snp
- **lowerLeftCorner**: position of the plot to add legend to (default c(0,0))
annDfPlus annotation dataframe plus strand
annDfMinus annotation dataframe minus strand
cex size of annotation text
ypos relative y-axis position for the annotation text
interspace space between each annotation block

Details
the function is preferably called from within the AllelicImbalance barplot method.

Author(s)
Jesper R. Gadin

Examples

#code placeholders
#< create a barplot without annotation >
#< add annotation >

---

ASEset-barplot barplot ASEset objects

Description
Generates barplots for ASEset objects. Two levels of plotting detail are provided: a detailed barplot
of read counts by allele useful for fewer samples and SNPs, and a less detailed barplot of the fraction
of imbalance, useful for more samples and SNPs.

Usage

barplot(height, ...)

## S4 method for signature 'ASEset'
barplot(
  height,
  type = "count",
  sampleColour.top = NULL,
  sampleColour.bot = NULL,
  legend = TRUE,
  pValue = TRUE,
  strand = "*",
  testValue = NULL,
  testValue2 = NULL,
  OrgDb = NULL,
  TxDb = NULL,
Arguments

height  An ASEset object
... for simpler generics when extending function
type    'count' or 'fraction'
sampleColour.top  User specified colours for top fraction
sampleColour.bot   User specified colours for bottom fraction
legend         Display legend
pValue          Display p-value
strand          four options, '+', '-', 'both' or '*'
testValue       if set, a matrix or vector with user p-values
ASEset-barplot

testValue2  
OrgDb  
TxDb  
annotationType  
main  
ylim  
yaxis  
xaxis  
ylab  
ylab.text  
xlab.text  
xlab  
legend.colnames  
las.ylab  
las.xlab  
cex.main  
cex.pValue  
cex.ylab  
cex.xlab  
cex.legend  
add  
lowerLeftCorner  
size  
addHorizontalLine  
add.frame  
filter.pValue.fraction  
legend.fill.size  
legend.interspace  
verbose  
top.fraction.criteria  
cex.annotation  
ypos.annotation  
annotation.interspace

if set, a matrix or vector with user p-values
an OrgDb object which provides annotation
a TxDb object which provides annotation
select one or more from 'gene','exon','transcript','cds'.
text to use as main label
set plot y-axis limit
whether the y-axis is to be displayed or not
whether the x-axis is to be displayed or not
showing labels for the tic marks
ylab text
xlab text
showing labels for the tic marks
gives colnames to the legend matrix
orientation of ylab text
orientation of xlab text
set main label size (max 2)
set pValue label size
set ylab label size
set xlab label size
set legend label size
boolean indicates if a new device should be started
integer that is only useful when add=TRUE
Used internally by locationplot. Rescales each small barplot window
adds a horizontal line that marks the default fraction of 0.5 - 0.5
boolean to give the new plot a frame or not
numeric between 0 and 1 that filter away pValues where the main allele has this frequency.
size of the fill/boxes in the legend (default:NULL)
set legend space between fills and text
Makes function more talkative
'maxcount', 'ref' or 'phase'
size of annotation text
relative ypos for annotation text
space between annotation text
Details

filter.pValue.fraction is intended to remove p-value annotation with very large difference in frequency, which could just be a sequencing mistake. This is to avoid p-values like 1e-235 or similar.

sampleColourUser specified colours, either given as named colours (‘red’, ‘blue’, etc) or as hexadecimal code. Can be either length 1 for all samples, or else of a length corresponding to the number of samples for individual colouring.

Author(s)

Jesper R. Gadin, Lasse Folkersen

See Also

• The ASEset class which the barplot function can be called up on.

Examples

data(ASEset)
barplot(ASEset[1])

---

ASEset-class ASEset objects

Description

Object that holds allele counts, genomic positions and map-bias for a set of SNPs

Usage

alleleCounts(x, strand = "*", return.class = "list")

## S4 method for signature 'ASEset'
alleleCounts(x, strand = "*", return.class = "list")

alleleCounts(x, ...) <- value

## S4 replacement method for signature 'ASEset'
alleleCounts(x, strand = "*", return.class = "array", ...) <- value

mapBias(x, ...)

## S4 method for signature 'ASEset'
mapBias(x, return.class = "list")

fraction(x, ...)

fraction(x, strand = "*",
    top.fraction.criteria = "maxcount",
    verbose = FALSE,
    ...
)

arank(x, return.type = "names", return.class = "list", strand = "*", ...)

frequency(x, ...)

genotype(x, ...)

## S4 method for signature 'ASEset'
genotype(x, return.class = "matrix")

genotype(x) <- value

## S4 replacement method for signature 'ASEset'
genotype(x) <- value

countsPerSnp(x, ...)

## S4 method for signature 'ASEset'
countsPerSnp(x, return.class = "matrix", return.type = "mean", strand = "*")
countsPerSample(x, ...)

## S4 method for signature 'ASEset'
countsPerSample(x, return.class = "matrix", return.type = "mean", strand = "*")

phase(x, ...)

## S4 method for signature 'ASEset'
phase(x, return.class = "matrix")

phase(x) <- value

## S4 replacement method for signature 'ASEset'
phase(x) <- value

mapBias(x) <- value

## S4 replacement method for signature 'ASEset'
mapBias(x) <- value
refExist(x)

## S4 method for signature 'ASEset'
refExist(x)

ref(x)

## S4 method for signature 'ASEset'
ref(x)

ref(x) <- value

## S4 replacement method for signature 'ASEset,ANY'
ref(x) <- value

altExist(x)

## S4 method for signature 'ASEset'
altExist(x)

alt(x)

## S4 method for signature 'ASEset'
alt(x)

alt(x) <- value

## S4 replacement method for signature 'ASEset,ANY'
alt(x) <- value

aquals(x, ...)

## S4 method for signature 'ASEset'
aquals(x)

aquals(x) <- value

## S4 replacement method for signature 'ASEset'
aquals(x) <- value

maternalAllele(x, ...)

## S4 method for signature 'ASEset'
maternalAllele(x)

paternalAllele(x, ...)

## S4 method for signature 'ASEset'
paternalAllele(x)
## S4 method for signature 'ASEset'
paternalAllele(x)

### Arguments

- **x**: ASEset object
- **strand**: which strand of '+', '-' or '*'
- **return.class**: return 'list' or 'array'
- **...**: additional arguments
- **value**: replacement variable
- **top.fraction.criteria**: 'maxcount', 'ref' or 'phase'
- **verbose**: makes function more talkative
- **return.type**: return 'names', rank or 'counts'

### Details

An ASEset object differs from a regular RangedSummarizedExperiment object in that the assays contains an array instead of matrix. This array has ranges on the rows, sampleNames on the columns and variants in the third dimension.

It is possible to use the commands barplot and locationplot on an ASEset object see more details in barplot and locationplot.

Three different alleleCount options are available. The simples one is the * option, and is for experiments where the strand information is not known e.g. non-stranded data. The unknown strand could also be for strand specific data when the aligner could not find any strand associated with the read, but this should normally not happen, and if it does probably having an extremely low mapping quality. Then there are an option too add plus and minus stranded data. When using this, it is essential to make sure that the RNA-seq experiment under analysis has in fact been created so that correct strand information was obtained. The most functions will by default have their strand argument set to '*'.

The phase information is stored by the convention of 'maternal chromosome|paternal chromosome', with 0 as reference allele and 1 as alternative allele. '|' when the phase is known and '/' when the phase is unknown. Internally the information will be stored as an three dimensional array, dim 1 for SNPs, dim 2 for Samples and dim 3 which is fixed and stores maternal chromosome, paternal chromosome and phased (1 equals TRUE).

### Value

An object of class ASEset containing location information and allele counts for a number of SNPs measured in a number of samples on various strand, as well as mapBias information. All data is stored in a manner similar to the SummarizedExperiment class.

### Table

```r
table(...)
```

Arguments:
... An ASEset object that contains the variants of interest

The generics for table does not easily allow more than one argument so in respect to the different strand options, table will return a SimpleList with length 3, one element for each strand.

**Frequency**

frequency(x, return.class = "list", strand = "*", threshold.count.sample = 15)

Arguments:

- *x* An ASEset object that contains the variants of interest
- *threshold.count.sample* if sample has fewer counts the function return NA.

**Constructor**

ASEsetFromCountList(rowRanges, countListNonStranded = NULL, countListPlus = NULL, countListMinus = NULL, countListUnknown = NULL, colData = NULL, mapBiasExpMean = array(), verbose=FALSE, ...)

Arguments:

- **rowRanges** A GenomicRanges object that contains the variants of interest
- **countListNonStranded** A list where each entry is a matrix with allele counts as columns and sample counts as rows
- **countListPlus** A list where each entry is a matrix with allele counts as columns and sample counts as rows
- **countListMinus** A list where each entry is a matrix with allele counts as columns and sample counts as rows
- **countListUnknown** A list where each entry is a matrix with allele counts as columns and sample counts as rows
- **colData** A DataFrame object containing sample specific data
- **mapBiasExpMean** A 3D array describing mapping bias. The SNPs are in the 1st dimension, samples in the 2nd dimension and variants in the 3rd dimension.
- **verbose** Makes function more talkative
- ... arguments passed on to SummarizedExperiment constructor

