

# Package ‘genArise’

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**Title** Microarray Analysis tool

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**Depends** R (>= 1.7.1), locfit, tkrplot, methods

**Imports** graphics, grDevices, methods, stats, tcltk, utils, xtable

**Description** genArise is an easy to use tool for dual color microarray data. Its GUI-Tk based environment let any non-experienced user performs a basic, but not simple, data analysis just following a wizard. In addition it provides some tools for the developer.

**biocViews** Microarray, TwoChannel, Preprocessing

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## Contents

a.arise . . . . .	3
alter.unique . . . . .	4
analysis.window . . . . .	4
annotations . . . . .	5
back.gui . . . . .	5
bg.correct . . . . .	6
classes . . . . .	6
create.project . . . . .	7
cys.plot . . . . .	7
DataSet-class . . . . .	8
filter.spot . . . . .	8
genArise . . . . .	9
genArise.init . . . . .	9
genMerge . . . . .	10
get.values . . . . .	11
get.Zscore . . . . .	11
global.norm . . . . .	12
graphic.choose . . . . .	12
grid.norm . . . . .	13
help . . . . .	14
i.arise . . . . .	14
imageLimma . . . . .	15
m.arise . . . . .	16
ma.plot . . . . .	16
make.swap . . . . .	17
meanUnique . . . . .	18
note . . . . .	18
old.project . . . . .	19
post.analysis . . . . .	19
principal . . . . .	20
project.select . . . . .	20
r.arise . . . . .	21
read.dataset . . . . .	22
read.spot . . . . .	22
reset.history . . . . .	23
ri.plot . . . . .	24
set.grid.properties . . . . .	24
set.history.project . . . . .	25
set.path.project . . . . .	25
set.project.properties . . . . .	26
Simon . . . . .	26
single.norm . . . . .	27
Spot-class . . . . .	27
spotUnique . . . . .	28
swap.select . . . . .	28
trim . . . . .	29

<i>a.arise</i>	3
write.dataSet . . . . .	29
write.spot . . . . .	30
write.zscore . . . . .	31
WT.dataset . . . . .	32
Zscore . . . . .	32
Zscore.plot . . . . .	33
Zscore.points . . . . .	34
<b>Index</b>	<b>35</b>

---

a.arise	<i>A Arise</i>
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---

### Description

Extract **A values** from a Spot.

### Usage

```
a.arise(mySpot)
```

### Arguments

mySpot            Spot object for one microarray.

### Value

List of A-values.  $(\log(\text{cy3}, 2) + \log(\text{cy5}, 2))/2$

### See Also

[m.arise.](#)

### Examples

```
## read the spot from a file and save it in spot
data(Simon)
## Extract A from spot and save in a
a <- a.arise(mySpot = Simon)
```

---

alter.unique	<i>Remove Duplicates</i>
--------------	--------------------------

---

**Description**

This function allows to remove from the spot repeated Id's. Before moving one of the repeated Id's the function compute the log ratio of both values with the same Id and delete the least absolute value if both of them are positive or negative. In other case delete both observations.

**Usage**

```
alter.unique(mySpot)
```

**Arguments**

mySpot            Spot object for one microarray.

**Value**

Spot object without duplicates.

**Examples**

```
data(Simon)
## filter the spot and save it in f.spot
f.spot <- filter.spot(Simon)
## remove duplicates and save it in u.spot
u.spot <- alter.unique(f.spot)
```

---

analysis.window	<i>Analysis.window</i>
-----------------	------------------------

---

**Description**

Auxiliar function of genArise GUI, in this window you can apply operations to original data.

**Usage**

```
analysis.window(texto, follow.wizard = FALSE, envir, swap)
```

**Arguments**

texto            Historial project string  
follow.wizard    Boolean value, if this argument is TRUE, an data analysis are performed  
envir            Environment where are the project variables  
swap             Is this a swap analysis or an individual analysis

**Value**

tkwidget

annotations

*Gene Annotations*

**Description**

Performed an HTML file

**Usage**

```
annotations(specie.data, specie, column, symbol,
output.file = "annotations.html")
```

**Arguments**

specie.data	A data frame
specie	Name of specie
column	Number of column where are the gene name in the data frame
symbol	An optional symbol besides GenBank ID
output.file	Name of output file

**Value**

HTML file with link for each spot in data frame

back.gui

*Return to the last window*

**Description**

Auxiliar function of genArise GUI.