**Author(s)**

Jesper R. Gadin, Lasse Folkersen

**Examples**

```r
# make example countList
set.seed(42)
countListPlus <- list()
snps <- c('snp1','snp2','snp3','snp4','snp5')
for(snp in snps){
  count<-matrix(rep(0,16),ncol=4,dimnames=list(}
c('sample1', 'sample2', 'sample3', 'sample4'),
c('A', 'T', 'G', 'C'))
# insert random counts in two of the alleles
for(allele in sample(c('A', 'T', 'G', 'C'), 2)) {
  count[, allele] <- as.integer(rnorm(4, mean = 50, sd = 10))
}
countListPlus[[snp]] <- count
}

# make example rowRanges
rowRanges <- GRanges(
  seqnames = Rle(c('chr1', 'chr2', 'chr3', 'chr1', 'chr1', 'chr3', 'chr1'),
                 ranges = IRanges(1:5, width = 1, names = head(letters, 5)),
                 snp = paste('snp', 1:5, sep = ''))
)

# make example colData
colData <- DataFrame(Treatment = c('ChIP', 'Input', 'Input', 'ChIP'),
                      row.names = c('ind1', 'ind2', 'ind3', 'ind4'))

# make ASEset
a <- ASEsetFromCountList(rowRanges, countListPlus = countListPlus,
                          colData = colData)

# example phase matrix (simple form)
p1 <- matrix(sample(c(1, 0), replace = TRUE, size = nrow(a) * ncol(a)),
              nrow = nrow(a), ncol = a)
p2 <- matrix(sample(c(1, 0), replace = TRUE, size = nrow(a) * ncol(a)),
              nrow = nrow(a), ncol = a)
p <- matrix(paste(p1, sample(c('|', '|', '/', size = nrow(a) * ncol(a), replace = TRUE),
                          p2, sep = ''),
               nrow = nrow(a), ncol = a))
phase(a) <- p

# generate ASEset from array
snps <- 999
samples <- 5
ar <- array(rep(unlist(lapply(1:snps, function(x) {
  sample(c(TRUE, FALSE, TRUE, FALSE), size = 4)
})), samples),
            dim = c(4, snps, samples))
ar2 <- array(sample(50:300, 4*snps*samples, replace = TRUE),
             dim = c(4, snps, samples))
ar2[ar] <- 0
ar2 <- aperm(ar2, c(2, 3, 1))
dimnames(ar2) <- list(paste('snp', 1:snps, sep = ''),
                       paste('sample', 1:samples, sep = ''),
                       c('A', 'C', 'G', 'T'))
gr <- GRanges(seqnames = c('chr2'), ranges = IRanges(start = 1:dim(ar2)[1], width = 1),
              strand = '*')
a <- ASEsetFromArrays(gr, countsUnknown = ar2)
Description

useful genotype filters

Usage

hetFilt(x, ...)

## S4 method for signature 'ASEset'
hetFilt(x, source = "genotype", ...)

Arguments

x  ASEset object
...
source  'genotype' or 'alleleCounts'

Details

hetFilt returns TRUE if the samples is heterozygote, based on stored genotype information present in the phase data.

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

#load example data
data(ASEset)
a <- ASEset
genotype(a) <- inferGenotypes(a)
hets <- hetFilt(a)
ASEset-gbarplot

gbarplot ASEset objects

Description

Generates gbarplots for ASEset objects. Two levels of plotting detail are provided: a detailed gbarplot of read counts by allele useful for fewer samples and SNPs, and a less detailed gbarplot of the fraction of imbalance, useful for more samples and SNPs.

Usage

gbarplot(x, type = "count", strand = "*", verbose = FALSE, ...)

Arguments

<table>
<thead>
<tr>
<th>x</th>
<th>An ASEset object</th>
</tr>
</thead>
<tbody>
<tr>
<td>type</td>
<td>'count' or 'fraction'</td>
</tr>
<tr>
<td>strand</td>
<td>four options, '+' , '-', 'both' or '*'</td>
</tr>
<tr>
<td>verbose</td>
<td>Makes function more talkative</td>
</tr>
<tr>
<td>...</td>
<td>for simpler generics when extending function</td>
</tr>
</tbody>
</table>

Details

This function serves the same purpose as the normal barplot, but with trellis graphics using lattice, to be able to integrate well with Gviz track functionality.

Author(s)

Jesper R. Gadin

See Also

- The ASEset class which the gbarplot function can be called up on.
- The barplot non trellis barplot.

Examples

data(ASEset)
gbarplot(ASEset[1])
**ASEset-glocationplot**

**glocationplot ASEset objects**

**Description**

plotting ASE effects over a specific genomic region using Gviz functionality

**Usage**

```r
glocationplot(
  x,
  type = "fraction",
  strand = "*",
  BamGAL = NULL,
  GenomeAxisTrack = FALSE,
  trackNameDeAn = paste("deTrack", type),
  TxDb = NULL,
  sizes = NULL,
  add = FALSE,
  verbose = FALSE,
  ...
)
```

**Arguments**

- `x`: an ASEset object.
- `type`: 'fraction' or 'count'
- `strand`: '+' or '-' or 'both'. This argument determines which strand is plotted. See `getAlleleCounts` for more information of choice of strand.
- `BamGAL`: GAlignmentsList covering the same genomic region as the ASEset
- `GenomeAxisTrack`: include an genomic axis track
- `trackNameDeAn`: trackname for deAnnotation track
- `TxDb`: a TxDb object which provides annotation
- `sizes`: vector with the sum 1. Describes the size of the tracks
- `add`: add to existing plot
- `verbose`: if set to TRUE it makes function more talkative
- `...`: arguments passed on to barplot function

**Details**

The glocationplot methods visualises the distribution of ASE over a larger region on one chromosome. It takes and ASEset object as well as additional information on plot type (see `gbarplot`), strand type (see `getAlleleCounts`), Annotation tracks are created from the Gviz package. It is obviously important to make sure that the genome build used is set correctly, e.g. 'hg19'. sizes has to be of the same length as the number of tracks used.
**Author(s)**

Jesper R. Gadin

**See Also**

- The `ASEset` class which the `glocationplot` function can be called up on.

**Examples**

```r
data(ASEset)
genome(ASEset) <- 'hg19'
glocationplot(ASEset,strand='+')

# for ASEsets with fewer SNPs the 'count' type plot is useful
glocationplot(ASEset,type='count',strand='+')
```

**Description**

plotting ASE effects over a specific genomic region

**Usage**

```r
ASEDAAnnotationTrack(
  x,
  GR = rowRanges(x),
  type = "fraction",
  strand = "*",
  trackName = paste("deTrack", type),
  verbose = TRUE,
  ...
)

## S4 method for signature 'ASEset'
ASEDAAnnotationTrack(
  x,
  GR = rowRanges(x),
  type = "fraction",
  strand = "*",
  trackName = paste("deTrack", type),
  verbose = TRUE,
  ...
)
```
CoverageDataTrack(
  x,
  GR = rowRanges(x),
  BamList = NULL,
  strand = NULL,
  start = NULL,
  end = NULL,
  trackNameVec = NULL,
  meanCoverage = FALSE,
  verbose = TRUE,
  ...
)

Arguments

  x      an ASEset object.
  GR     genomic range of plotting
  type   'fraction' or 'count'
  strand '+','-'. This argument determines which strand is plotted.
  trackName name of track (ASEDAnnotationTrack)
  verbose Setting verbose=TRUE gives details of procedure during function run
  ...    arguments passed on to barplot function
  BamList GAlignmnentsList object of reads from the same genomic region as the ASEset
  start   start position of reads to be plotted
  end     end position of reads to be plotted
  trackNameVec names of tracks (CoverageDataTrack)
  meanCoverage mean of coverage over samples (CoverageGataTrack)

Details

For information of how to use these tracks in more ways, visit the Gviz package manual.

Author(s)

Jesper R. Gadin

See Also

- The ASEset class which the functions can be called up on.
**Examples**

```r
data(ASEset)
x <- ASEset[,1:2]
r <- reads[1:2]
genome(x) <- 'hg19'
seqlevels(r) <- seqlevels(x)

GR <- GRanges(seqnames=seqlevels(x),
ranges=IRanges(start=min(start(x)),end=max(end(x))),
strand='+', genome=genome(x))
deTrack <- ASEDAnnotationTrack(x, GR=GR, type='fraction', strand='+')
covTracks <- CoverageDataTrack(x,BamList=r,strand='+')

lst <- c(deTrack,covTracks)
sizes <- c(0.5,rep(0.5/length(covTracks),length(covTracks)))
#temporarily do not run this function
#plotTracks(lst, from=min(start(x)), to=max(end(x)),
#sizes=sizes, col.line = NULL, showId = FALSE, main='mainText',
#cex.main=1, title.width=1, type='histogram')
```

---

**locationplot ASEset objects**

**Description**

plotting ASE effects over a specific genomic region

**Usage**

```r
locationplot(x, ...)
```

## S4 method for signature 'ASEset'

```r
locationplot(
  x,
  type = "fraction",
  strand = "*",
  yaxis = TRUE,
  xaxis = FALSE,
  xlab = FALSE,
  ylab = TRUE,
  xlab.text = "",
  ylab.text = "",
  legend.colnames = "",
  size = c(0.8, 1),
)
Arguments

x

an ASEset object.

... arguments passed on to barplot function

type

'fraction' or 'count'

strand

'+','-','*' or 'both'. This argument determines which strand is plotted. See getAlleleCounts for more information on strand.

yaxis

whether the y-axis is to be displayed or not

xaxis

whether the x-axis is to be displayed or not

xlab

showing labels for the tic marks

ylab

showing labels for the tic marks

xlab.text

xlab text

ylab.text

ylab text

legend.colnames

gives colnames to the legend matrix

size

will give extra space in the margins of the inner plots

main

text to use as main label

pValue

Display p-value

cex.main

set main label size

cex.ylab

set ylab label size

cex.legend

set legend label size

OrgDb

an OrgDb object from which to plot a gene map. If given together with argument TxDB this will only be used to extract genesymbols.

TxDB

a TxDb object from which to plot an exon map.

verbose

Setting verbose=TRUE gives details of procedure during function run

top.fraction.criteria

'maxcount', 'ref' or 'phase'

allow.whole.chromosome

logical, overrides 200kb region limit, defaults to FALSE
Details

The locationplot methods visualises how fractions are distributed over a larger region of genes on one chromosome. It takes and ASEset object as well as additional information on plot type (see \texttt{barplot}), strand type (see \texttt{getAlleleCounts}), colouring, as well as annotation. The annotation is taken either from the bioconductor OrgDb-sets, the TxDb sets or both. It is obviously important to make sure that the genome build used is the same as used in aligning the RNA-seq data.

Author(s)

Jesper R. Gadin, Lasse Folkersen

See Also

- The \texttt{ASEset} class which the locationplot function can be called up on.

Examples

```r

data(ASEset)
locationplot(ASEset)

#SNPs are plotted in the order in which they are found.
#This can be sorted according to location as follows:
locationplot(ASEset[order(start(rowRanges(ASEset)))]

#for ASEsets with fewer SNPs the 'count' type plot is
# useful for detailed visualization.
locationplot(ASEset,type='count',strand='*')
```

### scanForHeterozygotes

\texttt{scanForHeterozygotes}

Description

Identifies the positions of SNPs found in BamGR reads.

Usage

\texttt{scanForHeterozygotes(BamList, ...)}

```r

## S4 method for signature 'GAlignmentsList'
scanForHeterozygotes(
  BamList,
  minimumReadsAtPos = 20,
  maximumMajorAlleleFrequency = 0.9,
  minimumMinorAlleleFrequency = 0.1,
```
ASEset-scanForHeterozygotes

```r
minimumBiAllelicFrequency = 0.9,
verbose = TRUE,
```

Arguments

- **BamList**: A `GAlignmentsList` object
- **minimumReadsAtPos**: minimum number of reads required to call a SNP at a given position
- **maximumMajorAlleleFrequency**: maximum frequency allowed for the most common allele. Setting this parameter lower will minimise the SNP calls resulting from technical read errors, at the cost of missing loci with potential strong ASE.
- **minimumMinorAlleleFrequency**: minimum frequency allowed for the second most common allele. Setting this parameter higher will minimise the SNP calls resulting from technical read errors, at the cost of missing loci with potential strong ASE.
- **minimumBiAllelicFrequency**: minimum frequency allowed for the first and second most common allele. Setting a Lower value for this parameter will minimise the identification of loci with three or more alleles in one sample. This is useful if sequencing errors are suspected to be common.
- **verbose**: logical indicating if process information should be displayed

Details

This function scans all reads stored in a `GAlignmentsList` for possible heterozygote positions. The user can balance the sensitivity of the search by modifying the `minimumReadsAtPos`, `maximumMajorAlleleFrequency` and `minimumBiAllelicFrequency` arguments.

Value

`scanForHeterozygotes` returns a `GRanges` object with the SNPs for the `BamList` object that was used as input.

Author(s)

Jesper R. Gadin, Lasse Folkersen

See Also

- The `getAlleleCounts` which is a function that count the number of reads overlapping a site.
Examples

data(reads)
s <- scanForHeterozygotes(reads,verbose=FALSE)

ASEset.old  

Description

old version of an ASEset which needs to be updated

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

#load example data (Not Run)
data(ASEset.old)

ASEset.sim  

Description

ASEset with simulated data with SNPs within the first 200bp of chromosome 17, which is required to have example data for the refAllele function.

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

#load example data (Not Run)
data(ASEset.sim)
ASEsetFromBam

ASEset from bam file

Description

count alleles and create an ASEset direct from bam file instead of reading into R first.

Usage

ASEsetFromBam(gr, ...)

## S4 method for signature 'GRanges'
ASEsetFromBam(
  gr,
  pathToDir,
  PE = TRUE,
  flagsMinusStrand = c(83, 163),
  flagsPlusStrand = c(99, 147),
  strandUnknown = FALSE,
  ...
)

Arguments

gr GenomicRanges of SNPs to create ASEset for
...

passed on to ASEsetFromBam function

pathToDir Directory of bam files with index in same directory

PE if paired end or not (default: TRUE)

flagsMinusStrand flags that mark reads coming from minus strand

flagsPlusStrand flags that mark reads coming from plus strand

strandUnknown default: FALSE

Details

counts the alleles in a bam file based on GRanges positions.

Author(s)

Jesper R. Gadin
Examples

```r
data(GRvariants)
gr <- GRvariants
```

```r
# pathToDir <- system.file('inst/extdata/ERP000101_subset', package='AllelicImbalance')
#a <- ASEsetFromBam(gr, pathToDir)
```

Description

Generates lattice barplots for ASEset objects. Two levels of plotting detail are provided: a detailed barplot of read counts by allele useful for fewer samples and SNPs, and a less detailed barplot of the fraction of imbalance, useful for more samples and SNPs.

Usage

```r
barplotLatticeFraction(identifier, ...)
barplotLatticeCounts(identifier, ...)
```

Arguments

- `identifier`: the single snp name to plot
- `...`: used to pass on variables

Details

- `filter.pValue.fraction` is intended to remove p-value annotation with very large difference in frequency, which could just be a sequencing mistake. This is to avoid p-values like 1e-235 or similar.
- `sampleColour` user specified colours, either given as named colours ('red', 'blue', etc) or as hexadecimal code. Can be either length 1 for all samples, or else of a length corresponding to the number of samples for individual colouring.

Author(s)

Jesper R. Gadin, Lasse Folkesen

See Also

- The ASEset class which the barplot function can be called up on.
Examples

a <- ASEset
name <- rownames(a)[1]

barplotLatticeFraction(identifier=name, x=a, astrand="+")
barplotLatticeCounts(identifier=name, x=a, astrand="+")

Description

Performs a binomial test on an ASEset object.

Usage

```r
## S4 method for signature 'ASEset'
binom.test(x, n = "*")
```

Arguments

- `x` ASEset object
- `n` strand option

Details

the test can only be applied to one strand at the time.

Value

binom.test returns a matrix

Author(s)

Jesper R. Gadin, Lasse Folkersen

See Also

- The `chisq.test` which is another test that can be applied on an ASEset object.

Examples

```r
#load example data
data(ASEset)

#make a binomial test
binom.test(ASEset,"*")
```
Description
Performs a chisq.test on an ASEset object.

Usage
## S4 method for signature 'ASEset'
chisq.test(x, y = "*")

Arguments
- x: ASEset object
- y: strand option

Details
The test is performed on one strand in an ASEset object.

Value
chisq.test returns a matrix with the chisq.test P-value for each SNP and sample

Author(s)
Jesper R. Gadin, Lasse Folkesen

See Also
- The `binom.test` which is another test that can be applied on an ASEset object.

Examples
#load example data
data(ASEset)

#make a chi-square test on default non-stranded strand
chisq.test(ASEset)
Description

From a GAlignments calculate the real corresponding position for each read based on its cigar.

Usage

realCigarPosition.old(RleCigar, BpPos)
realCigarPositions.old(RleCigar)
realCigarPositionsList.old(RleCigarList)

Arguments

RleCigar   An Rle containing cigar information
BpPos      the absolute position on the chromosome of interest
RleCigarList  An RleList containing cigar information

Details

The main intention for these functions are to be the internal functions for scanForHeterozygotes and getAlleleCount.

Value

realCigarPosition returns the new position realCigarPositions returns a vector with the corrected positions to be subsetted from a read. realCigarPositionsList returns a list where each element i a vector with the corrected positions to be subsetted from a read.

Author(s)

Jesper R. Gadin

Examples

RleCigarList <- cigarToRleList('3M4I93M')
BpPos <- 5

newPos  <- realCigarPosition.old(RleCigar=RleCigarList[[1]], BpPos)
newPositions  <- realCigarPositions.old(RleCigar=RleCigarList[[1]])
newPositionsList  <- realCigarPositionsList.old(RleCigarList=RleCigarList)
countAllelesFromBam

alleleCounts from bam file

Description

count alleles before creating ASEs.

Usage

countAllelesFromBam(gr, ...)

## S4 method for signature 'GRanges'
countAllelesFromBam(
gr,  
pathToDir,  
flag = NULL,  
scanBamFlag = NULL,  
return.class = "array",  
verbose = TRUE,  
...  
)

Arguments

gr          GRanges that contains SNPs of interest
...          arguments to pass on
pathToDir    path to directory of bam files
flag         specify one flag to use as filter, default is no filtering. allowed flags are 99, 147, 83 and 163
scanBamFlag  set a custom flag to use as filter
return.class type of class for the returned object
verbose      makes function more talkative

Details

counts the alleles in a bam file based on GRanges positions.

Important excerpt from the details section of the internal applyPileups function: Regardless of 'param' values, the algorithm follows samtools by excluding reads flagged as unmapped, secondary, duplicate, or failing quality control.

Author(s)

Jesper R. Gadin
coverageMatrixListFromGAL

coverage matrix of GAlignmentsList

Description

Get coverage per nucleotide for reads covering a region

Usage

coverageMatrixListFromGAL(BamList, ...)

## S4 method for signature 'GAlignmentsList'
coverageMatrixListFromGAL(BamList, strand = "*", ignore.empty.bam.row = TRUE)

Arguments

BamList          GAlignmentsList containing reads over the region to calculate coverage
...              arguments to pass on
strand           strand has to be '+' or '-'
ignore.empty.bam.row

Details

a convenience function to get the coverage from a list of reads stored in GAlignmnetsList, and returns by default a list with one matrix, and information about the genomic start and stop positions.

Author(s)

Jesper R. Gadin

Examples

r <- reads
seqlevels(r) <- '17'
covMatList <- coverageMatrixListFromGAL(BamList=r, strand='+')
**Description**

Internal function that can draw gene regions on pre-specified surfaces. Necessary for the genomic-location plots.

**Usage**

```
decorateWithExons(x, exonsInRegion, xlim, ylim, chromosome)
```

**Arguments**

- `x` : ASEset object
- `exonsInRegion` : GRanges object with generegions. Can be obtained using `getExonsFromAnnotation`. Must contain a column 'tx_name'
- `xlim` : xlim values for the pre-specified surface
- `ylim` : ylim values for the pre-specified surface
- `chromosome` : character

**Details**

The main intention of this function is to be used when plotting several bar plots in the same window. This function add gene regions under the bars.

**Value**

`decorateWithExons` returns nothing, but draws genes

**Author(s)**

Jesper R. Gadin, Lasse Folkerse

**See Also**

- The `locationplot` which is uses this function internally.
- The `decorateWithGenes` which is another similar function that `locationplot` uses internally.

**Examples**

```
data(ASEset)
```
**decorateWithGenes**

---

**Description**

Internal function that can draw gene regions on pre-specified surfaces. Necessary for the genomic-location plots.

**Usage**

```r
decorateWithGenes(x, genesInRegion, xlim, ylim, chromosome)
```

**Arguments**

- `x` ASEset object
- `genesInRegion` GRanges object with gene regions. Can be obtained using `getGenesFromAnnotation`
- `xlim` xlim values for the pre-specified surface
- `ylim` ylim values for the pre-specified surface
- `chromosome` character

**Details**

The main intention of this function is to be used when plotting several bar plots in the same window. This function adds gene regions under the bars.

**Value**

`decorateWithGenes` returns nothing, but draws genes

**Author(s)**

Jesper R. Gadin, Lasse Folkersen

**See Also**

- The `locationplot` which is uses this function internally.
- The `decorateWithExons` which is another similar function that `locationplot` uses internally.

**Examples**

```r
data(ASEset)
```
defaultMapBias  

*Generate default mapbias from genotype*

**Description**

Create mapbias array from genotype matrix requires genotype information

**Usage**

```r
defaultMapBias(x, ...)  
```

```r
## S4 method for signature 'ASEset'
defaultMapBias(x, return.class = "array")
```

**Arguments**

- `x` : ASEset object
- `...` : internal arguments  
- `return.class` : "array" or "ASEset"

**Details**

Default mapbias will be 0.5 for bi-allelic snps and 1 for homozygots. For genotypes with NA, 0.5 will be placed on all four alleles. Therefore tri-allelic can not be used atm. Genotype information has to be placed in the genotype(x) assay.

**Author(s)**

Jesper R. Gadin, Lasse Folkersen

**Examples**

```r
#load example data
data(ASEset.sim)

fasta <- system.file('extdata/hg19.chr17.subset.fa', package='AllelicImbalance')
refAllele(ASEset.sim,fasta=fasta)
a <- refAllele(ASEset.sim,fasta=fasta)
```
defaultPhase

Description

used to populate the phase slot in an ASEset object

Usage

defaultPhase(i, ...)

## S4 method for signature 'numeric'
defaultPhase(i, j, ...)

Arguments

i       number of rows
...
arguments to forward to internal functions
j       number of columns

Details

will set everything to 0

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

i <- 5
j <- 10
defaultPhase(i, j)

detectAI

Description

detection of AllelicImbalance
detectAI

## Usage

detectAI(x, ...)