**Usage**

```
back.gui(envir)
```

**Arguments**

envir	Environment where are the project variables
-------	---

**Value**

tkwidget

---

bg.correct	<i>Background Correction</i>
------------	------------------------------

---

### Description

This function use the background data to eliminate unwanted effects in signal. The background correction establish the new Cy3 signal as  $Cy3 - BgCy3$  and the new Cy5 as  $Cy5 - BgCy5$ .

### Usage

```
bg.correct(mySpot)
```

### Arguments

mySpot            Spot object for one microarray.

### Value

Spot object with the background correction done.

### Examples

```
data(Simon)
## background correction and save it in c.spot
c.spot <- bg.correct(Simon)
```

---

classes	<i>Classes Defined by this Package</i>
---------	--

---

### Description

This package defines the following data classes.

**Spot** A class used to store spot data with the following attributes: Cy3, Cy5, BgCy3, BgCy5, Ids as they are read by `read.spot` or obtained from a function that return a spot object.

**DataSet** A class used to store spot data with the following attributes: Cy3, Cy5, Ids, Z-score.

---

create.project	<i>Create directorie for the project and its results and graphics</i>
----------------	---

---

**Description**

Auxiliar function for genAriseGUI. Create the directory's hierarchy of the project.

**Usage**

```
create.project(project.name, results.file = "Results",  
              graphics.file = "Graphics")
```

**Arguments**

project.name	Project directory name.
results.file	Filename of the project result.
graphics.file	Filename of the project graphics.

---

cys.plot	<i>Data Visualization: log<sub>2</sub>(Cy3) vs log<sub>2</sub>(Cy5)</i>
----------	---

---

**Description**

This function shows the plot of the values from the log Cy3 against log Cy5 intensities that belongs to an object of the Spot class.

**Usage**

```
cys.plot(mySpot, col = "green")
```

**Arguments**

mySpot	An Spot object
col	Color in which the points of the plot will be shown. This argument must be quoted and the possible values it can take are the same from the color funcion in the R base.

**Examples**

```
data(Simon)  
cys.plot(Simon)
```

---

DataSet-class	<i>DataSet - class</i>
---------------	------------------------

---

### Description

A simple list-based class for storing red and green channel foreground, z-scores and the Ids.

### Creating Objects from the Class

Objects can be created by calls of the form `new("DataSet", sets, type)` where `sets` is a list containing `Cy3`, `Cy5`, `Id` and `Zscore` and `type` is "ri" or "ma". Objects are normally created by [read.spot](#).

### Slots/List Components

This class contains no slots (other than `.Data`), but objects should contain the following list components:

- `Cy5`: numeric matrix containing the red (`cy5`) foreground intensities. Rows correspond to spots and columns to arrays.
- `Cy3`: numeric matrix containing the green (`cy3`) foreground intensities.
- `Id`: Ids from all the observations.
- `Zscore`: The result of  $(R - \text{mean}) / \text{sd}$  that define an intensity-dependent Z-score threshold to identify differential expression

All of these matrices should have the same dimensions.

### Methods

This class inherits directly from class `list` so any operation appropriate for lists will work on objects of this class.

---

<code>filter.spot</code>	<i>Intensity-based filtering of array elements</i>
--------------------------	--

---

### Description

This function keep only array elements with intensities that are 2 standard deviation above background.

### Usage

```
filter.spot(mySpot)
```

### Arguments

`mySpot` Spot object for one microarray.



**Value**

Array elements with intensities that are 2 standard deviation above background.