### S4 method for signature 'ASEset'

detectAI(
  x,
  return.class = "DetectedAI",
  strand = "*",
  threshold.frequency = 0,
  threshold.count.sample = 1,
  threshold.delta.frequency = 0,
  threshold.pvalue = 0.05,
  inferGenotype = FALSE,
  random.ref = FALSE,
  function.test = "binom.test",
  verbose = TRUE,
  gc = FALSE,
  biasMatrix = FALSE
)

### Arguments

- **x**: ASEset
- **...**: internal arguments
- **return.class**: class to return (atm only class 'logical')
- **strand**: strand to infer from
- **threshold.frequency**: least fraction to classify (see details)
- **threshold.count.sample**: least amount of counts to try to infer allele
- **threshold.delta.frequency**: minimum of frequency difference from 0.5 (or mapbias adjusted value)
- **threshold.pvalue**: pvalue over this number will be filtered out
- **inferGenotype**: infer genotypes based on count data in ASEset object
- **random.ref**: set the reference as random if you dont know. Affects interpretation of results.
- **function.test**: At the moment the only available option is 'binom.test'
- **verbose**: makes moment more talkative
- **gc**: use garbage collection when possible to save space
- **biasMatrix**: use biasMatrix in ASEset, or use default expected frequency of 0.5 for all sites

### Details

threshold.frequency is the least fraction needed to classify as bi tri or quad allelic SNPs. If 'all' then all of bi tri and quad allelic SNPs will use the same threshold. Everything under the treshold
will be regarded as noise. 'all' will return a matrix with snps as rows and uni bi tri and quad will be columns. For this function Anything that will return TRUE for tri-allelicwill also return TRUE for uni and bi-allelic for the same SNP an Sample.

return.type 'ref' return only AI when reference allele is more expressed. 'alt' return only AI when alternative allele is more expressed or 'all' for both 'ref' and 'alt' alleles. Reference allele is the one present in the reference genome on the forward strand.

threshold.delta.frequency and function.test will use the value in mapBias(x) as expected value. function.test will use the two most expressed alleles for testing. Make therefore sure there are no tri-allelic SNPs or somatic mutations among the SNPs in the ASEset.

inferGenotype(), set TRUE it should be used with as much samples as possible. If you split up the samples and run detectAI() on each sample separately, please make sure you have inferred the genotypes in before hand, alternatively used the genotypes detected by another variantCaller or chip-genotypes. Use ONLY biallelic genotypes.

Author(s)
Jesper R. Gadin

Examples

#load example data
data(ASEset)
a <- ASEset
dai <- detectAI(a)
thresholdCountSample(x, ...)

## S4 method for signature 'DetectedAI'
thresholdCountSample(x, return.class = "array")

thresholdDeltaFrequency(x, ...)

## S4 method for signature 'DetectedAI'
thresholdDeltaFrequency(x, return.class = "array")

thresholdPvalue(x, ...)

## S4 method for signature 'DetectedAI'
thresholdPvalue(x, return.class = "array")

### Arguments

- **x**: ASEset object or list of ASEsets
- **...**: pass arguments to internal functions
- **return.class**: type of class returned eg. "list" or "array".

### Details

The DetectedAI-class contains

### Author(s)

Jesper R. Gadin, Lasse Folkersen

### Examples

```r
data(ASEset)
a <- ASEset
dai <- detectAI(a)

#summary(gba)
#write.tables(dai)
```

---

**Description**

plot functions for the DetectedAI-class
Usage

frequency_vs_threshold_variable_plot(x, ...)

## S4 method for signature 'DetectedAI'
frequency_vs_threshold_variable_plot(
  x,
  var = "threshold.count.sample",
  hetOverlay = TRUE,
  smoothscatter = FALSE
)

detectedAI_vs_threshold_variable_plot(x, ...)

## S4 method for signature 'DetectedAI'
detectedAI_vs_threshold_variable_plot(
  x,
  var = "threshold.count.sample",
  summaryOverSamples = "sum",
  hetOverlay = TRUE,
  smoothscatter = FALSE
)

reference_frequency_density_vs_threshold_variable_plot(x, ...)

## S4 method for signature 'DetectedAI'
reference_frequency_density_vs_threshold_variable_plot(
  x,
  var = "threshold.count.sample"
)

detectedAI_vs_threshold_variable_multigraph_plot(x, ...)

## S4 method for signature 'DetectedAI'
detectedAI_vs_threshold_variable_multigraph_plot(x, ncol = 2, ...)

frequency_vs_threshold_variable_multigraph_plot(x, ...)

## S4 method for signature 'DetectedAI'
frequency_vs_threshold_variable_multigraph_plot(x, ncol = 2, ...)

reference_frequency_density_vs_threshold_variable_multigraph_plot(x, ...)

## S4 method for signature 'DetectedAI'
reference_frequency_density_vs_threshold_variable_multigraph_plot(
  x,
  ncol = 2,
  ...
)
Arguments

- `x` detectedAI object
- `...` pass on variables internally
- `var` string, see details for available options
- `hetOverlay` logical, if TRUE show nr of het SNPs used to calculate the reference allele frequency mean
- `smoothscatter` boolean, smoothscatter over the means
- `summaryOverSamples` 'mean' or 'sum'
- `ncol` nr of columns for multiplots

Details

Plot helper functions. The documentation will be improved before next release.

Author(s)

Jesper R. Gadin, Lasse Folkesen

Examples

```r
#some example code here
#generate example
data(ASEset)
a <- ASEset
dai <- detectAI(a,
threshold.count.sample=1:50,
threshold.frequency=seq(0,0.5,by=0.01),
threshold.delta.frequency=seq(0,0.5,by=0.01),
threshold.pvalue=rev(seq(0.001,0.05,by=0.005)) )

frequency_vs_threshold_variable_plot(dai)
detectedAI_vs_threshold_variable_plot(dai)
detectedAI_vs_threshold_variable_multigraph_plot(dai)
frequency_vs_threshold_variable_multigraph_plot(dai)
```

Description

Summary helper functions for the DetectedAI-class
Usage

```r
frequency_vs_threshold_variable_summary(x, ...)
```

```r
## S4 method for signature 'DetectedAI'
frequency_vs_threshold_variable_summary(
    x,
    var = "threshold.count.sample",
    return.class = "matrix",
    ...
)
```

```r
detectedAI_vs_threshold_variable_summary(x, ...)
```

```r
## S4 method for signature 'DetectedAI'
detectedAI_vs_threshold_variable_summary(x, var = "threshold.count.sample")
```

```r
usedSNPs_vs_threshold_variable_summary(x, ...)
```

```r
## S4 method for signature 'DetectedAI'
usedSNPs_vs_threshold_variable_summary(x, var = "threshold.count.sample")
```

Arguments

- `x`: detectedAI object
- `...`: pass on variables internally
- `var`: string, see details for available options
- `return.class`: 'matrix' or 'array'

Details

Summary helper functions. The documentation will be improved before next release.

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

```r
#some example code here
#generate example
data(ASEset)
a <- ASEset
dai <- detectAI(a,
    threshold.count.sample = 1:50,
    threshold.frequency = seq(0, 0.5, by = 0.01),
    threshold.delta.frequency = seq(0, 0.5, by = 0.01),
    threshold.pvalue = rev(seq(0.001, 0.05, by = 0.005))
)
```
fractionPlotDf

Plot Dataframe

Description
Summarizes information to ease creating plots

Usage
fractionPlotDf(x, snp, strand = "*", top.fraction.criteria = "maxcount", ...)

## S4 method for signature 'ASEset'
fractionPlotDf(x, snp, strand = "*", top.fraction.criteria = "maxcount", ...)

Arguments

x
ASEset

snp
rownames identifier for ASEset or row number

strand
'+', '-' or '*'

top.fraction.criteria
'maxcount', 'ref' or 'phase'

... arguments to forward to internal functions

Details
Main purpose is to reduce the amount of overall code and ease maintenance.

top.fraction.criteria can take three options, maxcount, ref and phase. The top allele will be every second row in the data frame, with start from row 2. The maxcount argument will put the allele with most reads on top of the bivariate fraction. Similarly the ref argument will put always the reference allele on top. The phase arguments puts the maternal phase always on top. The top.fraction.criteria for the ref or phase arguments requires that both ref and alt is set in mcols(ASEset).

Author(s)
Jesper R. Gadin, Lasse Folkersen

Examples
# test on example ASEset
data(ASEset)
a <- ASEset
df <- fractionPlotDf(a, 1, strand="+")
gba

global analysis wrapper

Description
A wrapper to make a global analysis based on paths for BAM, VCF and GFF files

Usage
gba(pathBam, ...)

## S4 method for signature 'character'
gba(pathBam, pathVcf, pathGFF = NULL, verbose)

Arguments
- pathBam: path to bam file
- ...: arguments to pass on
- pathVcf: path to vcf file
- pathGFF: path to gff file
- verbose: makes function more talkative

Author(s)
Jesper R. Gadin

Examples
empty as function doesn’t exist

genomatrix

genomatrix object

Description
genomatrix is an example of a matrix with genotypes

Author(s)
Jesper R. Gadin, Lasse Folkersen

Examples
#load example data (Not Run)
data(genomatrix)
Description

used to convert the genomatrix from the visually friendly matrix to phase array.

Usage

`genotype2phase(x, ...)

## S4 method for signature 'matrix'

`genotype2phase(

  x,
  ref = NULL,
  return.class = "array",
  levels = c("A", "C", "G", "T"),
  ...
)

Arguments

- `x`: matrix see examples
- `...`: pass on additional param
- `ref`: reference alleles
- `return.class`: 'array' or 'list'
- `levels`: vector of expected alleles

Details

To not introduce redundant information in the ASEset object, the genotype matrix is translated to a phase matrix, containing the same information. Does not allow tri-allelic or multi-allelic SNPs, and if present the multi-allelic SNPs will lose the least occurring genotype.

This function can handle indels, but if the reference allele is not provided, the rank matrix which is temporary created might use lots of memory, depending on the amount of indels among the genotypes. As conclusion, it is preferable to send in reference genome when converting to phase.

levels information is only important if the reference allele has to be guessed, and so if reference information is provided, the levels argument can be ignored.

Author(s)

Jesper R. Gadin, Lasse Folkersen
Examples

```r
#load example data
data(genomatrix)
data(ASEset)
p <- genotype2phase(genomatrix, ref(ASEset))
```

---

**getAlleleCounts**

**snp count data**

---

**Description**

Given the positions of known SNPs, this function returns allele counts from a BamGRL object.

**Usage**

```r
getAlleleCounts(BamList, ...)  
## S4 method for signature 'GAlignmentsList'
getAlleleCounts(
  BamList,
  GRvariants,
  strand = "*",
  return.class = "list",
  verbose = TRUE,
  ...
)
```

**Arguments**

- **BamList**
  - A GAlignmentsList object or GRangesList object containing data imported from a bam file.
- **...**
  - Parameters to pass on.
- **GRvariants**
  - A GRanges object that contains positions of SNPs to retrieve.
- **strand**
  - A length 1 character with value `'+', '-'`, or `'*'`. This argument determines if `getAlleleCounts` will retrieve counts from all reads, or only from reads marked as `'+', '-'` or `'*'` (unknown), respectively.
- **return.class**
  - 'list' or 'array'.
- **verbose**
  - Setting `verbose=TRUE` makes function more talkative.
getAlleleCounts

Details

This function is used to retrieve the allele counts from specified positions in a set of RNA-seq reads. The BamList argument will typically have been created using the impBamGAL function on bam-files. The GRvariants is either a GRanges with user-specified locations or else it is generated through scanning the same bam-files as in BamList for heterozygote locations (e.g. using scanForHeterozygotes). The GRvariants will currently only accept locations having width=1, corresponding to bi-allelic SNPs. In the strand argument, specifying '*' is the same as retrieving the sum count of '+' and '-' reads (and unknown strand reads in case these are found in the bam file). '*' is the default behaviour and can be used when the RNA-seq experiments strand information is not available.

Value

getAlleleCounts returns a list of several data.frame objects, each storing the count data for one SNP.

Author(s)

Jesper R. Gadin, Lasse Folkersen

See Also

- The scanForHeterozygotes which is a function to find possible heterozygote sites in a GenomicAlignments object

Examples

#load example data
data(reads)
data(GRvariants)

#get counts at the three positions specified in GRvariants
alleleCount <- getAlleleCounts(BamList=reads,GRvariants, strand='*')

#if the reads had contained stranded data, these two calls would
#have given the correct input objects for getAlleleCounts
alleleCountPlus <- getAlleleCounts(BamList=reads,GRvariants, strand='+')
alleleCountMinus <- getAlleleCounts(BamList=reads,GRvariants, strand='-')
getAlleleQuality

**Description**

Given the positions of known SNPs, this function returns allele quality from a BamGRL object.

**Usage**

```r
getAlleleQuality(BamList, ...)
```

## S4 method for signature 'GAlignmentsList'
```r
getAlleleQuality(
  BamList,
  GRvariants,
  fastq.format = "illumina.1.8",
  return.class = "array",
  verbose = TRUE,
  ...
)
```

**Arguments**

- `BamList` A `GAlignmentsList` object or `GRangesList` object containing data imported from a bam file.
- `...` Parameters to pass on.
- `GRvariants` A `GRanges` object that contains positions of SNPs to retrieve.
- `fastq.format` Default "illumina.1.8".
- `return.class` 'list' or 'array'.
- `verbose` Setting `verbose=TRUE` makes function more talkative.

**Details**

This function is used to retrieve the allele quality strings from specified positions in a set of RNA-seq reads. The `BamList` argument will typically have been created using the `impBamGAL` function on bam-files. The `GRvariants` is either a `GRanges` with user-specified locations or else it is generated through scanning the same bam-files as in `BamList` for heterozygote locations (e.g. using `scanForHeterozygotes`). The `GRvariants` will currently only accept locations having width=1, corresponding to bi-allelic SNPs. The strand type information will be kept in the returned object. If the strand is marked as unknown "*", it will be forced to the "+" strand.

Quality information is extracted from the `BamList` object, and requires the presence of `mcols(BamList)["qual"]` to contain quality sequences.

**Value**

`getAlleleQuality` returns a list of several data.frame objects, each storing the count data for one SNP.
Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

#load example data
data(reads)
data(GRvariants)

#get counts at the three positions specified in GRvariants
alleleQualityArray <- getAlleleQuality(BamList=reads,GRvariants)

#place in ASEset object
alleleCountsArray <- getAlleleCounts(BamList=reads,GRvariants,
strand='*', return.class="array")

a <- ASEsetFromArrays(GRvariants, countsUnknown = alleleCountsArray)
aquals(a) <- alleleQualityArray

getAreaFromGeneNames  Get Gene Area

Description

Given a character vector with genesymbols and an OrgDb object, this function returns a GRanges giving the coordinates of the genes.

Usage

getAreaFromGeneNames(genesymbols, ...)

## S4 method for signature 'character'
getAreaFromGeneNames(
  genesymbols, 
  OrgDb, 
  leftFlank = 0, 
  rightFlank = 0, 
  na.rm = FALSE, 
  verbose = TRUE 
)

Arguments

genesymbols A character vector that contains genesymbols of genes from which we wish to retrieve the coordinates

... arguments to pass on

OrgDb An OrgDb object containing gene annotation
getDefaultMapBiasExpMean

`getDefaultMapBiasExpMean`  

```r
getDefaultMapBiasExpMean
```

Description

an allele frequency array

Usage

```r
genericMapBiasExpMean(alleleCountList, ...)  
genericMapBiasExpMean3D(alleleCountList, ...)
```

## S4 method for signature 'list'
```r  
genericMapBiasExpMean(alleleCountList)  
```
## S4 method for signature 'ANY'
getDefaultMapBiasExpMean3D(alleleCountList)

### Arguments

- **alleleCountList**
  A GRangesList object containing read information

- **...**
  parameters to pass on

### Details
This function will assume there is no bias that comes from the mapping of reads, and therefore create a matrix with expected frequency of 0.5 for each allele.

### Value

`getDefaultMapBiasExpMean` returns a matrix with a default expected mean of 0.5 for every element.

### Author(s)

Jesper R. Gadin, Lasse Folkersen

### Examples

```r
# load example data
data(ASEset)
# access SnpAflist
alleleCountList <- alleleCounts(ASEset)
# get default map bias exp mean
matExpMean <- getDefaultMapBiasExpMean(alleleCountList)
```

---

### Description

Get rsIDs from locations of SNP

Given a GRanges object of SNPs and a SNPlocs annotation, this function attempts to replace the names of the GRanges object entries with rs-IDs.

### Usage

```r
getSnpIdFromLocation(GR, ...)
```

```r
## S4 method for signature 'GRanges'
getSnpIdFromLocation(GR, SNPloc, return.vector = FALSE, verbose = TRUE)
```
Arguments

- **GR**
  A GRanges that contains positions of SNPs to look up
- **SNPloc**
  A SNPlocs object containing information on SNP locations (e.g. SNPlocs.Hsapiens.dbSNP144.GRCh37)
- **return.vector**
  Setting return.vector=TRUE returns vector with rsIds
- **verbose**
  Setting verbose=TRUE makes function more talkative

Details

This function is used to try to identify the rs-IDs of SNPs in a GRanges object.

Value

getSnpIdFromLocation returns the same GRanges object it was given with, but with updated with rs.id information.

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

```r
is_32bit_windows <- .Platform$OS.type == "windows" &&
  .Platform$r_arch == "i386"
if (!is_32bit_windows && require(SNPlocs.Hsapiens.dbSNP144.GRCh37)) {
  #load example data
data(ASEset)

  #get counts at the three positions specified in GRvariants
  updatedGRanges <- getSnpIdFromLocation(rowRanges(ASEset),
    SNPlocs.Hsapiens.dbSNP144.GRCh37)
}
```

GlobalAnalysis-class

GlobalAnalysis class

Description

Object that holds results from a global AI analysis including reference bias estimations and AI detection.

Arguments

- **x**
  ASEset object or list of ASEsets
- **TxDb**
  A transcriptDb object
- **...**
  pass arguments to internal functions
**Details**

The GlobalAnalysis-class contains summaries and "pre-configured and pre-calculated lattice plots" needed to create an AI-report.

**Value**

An object of class GlobalAnalysis containing all data to make report.

**Author(s)**

Jesper R. Gadin, Lasse Folkersen

**Examples**

```r
data(ASEset)
#a <- ASEset
#gba <- gba(a)

#report(gba)
#write.tables(gba)
#graphs(gba)
#as.list(gba)
```

---

**GRvariants**

**GRvariants object**

**Description**

This data is a GRanges object that contains the ranges for three example SNPs.

**Author(s)**

Jesper R. Gadin, Lasse Folkersen

**See Also**

- The reads which is another example object

**Examples**

```r
#load example data
data(GRvariants)
```
histplot

histplot

histogram plots

Description

uses base graphics hist plot

Usage

## S4 method for signature 'ASEset'
hist(x, strand = "*", type = "mean", log = 1, ...)

Arguments

x ReferenceBias object or ASEset object
strand '+', '-', or '*'
type 'mean' (only one option atm)
log an integer to log each value (integer 10 for log10)
... arguments to forward to interal boxplots function

Details

The histogram will show the density over frequencies for each sample

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

###load example data

#data(ASEset)
#a <- ASEset
#hist(a)
import-bam

implodeList.old  implode list of arguments into environment

Description

apply on list of variables to be put in the local environment

Usage

implodeList.old(x)

Arguments

x  list of variables

Details

help the propagation of e.g. graphical parameters

Author(s)

Jesper R. Gadin

Examples

lst <- list(hungry='yes', thirsty='no')
implodeList.old(lst)
#the check ls()
ls()

import-bam  Import Bam

Description

Imports a specified genomic region from a bam file using a GRanges object as search area.

Usage

impBamGAL(UserDir, ...)

## S4 method for signature 'character'
impBamGAL(
  UserDir,
  searchArea,
  files = NULL,
import-bam

XStag = FALSE,
verbose = TRUE,
...
)

Arguments

UserDir  The relative or full path of folder containing bam files.

... arguments to pass on

searchArea  A GenomicRanges object that contains the regions of interest

files  use character vector to specify one or more files to import. The default imports all bam files from the directory.

XStag  Setting XStag=TRUE stores the strand specific information in the mcols slot 'XS'

verbose  makes the function more talkative.

Details

If the sequence data is strand-specific you may want to set XStag=TRUE. The strand specific information has then to be stored in the meta columns with column name 'XS'. If the aligner did not set the XS-tag and the data is strand-specific it is still be possible to infer the strand from the bit flags after importing the reads to R. Depending on the strand-specific protocol different combinations of the flags will have to be used. For illumina fr-secondstrand, 83 and 163 are minus strand reads and 99 and 147 are plus strand reads.

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

#Declare searchArea
searchArea <- GRanges(seqnames=c('17'), ranges=IRanges(79478301,79478361))

#Relative or full path
pathToFiles <- system.file('extdata/ERP000101_subset', package='AllelicImbalance')

#all files in directory
reads <- impBamGAL(pathToFiles, searchArea, verbose=FALSE)
#specified files in directory
reads <- impBamGAL(pathToFiles, searchArea, files=c("ERR009160.bam", "ERR009167.bam"), verbose=FALSE)
**Description**

Imports bla bal bal a specified genomic region from a bam file using a GenomicRanges object as search area.

**Usage**

```r
impBamGRL.old(UserDir, searchArea, verbose = TRUE)
```

**Arguments**

- **UserDir**: The relative or full path of folder containing bam files.
- **searchArea**: A GenomicRanges object that contains the regions of interest.
- **verbose**: Setting verbose=TRUE gives details of procedure during function run.

**Details**

These functions are right on tahea wrappers to import bam files into R and store them into either GRanges, GAlignments or GappedAlignmentpairs objects.

It is recommended to use the impBamGAL() which takes information of gaps into account. It is also possible to use the other variants as well, but then pre-filtering becomes important keeps to understand because gapped, intron-spanning reads will cause problems. This is because the GRanges objects can not handle if gaps are present and will then give a wrong result when calculating the allele (SNP) count table.

**Value**

- `impBamGRL` returns a GRangesList object containing the RNA-seq reads in the region defined by the searchArea argument.
- `impBamGAL` returns a list with GAlignments objects containing the RNA-seq reads in the region defined by the searchArea argument.
- `funImpBamGAPL` returns a list with GappedAlignmentPairs object containing the RNA-seq reads in the region defined by the searchArea argument.