**References**

John Quackenbush "Microarray data normalization and transformation". Nature Genetics. Vol.32 supplement pp496-501 (2002)

**Examples**

```
data(Simon)
## background correction and save it in c.spot
c.spot <- bg.correct(Simon)
## normalize spot
n.spot <- grid.norm(c.spot, nr = 23, nc = 24)
## filtering the spot
filter.spot(n.spot)
```

---

genArise

*GUI: Graphical User Interface*

---

**Description**

This is the main function and display the GUI of genArise.

**Usage**

```
genArise()
```

---

genArise.init

*genArise.init*

---

**Description**

Auxiliar function of genArise GUI, this function show a principal menu of genAriseGUI

**Usage**

```
genArise.init(envir)
```

**Arguments**

envir            Environment where are the project variables

**Value**

tkwidget

genMerge

*genMerge: Post-Genomic Analysis*

---

**Description**

After we finished our slice analysis we get a up-regulated and down-regulated set. This will be the set of study genes for genMege. Given this set, genMerge retrieves functional genomic data for each gene and provides statistical rank scores for over-representation of particular functions in the dataset.

**Usage**

```
genMerge(gene.association, description, population.genes,  
study.genes, output.file = "GenMerge.txt")
```

**Arguments**

gene.association	The gene-association file links gene names with a particular datum of information using a shorthand of gene-association IDS
description	The description file contains human-readable description of gene-association IDS
population.genes	Set of all genes detected on a array
study.genes	Set of genes may be those that are up-regulated or down-regulated or both of them.
output.file	The name of output file that includes all results obtained after this analisys.

**Note**

This function is completely based on GeneMerge from Cristian I. Castillo-Davis and Daniel L. Hartl

**References**

Cristian I. Castillo-Davis Department of Statistics Harvard University <http://www.oeb.harvard.edu/hartl/lab/publications/GeneMerge>

---

get.values	<i>Auxiliar function for post-analysis</i>
------------	--

---

**Description**

This function get values from an DataSet object.

This is just a function for the GUI, and can not be used in the command line.

**Usage**

```
get.values(list.values, genes.values, up.down, min.val, max.val)
```

**Arguments**

list.values	Zscore values from DataSet object
genes.values	Ids values from DataSet object
up.down	If the analysis will be done with "up" or "down" regulated
min.val	Minimal value of the range
max.val	Maximal value of the range

**Value**

An Ids list

---

get.Zscore	<i>Swap from Files</i>
------------	------------------------

---

**Description**

Read both files, but only extract the interested columns and create a Spot object.

**Usage**

```
get.Zscore( spot, name, Zscore.min=NULL, Zscore.max=NULL, all=FALSE, envir)
```

**Arguments**

spot	a connection or a character string giving the name of the file to read where each column represent the spot components.
name	a connection or a character string giving the name of the file to read where each column represent the spot components.
Zscore.min	column that represent Cy3.
Zscore.max	column that represent Cy5.
all	column that represent BgCy3.
envir	Environment where are the genArise variables.

**See Also**

[write.spot.](#)

---

global.norm

*Global Normalization of Spot*

---

**Description**

This function normalize R and I values and fit the value of Cy5 from his argument. In this function the normalize algorithm will be applied to all observations to get the lowess factor and then fit Cy5 with this factor. The observations with values R = 0 are deleted because they have no change in their expression levels.

**Usage**

```
global.norm(mySpot)
```

**Arguments**

mySpot            A spot object

**Value**

A new spot object but normalized, It means with a different Cy5 that is the result of the fit with the lowess factor.

**Examples**

```
data(Simon)
# Background Correction
c.spot <- bg.correct(Simon)
#Normalized data
n.spot <- global.norm(c.spot)
```

---

graphic.choose

*Graphic choose*

---

**Description**

This function show the plot of an spot subject. This plot are identify with the graphic.type.value

**Usage**

```
graphic.choose(spot.object, graphic.type)
```

**Arguments**

spot.object     An object ob Spot class  
graphic.type    representative integer of type graphic will be plot

**Value**

Plot device

---

grid.norm                    *Normalization by grid of Spot*

---

**Description**

This function normalize R and I values and fit the value of Cy5 for each grid in the spot that it receives as argument. In this function the dimension of grid is (meta-row \* meta-column).

**Usage**

```
grid.norm(mySpot, nr, nc)
```

**Arguments**

mySpot            Spot object for one microarray.  
nr                Total of meta-row.  
nc                Total of meta-column.

**Value**

Spot object with the grid normalization done.

**Examples**

```
data(Simon)  
## background correction and save it in c.spot  
c.spot <- bg.correct(Simon)  
## normalization and save it in n.spot  
n.spot <- grid.norm(c.spot, 23, 24)
```

---

help	<i>Help of genArise</i>
------	-------------------------

---

**Description**

Display the help of genArise in the GUI. This is just a function for the GUI, and can not be used in the command line.

**Usage**

```
help()
```

---

i.arise	<i>I Arise</i>
---------	----------------

---

**Description**

Extract **I** from a Spot.

**Usage**

```
i.arise(mySpot)
```

**Arguments**

mySpot            Spot object for one microarray.

**Value**

List of I-values

**See Also**

[r.arise.](#)

**Examples**

```
data(Simon)
## Extract I from spot and save in i
i.arise(Simon)
```

---

imageLimma	<i>Image Plot of Microarray</i>
------------	---------------------------------

---

**Description**

Plot an image of colours representing the log intensity ratio for each spot on the array. This function can be used to explore whether there are any spatial effects in the data.

**Usage**

```
imageLimma(z, row, column, meta.row, meta.column,
low = NULL, high = NULL)
```

**Arguments**

z	numeric vector or array. This vector can contain any spot statistics, such as log intensity ratios, spot sizes or shapes, or t-statistics. Missing values are allowed and will result in blank spots on the image
row	rows in the microarray
column	columns in the microarray
meta.row	metarows in the microarray
meta.column	metacolumns in the microarray
low	color associated with low values of 'z'. May be specified as a character string such as "green", "white" etc, or as a rgb vector in which 'c(1,0,0)' is red, 'c(0,1,0)' is green and 'c(0,0,1)' is blue. The default value is "green" if 'zerocenter=T' or "white" if 'zerocenter=F'.
high	color associated with high values of 'z'. The default value is "red" if 'zerocenter=T' or "blue" if 'zerocenter=F'.

**Note**

This function is based in the imageplot function from limma package.

**References**

Gordon K. Smyth (2004) "Linear Models and Empirical Bayes Methods for Assessing Differential Expression in Microarray Experiments", *Statistical Applications in Genetics and Molecular Biology*: Vol. 3: No. 1, Article 3. <http://www.bepress.com/sagmb/vol3/iss1/art3>

**Examples**

```
data(Simon)
spot.data <- attr(Simon, "spotData")
M <- log(spot.data$Cy5, 2) - log(spot.data$Cy3, 2)
imageLimma(z = M, row = 23, column = 24, meta.row = 2, meta.column = 2,
low = NULL, high = NULL)
```

m.arise

*M Arise*

---

**Description**

Extract **M values** from a Spot.

**Usage**

```
m.arise(mySpot)
```

**Arguments**

mySpot            Spot object for one microarray.

**Value**

List of M-values

**See Also**

[a.arise.](#)

**Examples**

```
data(Simon)
## Extract M from spot and save in m
m <- m.arise(Simon)
```

---

ma.plot*Data Visualization: M vs. A plot*

---

**Description**

This function allows to plot **M -vs- A** in spot.

**Usage**

```
ma.plot(mySpot, col = "green")
```

**Arguments**

mySpot            Spot for one microarray.  
col                color of points in graphic



**Examples**

```
data(Simon)
## plot the signals for spot.
ma.plot(Simon)
```

---

make.swap

*Swap analysis*


---

**Description**

Read both files, but only extract the interested columns and create a Spot object.