**Author(s)**

Jesper R. Gadin, Lasse Folkersen

**Examples**

```r
# Declare searchArea
searchArea <- GRanges(seqnames=c('17'), ranges=IRanges(79478301,79478361))

# Relative or full path
pathToFiles <- system.file('extdata/ERP000101_subset', package='AllelicImbalance')
```
**import-bcf**

**Import Bcf Selection**

**Description**
Imports a selection of a bcf file or files specified by a GenomicRanges object as search area.

**Usage**

```r
impBcfGRL(UserDir, ...)

## S4 method for signature 'character'
impBcfGRL(UserDir, searchArea = NULL, verbose = TRUE, ...)

impBcfGR(UserDir, ...)

## S4 method for signature 'character'
impBcfGR(UserDir, searchArea = NULL, verbose = TRUE, ...)
```

**Arguments**

- `UserDir` The relative or full path of folder containing bam files.
- `...` parameters to pass on
- `searchArea` A GenomicRanges object that contains the regions of interest
- `verbose` Setting verbose=TRUE gives details of the procedure during function run.

**Details**
A wrapper to import bcf files into R in the form of GenomicRanges objects.

**Value**

- `BcfImpGRList` returns a GRangesList object. `BcfImpGR` returns one GRanges object of all unique entries from one or more bcf files.

**Note**
Make sure there is a complementary index file `*.bcf.csi` for each bcf file in `UserDir`. If there is not, then the functions `impBcfGRL` and `impBcfGR` will try to create them.

**Author(s)**
Jesper R. Gadin, Lasse Folkersen
See Also

- The `impBamGRL` for importing bam files
- The `getAlleleCounts` for how to get allele(SNP) counts
- The `scanForHeterozygotes` for how to find possible heterozygote positions

Examples

```r
# Declare searchArea
searchArea <- GRanges(seqnames=c('17'), ranges=IRanges(79478301, 79478361))

# Relative or full path
pathToFiles <- system.file('extdata/ERP000101_subset', package='AllelicImbalance')

# Import
reads <- impBcfGRL(pathToFiles, searchArea, verbose=FALSE)
```

## inferAlleles

### inference of SNPs of ASEset

#### Description

Inference of SNPs

#### Usage

```r
inferAlleles(
  x,
  strand = "*",
  return.type = "bi",
  threshold.frequency = 0,
  threshold.count.sample = 1,
  inferOver = "eachSample",
  allow.NA = FALSE
)
```

#### Arguments

- `x`: ASEset
- `strand`: strand to infer from
- `return.type`: 'uni' 'bi' 'tri' 'quad' 'all'
- `threshold.frequency`: least fraction to classify (see details)
- `threshold.count.sample`: least amount of counts to try to infer allele
- `inferOver`: 'eachSample' or 'allSamples'
- `allow.NA`: treat NA as zero when TRUE
**inferAltAllele**

**Details**
threshold.frequency is the least fraction needed to classify as bi tri or quad allelic SNPs. If 'all' then all of bi tri and quad allelic SNPs will use the same threshold. Everything under the threshold will be regarded as noise. 'all' will return a matrix with snps as rows and uni bi tri and quad will be columns. For this function Anything that will return TRUE for tri-allelic will also return TRUE for uni and bi-allelic for the same SNP an Sample.

**Author(s)**
Jesper R. Gadin

**Examples**

```r
data(ASEset)
i <- inferAlleles(ASEset)
```

---

**inferAltAllele**

<table>
<thead>
<tr>
<th>inferAltAllele</th>
<th>inferAltAllele</th>
</tr>
</thead>
</table>

**Description**
inference of the alternate allele based on count data

**Arguments**
- **x** matrix see examples
- **return.class** class of returned object
- **allele.source** 'arank'
- **verbose** make function more talkative
- **...** arguments to forward to internal functions

**Details**
The inference essentially ranks all alleles and the most expressed allele not declared as reference will be inferred as the alternative allele. At the moment only inference of bi-allelic alternative alleles are available.

**Author(s)**
Jesper R. Gadin, Lasse Folkersen
Examples

# load data
data(ASEset)

alt <- inferAltAllele(ASEset)

inferGenotypes

inference of genotypes from ASEset count data

Description

inference of genotypes

Usage

inferGenotypes(
  x,
  strand = "*",
  return.class = "matrix",
  return.allele.allowed = "bi",
  threshold.frequency = 0,
  threshold.count.sample = 1
)

Arguments

x      ASEset
strand strand to infer from
return.class 'matrix' or 'vector'
return.allele.allowed
  vector with 'bi' 'tri' or 'quad'. 'uni' Always gets returned
threshold.frequency least fraction to classify (see details)
threshold.count.sample least amount of counts to try to infer allele

Details

Often necessary information to link AI to SNPs outside coding region

Author(s)

Jesper R. Gadin
Examples

data(ASEset)
g <- inferGenotypes(ASEset)

initialize-ASEset

Initialize ASEset

Description

Functions to construct ASEset objects

Usage

ASEsetFromCountList(
  rowRanges,
  countListUnknown = NULL,
  countListPlus = NULL,
  countListMinus = NULL,
  colData = NULL,
  mapBiasExpMean = NULL,
  phase = NULL,
  aquals = NULL,
  verbose = FALSE,
  ...
)

ASEsetFromArrays(
  rowRanges,
  countsUnknown = NULL,
  countsPlus = NULL,
  countsMinus = NULL,
  colData = NULL,
  mapBiasExpMean = NULL,
  phase = NULL,
  genotype = NULL,
  aquals = NULL,
  verbose = FALSE,
  ...
)

Arguments

rowRanges A GenomicRanges object that contains the variants of interest
countListUnknown A list where each entry is a matrix with allele counts as columns and sample counts as rows
countListPlus  A list where each entry is a matrix with allele counts as columns and sample counts as rows

countListMinus A list where each entry is a matrix with allele counts as columns and sample counts as rows

colData A DataFrame object containing sample specific data

cmapBiasExpMean A 3D array where the SNPs are in the 1st dimension, samples in the 2nd dimension and variants in the 3rd dimension.

phase A matrix or an array containing phase information.

aquals A 4-D array containing the count information, see details

verbose Makes function more talkative

... arguments passed on to SummarizedExperiment constructor

countsUnknown An array containing the count information

countsPlus An array containing the count information

countsMinus An array containing the count information

genotype matrix

Details

The resulting ASEset object is based on the RangedSummarizedExperiment class, and will therefore inherit the same accessors and ranges operations.

If both countListPlus and countListMinus are given they will be used to calculate countListUnknown, which is the sum of the plus and minus strands.

countListPlus, countListMinus and countListUnknown are i.e. the outputs from the getAlleleCounts function.

aquals is new for the devel branch and will be changed slightly before the relase to include better granularity.

Value

ASEsetFromCountList returns an ASEset object.

Note

ASEsetFromCountList requires the same input data as a RangedSummarizedExperiment, but with minimum one assay for the allele counts.

Author(s)

Jesper R. Gadin, Lasse Folkersen
Examples

#make example alleleCountListPlus
set.seed(42)
countListPlus <- list()
snps <- c('snp1', 'snp2', 'snp3', 'snp4', 'snp5')
for(snp in snps){
  count <- matrix(rep(0, 16), ncol=4, dimnames=list(c('sample1', 'sample2', 'sample3', 'sample4'), c('A', 'T', 'G', 'C')))
  #insert random counts in two of the alleles
  for(allele in sample(c('A', 'T', 'G', 'C'), 2)){
    count[, allele] <- as.integer(rnorm(4, mean=50, sd=10))
  }
  countListPlus[[snp]] <- count
}

#make example alleleCountListMinus
countListMinus <- list()
snps <- c('snp1', 'snp2', 'snp3', 'snp4', 'snp5')
for(snp in snps){
  count <- matrix(rep(0, 16), ncol=4, dimnames=list(c('sample1', 'sample2', 'sample3', 'sample4'), c('A', 'T', 'G', 'C')))
  #insert random counts in two of the alleles
  for(allele in sample(c('A', 'T', 'G', 'C'), 2)){
    count[, allele] <- as.integer(rnorm(4, mean=50, sd=10))
  }
  countListMinus[[snp]] <- count
}

#make example rowRanges
rowRanges <- GRanges(
  seqnames = Rle(c('chr1', 'chr2', 'chr3', 'chr1'),
                 ranges = IRanges(1:5, width = 1, names = head(letters, 5)),
                 snp = paste('snp', 1:5, sep=''))
)

#make example colData
colData <- DataFrame(Treatment=c('ChIP', 'Input', 'Input', 'ChIP'),
                      row.names=c('ind1', 'ind2', 'ind3', 'ind4'))

#make ASEset
a <- ASEsetFromCountList(rowRanges, countListPlus=countListPlus,
                         countListMinus=countListMinus, colData=colData)
Description

Functions to construct DetectedAI objects

Usage

DetectedAIFromArray(
  x = "ASEset",
  strand = "x",
  reference.frequency = NULL,
  threshold.frequency = NULL,
  threshold.count.sample = NULL,
  threshold.delta.frequency = NULL,
  threshold.pvalue = NULL,
  threshold.frequency.names = NULL,
  threshold.count.sample.names = NULL,
  threshold.delta.frequency.names = NULL,
  threshold.pvalue.names = NULL,
  ...
)

Arguments

x ASEset
strand set strand to detectAI over "+","-","x"
reference.frequency frequencies of reference alleles based allele counts
threshold.frequency logical array for frequency thresholds
threshold.count.sample logical array for per sample allele count thresholds
threshold.delta.frequency logical array for delta frequency thresholds.
threshold.pvalue logical array for pvalue thresholds (max 1, min 0)
threshold.frequency.names character vector
threshold.count.sample.names character vector
threshold.delta.frequency.names character vector
threshold.pvalue.names character vector
...
internal arguments

Details

produces a class container for reference bias calculations
initialize-GlobalAnalysis

Author(s)
Jesper R. Gadin, Lasse Folkersen

Examples

```r
data(ASEset)
a <- ASEset
dai <- detectAI(a)
```

Description

Functions to construct GlobalAnalysis objects

Usage

```r
GAnalysis(x = "ASEset", ...)
```

Arguments

- `x`: ASEset
- `...`: internal arguments

Details

produces a class container for a global analysis

Author(s)
Jesper R. Gadin, Lasse Folkersen

Examples

```r
data(ASEset)
a <- ASEset
# gba <- gba(a)
```
initialize-RiskVariant

Initialize RiskVariant

Description

Functions to construct RiskVariant objects

Usage

RiskVariantFromGRangesAndPhaseArray(x, phase, ...)

Arguments

x GRanges object for the SNPs
phase array with phaseinfo
... internal arguments

Details

produces a class container for reference bias calculations

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

data(ASEset)
#p <- getPhaseFromSomewhere
#rv <- RiskVariantFromGRangesAndPhaseArray(x=GRvariants, phase=p)

legendBarplot

add legend to AllelicImbalance barplot

Description

adds a very customizable legend function for AllelicImbalance barplots.
Usage

legendBarplot(
  lowerLeftCorner,
  size,
  rownames,
  colnames,
  boxsize = 1,
  boxspace = 1,
  fgCol,
  bgCol,
  ylegendPos = 1,
  xlegendPos = 0.96,
  cex = 1
)

Arguments

lowerLeftCorner
  position of the plot to add legend to (default c(0,0))
size
  scale the plot, default is 1
rownames
  rownames in legend
colnames
  colnames in legend
boxsize
  size of each box fill
boxspace
  space inbetween the box fill
fgCol
  color for allele1
bgCol
  color for allele2
ylegendPos
  placement of the legend within the plot for y
xlegendPos
  placement of the legend within the plot for x
cex
  size of legend text

Details

the function is preferably called from within the AllelicImbalance barplot method.