**Usage**

```
make.swap(spot1, spot2, Cy3, Cy5, BgCy3, BgCy5, Id, Symdesc, header = FALSE, is.ifc = FALSE, envir, nr, nc)
```

**Arguments**

spot1	a connection or a character string giving the name of the file to read where each column represent the spot components.
spot2	a connection or a character string giving the name of the file to read where each column represent the spot components.
Cy3	column that represent Cy3.
Cy5	column that represent Cy5.
BgCy3	column that represent BgCy3.
BgCy5	column that represent BgCy5.
Id	column that represent Id.
Symdesc	optional identifier besides the Id column.
header	the logical value of the header input file
is.ifc	If is.ifc = TRUE this experiment was done in the Unit of Microarray from Cellular Physiology Institute.
envir	Environment where are the genArise variables.
nr	Total of meta-row.
nc	Total of meta-column.

**See Also**

[write.spot.](#)

---

`meanUnique`*Remove Duplicates*

---

**Description**

This function allows to remove from the spot repeated Id's. Before moving one of the repeated Id's the function compute the average of Cy3 intensity and Cy5 intensity.

**Usage**

```
meanUnique(mySpot)
```

**Arguments**

`mySpot` Spot object for one microarray.

**Value**

Spot object without duplicates

**Examples**

```
data(Simon)
c.spot <- bg.correct(Simon)
n.spot <- global.norm(c.spot)
f.spot <- filter.spot(n.spot)
meanUnique(f.spot)
```

---

`note`*note*

---

**Description**

Call a editor for note about actual experiment

**Usage**

```
note(envir)
```

**Arguments**

`envir` Environment where are the experiment variables

---

old.project	<i>Open previous project</i>
-------------	------------------------------

---

**Description**

Show the information that was obtained from the analysis of a previous project. This is just an auxiliary function for genAriseGUI, and can not be used in the command line.

**Usage**

```
old.project(project.name,envir, parent)
```

**Arguments**

project.name	path of project file (PRJ)
envir	Environment where are the genArise variables
parent	The parent widget

**Value**

tkwidget

---

post.analysis	<i>Set-combinatorial analysis</i>
---------------	-----------------------------------

---

**Description**

This function allows you to perform a set combinatorial analysis between the results previously obtained in different projects. This function is called post.analysis and it is mandatory that you have done the Zscore operation in all the selected projects. It is important to clarify that this function receives a list of files with extension prj as argument and for this reason you can't use it if the results to compare was not obtained by the genArise GUI.

**Usage**

```
post.analysis(values, min.val, max.val, up.down, output)
```

**Arguments**

values	A list of projects to compare
min.val	The minimal value of the range
max.val	The maximal value of the range
up.down	If the analysis will be done with "up" or "down" regulated
output	The directory that will contain all the output files

**Value**

Once obtained the ids list for each project a number of files with extension set are created in a directory. The name of this files consists in a sequence of 0 and 1. The number of digits in the file names is the same to the number of projects in the list passed as argument to the function. There is then, a relation between the number of digits in the file names and the projects. This relation is defined by the position specified in the file order.txt in the same directory you have passed as another argument in the function.

---

principal	<i>Principal window of genAriseGUI</i>
-----------	--

---

**Description**

This function show a window with the information of experiment like name and dimensions, too plot an image of colours representing the log intensity ratio for each spot on the array. This is just an auxiliar function for genAriseGUI, and can not beused in the command line.

**Usage**

```
principal(envir, swap)
```

**Arguments**

envir	Environment where are the genArise variables
swap	Is this a swap analysis or an individual analysis

**Value**

tkwidget

---

project.select	<i>File selector</i>
----------------	----------------------

---

**Description**

Previous window to post-analysis. In this window you can select one or several files (projects) and arguments to be used by post analysis function.

This is just an auxiliar function for genAriseGUI, and can not be used in the command line.

**Usage**

```
projects.select(envir, nombre)
```

**Arguments**

envir            Environment where are the genArise variables  
nombre          Name of directory where the post-analysis results will be placed.

**Value**

tkwidget

---

r.arise                      *Get R value*

---

**Description**

Get the **R values** from an object of the Spot class.

**Usage**

```
r.arise(mySpot)
```

**Arguments**

mySpot            An object of the Spot class

**Value**

A vector containing the R value ( $\log(\text{Cy5}/\text{Cy3})$ ) for each observation of the spot object.