Author(s)

Jesper R. Gadin

Examples

#code placeholders
#< create a barplot with legend >
#< add legend >
**LinkVariantAlmlof-class**

**LinkVariantAlmlof class**

**Description**

Object that holds results from AI detection.

**Usage**

\[
pvalue(x, \ldots)
\]

```r
## S4 method for signature 'LinkVariantAlmlof'
pvalue(x)
```

**Arguments**

- **x** LinkVariantAlmlof object
- **...** pass arguments to internal functions

**Details**

The LinkVariantAlmlof-class contains

**Author(s)**

Jesper R. Gadin, Lasse Folkersen

**Examples**

```r
#some code
```

---

**LinkVariantAlmlof-plot**

*plot LinkVariantAlmlof objects*

**Description**

plot an object of type LinkVariantAlmlof

**Usage**

\[
plot(x, y, \ldots)
\]

```r
## S4 method for signature 'LinkVariantAlmlof,ANY'
plot(x, y, \ldots)
```

---
Arguments

x LinkVariantAlmlof object
y not used
... pass on arguments to internal methods

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

data(ASEset)
a <- ASEset

# Add phase
set.seed(1)
p1 <- matrix(sample(c(1,0), replace=TRUE, size=nrow(a)*ncol(a)), nrow=nrow(a), ncol(a))
p2 <- matrix(sample(c(1,0), replace=TRUE, size=nrow(a)*ncol(a)), nrow=nrow(a), ncol(a))
p <- matrix(paste(p1, sample(c("|","","/"), size=nrow(a)*ncol(a), replace=TRUE), p2, sep=""), nrow=nrow(a), ncol(a))

phase(a) <- p

# Add alternative allele information
mcols(a)[["alt"]]<- inferAltAllele(a)

# init risk variants
p.ar <- phaseMatrix2Array(p)
rv <- RiskVariantFromGRangesAndPhaseArray(x=GRvariants, phase=p.ar)

# colnames has to be same and same order in ASEset and RiskVariant
colnames(a) <- colnames(rv)

# in this example each and every snp in the ASEset defines a region
r1 <- granges(a)

# in this example two overlapping subsets of snps in the ASEset defines the region
r2 <- split(granges(a)[c(1,2,2,3)], c(1,2,2))

# link variant almlof (lva)
lv1 <- lva(a, rv, r1)
lv2 <- lva(a, rv, r2)
plot(lv2[1])

---

lva

Description

make an almlof regression for arrays
Usage

lva(x, ...)

## S4 method for signature 'ASEset'
lva(
  x,
  rv,
  region,
  settings = list(),
  return.class = "LinkVariantAlmlof",
  type = "lm",
  verbose = FALSE,
  covariates = matrix(),
  ...
)

Arguments

x
ASESet object with phase and 'ref'/alt' allele information

... arguments to forward to internal functions

rv
RiskVariant object with phase and 'ref'/alt' allele information

region
RiskVariant object with phase and alternative allele information

settings
RiskVariant object with phase and alternative allele information

return.class
'LinkVariantAlmlof' (more options in future)

type
"lm" or "nlme", "nlme" needs subject information

verbose
logical, if set TRUE, then function will be more talkative

covariates
add data.frame with covariates (only integers and numeric)

Details

internal method that takes one array with results from regionSummary and one matrix with group information for each risk SNP (based on phase)

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

data(ASEset)
a <- ASEset
# Add phase
set.seed(1)
p1 <- matrix(sample(c(1,0),replace=TRUE, size=nrow(a)*ncol(a)),nrow=nrow(a), ncol(a))
p2 <- matrix(sample(c(1,0),replace=TRUE, size=nrow(a)*ncol(a)),nrow=nrow(a), ncol(a))
p <- matrix(paste(p1,sample(c("\"","\"","/"), size=nrow(a)*ncol(a), replace=TRUE), p2, sep=""), nrow=nrow(a), ncol(a))
phase(a) <- p

# add alternative allele information
mcols(a)["alt"] <- inferAltAllele(a)

# init risk variants
p.ar <- phaseMatrix2Array(p)
rv <- RiskVariantFromGRangesAndPhaseArray(x=GRvariants, phase=p.ar)

# colnames has to be same and same order in ASEset and RiskVariant
colnames(a) <- colnames(rv)

# in this example each and every snp in the ASEset defines a region
r1 <- granges(a)

# use GRangesList to merge and use regions defined by each element of the
# GRangesList
r1b <- GRangesList(r1)
r1c <- GRangesList(r1, r1)

# in this example two overlapping subsets of snps in the ASEset defines the region
r2 <- split(granges(a)[c(1,2,2,3),c(1,1,2,2)])

# link variant alnl0f (lva)
lva(a, rv, r1)
lva(a, rv, r1b)
lva(a, rv, r1c)
lva(a, rv, r2)

# Use covariates (integers or numeric)
cov <- data.frame(age=sample(20:70, ncol(a)), sex=rep(c(1,2), each=ncol(a)/2),
row.names=colnames(a))
lva(a, rv, r1, covariates=cov)
lva(a, rv, r1b, covariates=cov)
lva(a, rv, r1c, covariates=cov)
lva(a, rv, r2, covariates=cov)

# link variant alnl0f (lva), using nlme
a2 <- a
ac <- assays(a2)["countsPlus"]
jit <- sample(c(seq(-0.10,0,length=5), seq(0,0.10,length=5)), size=length(ac), replace=TRUE)
assays(a2, withDimnames=FALSE)["countsPlus"] <- round(ac * (1+jit),0)
ab <- cbind(a, a2)
colData(ab)[["subject.group"]]<- c(1:ncol(a),1:ncol(a))
rv2 <- rv[,c(1:ncol(a),1:ncol(a))]
colnames(ab) <- colnames(rv2)

lva(ab, rv2, r1, type="nlme")
lva(ab, rv2, r1b, type="nlme")
lva(ab, rv2, r1c, type="nlme")
lva(ab, rv2, r2, type="nlme")
**Description**

make an almlof regression for arrays (internal function)

**Usage**

```r
lva.internal(x, ...)
```

```r
## S4 method for signature 'array'
lva.internal(
x,
grp,
element = 3,
type = "lm",
subject = NULL,
covariates = matrix(),
...)
```

**Arguments**

- `x` regionSummary array phased for maternal allele
- `...` arguments to forward to internal functions
- `grp` group 1-3 (1 for 0:0, 2 for 1:0 or 0:1, and 3 for 1:1)
- `element` which column in x contains the values to use with lm.
- `type` which column in x contains the values to use with lm.
- `subject` which samples belongs to the same individual
- `covariates` add data.frame with covariates (only integers and numeric)

**Details**

internal method that takes one array with results from regionSummary and one matrix with group information for each risk SNP (based on phase). Input and output objects can change format slightly in future.

**Author(s)**

Jesper R. Gadin, Lasse Folkersen
Examples

```r
data(ASEset)
a <- ASEset
# Add phase
set.seed(1)
p1 <- matrix(sample(c(1,0), replace=TRUE, size=nrow(a)*ncol(a)), nrow=nrow(a), ncol(a))
p2 <- matrix(sample(c(1,0), replace=TRUE, size=nrow(a)*ncol(a)), nrow=nrow(a), ncol(a))
p <- matrix(paste(p1, sample(c("|","|","/"), size=nrow(a)*ncol(a), replace=TRUE), p2, sep=""), nrow=nrow(a), ncol(a))

phase(a) <- p

# add alternative allele information
mcols(a)[["alt"]]<- inferAltAllele(a)

# in this example two overlapping subsets of snps in the ASEset defines the region
region <- split(granges(a)[c(1,2,2,3)], c(1,1,2,2))
rs <- regionSummary(a, region, return.class="array", return.meta=FALSE)

# use (change to generated riskSNP phase later)
phs <- array(c(phase(a,return.class="array")[1,,c(1, 2)],
              phase(a,return.class="array")[2,,c(1, 2)]), dim=c(20,2,2))
grp <- matrix(2, nrow=dim(phs)[1], ncol=dim(phs)[2])
grp[(phs[,1] == 0) & (phs[,2] == 0)] <- 1
grp[(phs[,1] == 1) & (phs[,2] == 1)] <- 3
# only use mean.fr at the moment, which is col 3
lva.internal(x=assays(rs)[["rs1"]],grp=grp, element=3)
```
mapBiasRef

Arguments

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>fastaIn</td>
<td>character string of the path for the fasta file to be used</td>
</tr>
<tr>
<td>...</td>
<td>arguments to pass on</td>
</tr>
<tr>
<td>fastaOut</td>
<td>character string of the path for the masked fasta file (no extension)</td>
</tr>
<tr>
<td>posToReplace</td>
<td>GRanges object with the genomic ranges to replace</td>
</tr>
<tr>
<td>splitOnSeqlevels</td>
<td>write on file for each seqlevel to save memory</td>
</tr>
<tr>
<td>verbose</td>
<td>makes function more talkative</td>
</tr>
</tbody>
</table>

Author(s)

Jesper R. Gadin

Examples

```r
data(ASEset.sim)
gr <- rowRanges(ASEset.sim)
fastaIn <- system.file('extdata/hg19.chr17.subset.fa', package='AllelicImbalance')
makeMaskedFasta(fastaIn=fastaIn, fastaOut="fastaOut", posToReplace=gr)
```

**Description**

Create a matrix of bias for the reference allele

**Usage**

```r
mapBiasRef(x, ...)
```

## S4 method for signature 'ASEset'

```r
mapBiasRef(x)
```

**Arguments**

<table>
<thead>
<tr>
<th>Argument</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>x</td>
<td>ASEset object</td>
</tr>
<tr>
<td>...</td>
<td>internal arguments</td>
</tr>
</tbody>
</table>

**Details**

select the expected frequency for the reference allele
Author(s)
Jesper R. Gadin, Lasse Folkersen

Examples

```r
# load example data
data(ASEset)
a <- ASEset

mat <- mapBiasRef(a)
```

---

Description
filter on minCountFilt snps

Usage

```r
minCountFilt(x, ...)
```

## S4 method for signature 'ASEset'

```r
minCountFilt(
  x,
  strand = "*",
  threshold.counts = 1,
  sum = "all",
  replace.with = "zero",
  return.class = "ASEset"
)
```

Arguments

- `x` ASEset object
- `...` internal param
- `strand` strand to infer from
- `threshold.counts`
- `sum` 'each' or 'all'
- `replace.with` only option 'zero'
- `return.class` 'ASEset', 'array' or 'matrix'

Details
Description info here
Author(s)
Jesper R. Gadin, Lasse Folkersen

Examples
#load example data
data(ASEset)
a <- ASEset

minCountFilt(a)

Description
filter on minFreqFilt snps

Usage
minFreqFilt(x, ...)

## S4 method for signature 'ASEset'
minFreqFilt(
x,
    strand = "*",
    threshold.frequency = 0.1,
    replace.with = "zero",
    return.class = "ASEset",
    sum = "all"
)

Arguments
x  ASEset object
... internal param
strand  strand to infer from
threshold.frequency  least fraction to classify (see details)
replace.with  only option 'zero'
return.class  'ASEset', 'array' or 'matrix'
sum  'each' or 'all'

Details
Description info here
Author(s)
Jesper R. Gadin, Lasse Folkersen

Examples

# load example data
data(ASEset)
a <- ASEset

minFreqFilt(a)

multiAllelicFilt

multi-allelic filter methods

Description
filter on multiallelic snps

Usage

multiAllelicFilt(x, ...)

## S4 method for signature 'ASEset'

multiAllelicFilt(  
x,  
strand = "*",  
threshold.count.sample = 10,  
threshold.frequency = 0.1,  
filterOver = "eachSample"
)

Arguments

x ASEset object
... internal param
strand strand to infer from
threshold.count.sample least amount of counts to try to infer allele
threshold.frequency least fraction to classify (see details)
filterOver 'eachSample' or 'allSamples'

Details

based on the allele counts for all four variants A, T, G and C and returns true if there is counts enough suggesting a third or more alleles. The sensitivity can be specified using 'threshold.count.sample' and 'threshold.frequency'.
Author(s)
Jesper R. Gadin, Lasse Folkesen

Examples

```r
# load example data
data(ASEset)
a <- ASEset
multiAllelicFilt(a)
```

Description
Convert the phase from the internally stored phase, ref and alt information

Usage

```r
phase2genotype(x, ...)
```

```r
## S4 method for signature 'array'
phase2genotype(x, ref, alt, return.class = "matrix", ...)
```

Arguments

- `x` array see examples
- `...` pass on additional param
- `ref` reference allele vector
- `alt` alternative allele vector
- `return.class` 'matrix' or 'array'

Details
To not introduce redundant information in the ASEset object, the genotype matrix is accessed from the phase matrix, which together with ref and alt allele information contains the same information (not taken into account three-allelic or more SNPs).

The genotype matrix retrieved from an ASEset object can differ from the genotype matrix stored in the object if reference and alternative alleles were not used or has changed since the phase genotype matrix was stored. Basically, it is preferable to provide reference and alternative information when storing the genotype matrix.

If possible, it is better to not use a genotype matrix, but instead relying completely on storing a phase matrix (or array) together with reference and alternative allele information.
phaseArray2phaseMatrix

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

#load example data
data(ASEset)
data(genomatrix)
p <- genotype2phase(genomatrix, ref(ASEset), return.class="array")
ref <- ref(ASEset)
alt <- inferAltAllele(ASEset)

gt <- phase2genotype(p, ref, alt, return.class="matrix")

Description

used to convert the phase from the visually friendly matrix to array.

Usage

phaseArray2phaseMatrix(x, ...)

## S4 method for signature 'array'
phaseArray2phaseMatrix(x, ...)

Arguments

x array see examples

... arguments to forward to internal functions

Details

A more effective way to store the phase data in the ASEset object

Author(s)

Jesper R. Gadin, Lasse Folkersen
Examples

#load data
data(ASEset)
a <- ASEset
deference phase matrix
p1 <- matrix(sample(c(1,0),replace=TRUE, size=nrow(a)*ncol(a)),nrow=nrow(a), ncol(a))
p2 <- matrix(sample(c(1,0),replace=TRUE, size=nrow(a)*ncol(a)),nrow=nrow(a), ncol(a))
p <- matrix(paste(p1,sample(c("|","","/"), size=nrow(a)*ncol(a), replace=TRUE), p2, sep=""), nrow=nrow(a), ncol(a))
ar <- phaseMatrix2Array(p)
#Convert back
mat <- phaseArray2phaseMatrix(ar)

Description

used to convert the phase from the visually friendly matrix to array.

Usage

phaseMatrix2Array(x, ...)
    ## S4 method for signature 'matrix'
    phaseMatrix2Array(x, dimnames = NULL, ...)

Arguments

  x           matrix see examples
  ...         arguments to forward to internal functions
dimnames     list with dimnames

Details

A more effectice way of store the phase data in the ASEset object

Author(s)

   Jesper R. Gadin, Lasse Folkersen
randomRef

Examples

# load data
data(ASEset)
a <- ASEset

# example phase matrix
p1 <- matrix(sample(c(1,0),replace=TRUE, size=nrow(a)*ncol(a)),nrow=nrow(a), ncol(a))
p2 <- matrix(sample(c(1,0),replace=TRUE, size=nrow(a)*ncol(a)),nrow=nrow(a), ncol(a))
p <- matrix(paste(p1,sample(c("|","","/"), size=nrow(a)*ncol(a), replace=TRUE), p2, sep=""),
nrow=nrow(a), ncol(a))
ar <- phaseMatrix2Array(p)

randomRef

Random ref allele from genotype

Description

Create a vector of random reference alleles

Usage

randomRef(x, ...)

## S4 method for signature 'ASEset'
randomRef(x, source = "alleleCounts", ...)

Arguments

x         ASEset object
...
source    'alleleCounts'

Details

Randomly shuffles which of the two alleles for each genotype that is indicated as reference allele, based on either allele count information or previous ref and alt alleles.

When the source is 'alleleCounts', the two most expressed alleles are taken as reference and alternative allele.

Author(s)

Jesper R. Gadin, Lasse Folkersen
Examples

```r
# load example data
data(ASEset.sim)
a <- ASEset.sim

ref(a) <- randomRef(a, source = 'alleleCounts')
```

**Description**

This data set corresponds to the BAM-file data import illustrated in the vignette. The data set consists of a chromosome 17 region from 20 RNA-seq experiments of HapMap samples.

**Author(s)**

Jesper R. Gadin, Lasse Folkersen

**References**


**See Also**

- The `GRvariants` which is another example object

**Examples**

```r
# load example data (Not Run)
# data(reads)
```

---

**refAllele**

**Reference allele**

**Description**

Extract the allele based on SNP location from the reference fasta file

**Usage**

`refAllele(x, fasta)`
regionSummary

Arguments

- `x` ASEset object
- `fasta` path to fasta file, index should be located in the same folder

Details

The alleles will be placed in the rowRanges() meta column 'ref'

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

```r
#load example data
data(ASEset.sim)

fasta <- system.file('extdata/hg19.chr17.subset.fa', package='AllelicImbalance')
a <- refAllele(ASEset.sim, fasta=fasta)
```

Description

Gives a summary of AI-consistency for a transcript

Usage

```r
regionSummary(x, ...)
```

## S4 method for signature 'ASEset'

```r
regionSummary(x, region, strand = "*", return.class = "RegionSummary", ...)
```

Arguments

- `x` ASEset object
- `...` arguments to forward to internal functions
- `region` to summarize over, the object can be a GRanges, GRangesList
- `strand` can be "+", "-" or "*
- `return.class` "array" or "list".
RegionSummary-class

Details

From a given set of e.g. transcripts exon ranges the function will return a summary for the sum of all exons. Phase information, reference and alternative allele is required.

A limitation comes to the strand-specificness. At the moment it is not possible to call over more than one strand type using the strands in region. This will be improved before going to release.

to calculate the direction and binomial p-values of AI the mapbias stored in the ASEset is used. see ‘?mapBias’.

Author(s)

Jesper R. Gadin, Lasse Folkesen

Examples

data(ASEset)
a <- ASEset
# Add phase
set.seed(1)
p1 <- matrix(sample(c(1,0),replace=TRUE, size=nrow(a)*ncol(a)),nrow=nrow(a),ncol(a))
p2 <- matrix(sample(c(1,0),replace=TRUE, size=nrow(a)*ncol(a)),nrow=nrow(a),ncol(a))
p <- matrix(paste(p1,sample(c("|","|","/"), size=nrow(a)*ncol(a), replace=TRUE), p2, sep=""),
nrow=nrow(a), ncol(a))

phase(a) <- p

#add alternative allele information
mcols(a)[["alt"]] <- inferAltAllele(a)

# in this example each and all snps in the ASEset defines the region
region <- granges(a)
t <- regionSummary(a, region)

# in this example two overlapping subsets of snps in the ASEset defines the region
region <- split(granges(a)[c(1,2,2,3),c(1,1,2,2)])
t <- regionSummary(a, region)

RegionSummary-class RegionSummary class

Description

Object that holds results from the regionSummary method
Usage

sumnames(x, ...)

## S4 method for signature 'RegionSummary'
sumnames(x)

basic(x, ...)

## S4 method for signature 'RegionSummary'
basic(x)

Arguments

x RegionSummary object
...

pass arguments to internal functions

Details

The RegionSummary-class objects contains summaries for specified regions

Author(s)

Jesper R. Gadin, Lasse Folkersen

Examples

#some code

RiskVariant-class

RiskVariant class

Description

Object that holds results from AI detection.

Usage

## S4 method for signature 'RiskVariant'
ref(x)

## S4 replacement method for signature 'RiskVariant,ANY'
ref(x) <- value

## S4 method for signature 'RiskVariant'
alt(x)
## S4 replacement method for signature 'RiskVariant,ANY'
alt(x) <- value

## S4 method for signature 'RiskVariant'
phase(x, return.class = "matrix")

## S4 replacement method for signature 'RiskVariant'
phase(x) <- value

### Arguments

- **x**: RiskVariant object or list of RiskVariants
- **value**: argument used for replacement
- **return.class**: type of class returned eg. "list" or ""array".

### Details

The RiskVariant-class contains

### Author(s)

Jesper R. Gadin, Lasse Folkersen

### Examples

#some code

---

**scanForHeterozygotes.old**

scanForHeterozygotes-old

---

**Description**

Identifies the positions of SNPs found in BamGR reads.

**Usage**

scanForHeterozygotes.old(
    BamList,
    minimumReadsAtPos = 20,
    maximumMajorAlleleFrequency = 0.9,
    minimumBiAllelicFrequency = 0.9,
    maxReads = 15000,
    verbose = TRUE
)
Arguments

  BamList          A GAlignmentsList object
  minimumReadsAtPos  minimum number of reads required to call a SNP at a given position
  maximumMajorAlleleFrequency  maximum frequency allowed for the most common allele. Setting this parameter lower will minimise the SNP calls resulting from technical read errors, at the cost of missing loci with potential strong ASE
  minimumBiAllelicFrequency  minimum frequency allowed for the first and second most common allele. Setting a lower value for this parameter will minimise the identification of loci with three or more alleles in one sample. This is useful if sequencing errors are suspected to be common.
  maxReads        max number of reads of one list-element allowed
  verbose         logical indicating if process information should be displayed

Details

This function scans all reads stored in a GAlignmentsList for possible heterozygote positions. The user can balance the sensitivity of the search by modifying the minimumReadsAtPos, maximumMajorAlleleFrequency and minimumBiAllelicFrequency arguments.

Value

scanForHeterozygotes.old returns a GRanges object with the SNPs for the BamList object that was used as input.

Author(s)

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See Also

- The getAlleleCounts which is a function that count the number of reads overlapping a site.

Examples

data(reads)
s <- scanForHeterozygotes.old(reads, verbose=FALSE)
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