**See Also**

[i.arise.](#)

**Examples**

```
data(Simon)
#Get R-value from an object of the Spot class and save the result
R <- r.arise(Simon)
#Show the R-values
```

---

read.dataset	<i>Read Dataset from File</i>
--------------	-------------------------------

---

### Description

Read all file and extract the interested columns to create a DataSet object (this file contain the zscore with all the genes after the duplicates filtering and makes not distinction between up-regulated and down-regulated. If you want to make this distinction you must write the data with the function write.dataSet, but there is no way to read this files with this function).

### Usage

```
read.dataset(file.name, cy3 = 1, cy5 = 2, ids = 3, symdesc = NULL,
zscore = 4, type = 6, header = FALSE, sep = "\t")
```

### Arguments

file.name	a connection or a character string giving the name of the file to read where each column represent the dataset components.
cy3	column that represent Cy3.
cy5	column that represent Cy5.
ids	column that represent Id.
symdesc	optional identifier besides Id column.
zscore	column that represent the zscore value.
type	column that represent if the experiment was performed as R vs I or M vs A.
header	the logical value of the header input file
sep	the separator in the inputfile

### See Also

[write.zscore.](#)

---

read.spot	<i>Read Spot from File</i>
-----------	----------------------------

---

### Description

Read all file, but only extract the interested columns and create a Spot object.

### Usage

```
read.spot(file.name, cy3, cy5, bg.cy3, bg.cy5, ids, symdesc, header =
FALSE, sep = "\t", is.ifc = FALSE, envir)
```

**Arguments**

file.name	a connection or a character string giving the name of the file to read where each column represent the spot components.
cy3	column that represent Cy3.
cy5	column that represent Cy5.
bg.cy3	column that represent BgCy3.
bg.cy5	column that represent BgCy5.
ids	column that represent Id.
symdesc	(optional) identifier besides Id column.
header	the logical value of the header input file
sep	the separator in the inputfile
is.ifc	If is.ifc = TRUE this experiment was done in the Unit of Microarray from Cellular Physiology Institute.
envir	Environment where are the genArise variables. You don't need to specify this argument.

**See Also**

[write.spot.](#)

---

reset.history	<i>Reset the prj history file</i>
---------------	-----------------------------------

---

**Description**

Clean all the operations saved in the prj history file.

**Usage**

```
reset.history(history.file, text)
```

**Arguments**

history.file	The name of the prj history file.
text	The new content of the prj history file.

**Value**

The history file without operations.

---

ri.plot *Data Visualization: R vs I*

---

### Description

This function allows to plot **R-values** vs **I-values I-value** from a Spot object

### Usage

```
ri.plot(mySpot, col = "green")
```

### Arguments

mySpot	Spot Object
col	Color in which the points of the plot will be shown. This argument must be quoted and the possible values it can take are the same from the colors function in the R base package.

### See Also

colors()

### Examples

```
data(Simon)
ri.plot(Simon)
```

---

set.grid.properties *set.grid.properties*

---

### Description

Auxiliary function for genAriseGUI

### Usage

```
set.grid.properties(envir, name, nr, nc, nmr, nmc)
```

### Arguments

envir	Environment where the variables are stored
name	The name of the experiment
nr	Total rows in the array (each row represent a spot)
nc	Total columns in the array
nmr	Total of meta-rows
nmc	Total of meta-columns



---

set.history.project     *Save the history of a project*

---

**Description**

Save in the history file each operation performed while the analysis. This is just to get the open this particular project in the future. This is just an auxiliary function for the GUI, and can not be used in the command line.

**Usage**

```
set.history.project(history.file, id.name, data.file)
```

**Arguments**

history.file	The name of the prj history file.
id.name	The name of the operation.
data.file	The file with the results of the operation.

**Value**

The history file with the new performed operation.

---

set.path.project     *set.path.project*

---

**Description**

Auxiliar function for genAriseGUI

**Usage**

```
set.path.project(path, results.file, graphics.file, envir)
```

**Arguments**

path	Project path value
results.file	Name of directory where results file will be
graphics.file	Name of directory where pdf graphics will be
envir	Environment where are the experiment variables

```
set.project.properties  
      set.project.properties
```

---

**Description**

Auxiliar function for genAriseGUI

**Usage**

```
set.project.properties(envir)
```

**Arguments**

envir                    Environment where are the experiment variables

---

Simon                    *Dataset: Little fragment of a microarray from IFC UNAM*

---

**Description**

This structure is a data fragment of a yeast microarray from the Microarrays Unit in IFC UNAM. The original microarray contains 6 meta-rows and 4 meta-columns, however this data just belongs to the first meta-row order in a way of 2 meta-rows and 2 meta-columns.

**Usage**

```
data(Simon)
```

**Format**

A list that contains 1104 observations, because the dimensions of this example are: 2 meta-rows, 2 meta-columns, 23 rows, 24 columns.

**Examples**

```
data(Simon)  
#A preview from the chip  
datos <- attr(Simon, "spotData")  
M <- log(datos$Cy3, 2) - log(datos$Cy5, 2)  
imageLimma(M, 23, 24, 2, 2)
```

---

 single.norm

*Swap from Files*


---

**Description**

Read both files, but only extract the interested columns and create a Spot object.

**Usage**

```
single.norm(envir)
```

**Arguments**

envir            Environment where are the genArise variables.

**See Also**

[write.spot.](#)

---

 Spot-class

*Spot - class*


---

**Description**

A simple list-based class for storing red and green channel foreground and background intensities for a batch of spotted microarrays and the Ids.

**Creating Objects from the Class**

Objects can be created by calls of the form `new("Spot", spot)` where `spot` is a list. Objects are normally created by [read.spot](#).

**Slots/List Components**

This class contains no slots (other than `.Data`), but objects should contain the following list components:

Cy5:        numeric matrix containing the red (cy5) foreground intensities. Rows correspond to spots and columns to arrays.  
 Cy3:        numeric matrix containing the green (cy3) foreground intensities.  
 BgCy5:     numeric matrix containing the red (cy5) background intensities.  
 BgCy3:     numeric matrix containing the green background intensities.  
 Id:        Ids from all the observations.

All of these matrices should have the same dimensions.

**Methods**

This class inherits directly from class `list` so any operation appropriate for lists will work on objects of this class.

---

spotUnique	<i>Replicate filtering</i>
------------	----------------------------

---

**Description**

We consider replicate measures of two samples and adjust the  $\log(\text{ratio}, 2)$  measures for each gene so that the transformed values are equal. To do this we take the geometric mean. This procedure can be extended to averaging over  $n$  replicates.

**Usage**

```
spotUnique(mySpot)
```

**Arguments**

mySpot            Spot object for one microarray.

**Value**

Spot object without duplicates

**Examples**

```
data(Simon)
c.spot <- bg.correct(Simon)
f.spot <- filter.spot(c.spot)
spotUnique(mySpot = f.spot)
```

---

swap.select	<i>Dye swap files selector</i>
-------------	--------------------------------

---

**Description**

This is just an auxiliary function for genAriseGUI, and can not be used in the command line.

**Usage**

```
swap.select(envir)
```

**Arguments**

envir            Environment where are the genArise variables

**Value**

tkwidget

---

`trim`

---

*Trim***Description**

Extract white spaces at the beginning or end of a word.

**Usage**`trim(word)`**Arguments**

`word` A string of characters possibly with white spaces at the beginning or end of the string.

**Value**

Returns a string of characters, with leading and trailing whitespace omitted.

**Examples**

```
trim("      This is a String      ")
## return [1] "This is a String"
```

---

`write.dataSet`

---

*Write dataSet***Description**

Write the values for observations of an object of `DataSet` class in an output file. These values are written in columns with the following order: `Cy3`, `Cy5`, `Cy3 Background`, `Cy5 Background`, `Ids` and finally the `Zscore` value. By default this output file has no header.

**Usage**

```
write.dataSet(dataSet.spot, fileName, quote
= FALSE, col.names = FALSE, row.names = FALSE,
Zscore.min = NULL, Zscore.max = NULL, sep = "\t")
```

**Arguments**

dataSet.spot	An object of DataSet class
fileName	The name of the output file where the data will be written. This argument must be quoted.
quote	If quote = TRUE, all values in the file will be quoted.
col.names	If col.names = TRUE, an integer is written in every column as header. By default col.names = FALSE.
row.names	If row.names = TRUE will be an extra column that numerates every rows in the file.
Zscore.min	The lower value in a range, if Zscore.min = NULL then the file will contain all values bellow Zscore.max
Zscore.max	The greater value in a range, if Zscore.max = NULL then file will be contain all values above Zscore.min. Both values, Zscore.min and Zscore.max can not be NULL
sep	Character to separate the columns in file. By default sep = "\t".

**Examples**

```
data(WT.dataset)
write.dataSet(dataSet.spot = WT.dataset, fileName = "Example.csv", Zscore.min = 1,
Zscore.max = 1.5, sep = "\t")
```

---

write.spot

*Write Spot*

---

**Description**

Write the values for observations of an object of Spot class in an output file. This values are written in columns with the follow order: Cy3, Cy5, Cy3 Background, Cy5 Background and finally Ids. By default this file has no header.

**Usage**

```
write.spot(spot, fileName, quote = FALSE, sep = "\t",
col.names = FALSE, row.names = FALSE)
```

**Arguments**

spot	An object of Spot class
fileName	The name of the output file where the data will be written. This argument must be quoted.
quote	If quote = TRUE, all values in the file will be quoted.
sep	Character to separate the columns in file. By default sep = "\t".

- `col.names`      If `col.names = TRUE`, an integer is written in every column as header. By default `col.names = FALSE`.
- `row.names`      If `row.names = TRUE` will be an extra column that numerates every rows in the file.
- `read.spot.`

## Examples

```
data(Simon)
write.spot(spot = Simon, fileName = "Example.csv", quote = FALSE, sep =
"\t", col.names = FALSE, row.names = FALSE)
```

---

<code>write.zscore</code>	<i>Write Z-score data</i>
---------------------------	---------------------------

---

## Description

Write the values for observations of an object of `DataSet` class in an output file. This values are written in columns tab separated with the follow order: Cy3, Cy5, Cy3 Background, Cy5 Background, Ids and finally the z-score value. The header of the output file is the selected type for the z-score (ri or ma).

## Usage

```
write.zscore(dataSet.spot, fileName, sep = "\t")
```

## Arguments

- `dataSet.spot`    An object of `DataSet` class
- `fileName`        The name of the output file where the data will be written. This argument must be quoted.
- `sep`              Character to separate the columns in file. By default `sep = "\t"`.

## Examples

```
data(WT.dataset)
write.zscore(dataSet.spot = WT.dataset, fileName = "Zscore.csv", sep =
"\t")
```

WT.dataset

*Microarray from the IFC*

---

**Description**

This data set is a Microarray from the IFC.

**Usage**

```
data(WT.dataset)
```

**Format**

A vector containing 4036 observations.

**Examples**

```
data(WT.dataset)
Zscore.plot(WT.dataset)
```

---

Zscore

*Z-scores for identifying differential expression*

---

**Description**

This function identify differential expressed genes by calculating an intensity-dependent Z-score. This function use a sliding window to calculate the mean and standard deviation within a window surrounding each data point, and define a Z-score where Z measures the number of standard deviations a data point is from the mean.

**Usage**

```
Zscore(spot.object,type>window.size)
```

**Arguments**

spot.object	A spot object
type	Type of analysis: "ri" is for a R-I analysis and "ma" is for M-A analysis
window.size	Size of the sliding window

**Value**

A dataSet object with attributes Cy3, Cy5, Id, Z-score.



**Examples**

```

data(Simon)
# Background Correction
c.spot <- bg.correct(Simon)
#Normalized data
n.spot <- grid.norm(c.spot,23,24)
#Filter spot
f.spot <- filter.spot(n.spot)
#Replicate filtering
u.spot <- spotUnique(f.spot)
#Zscore analysis
s.spot <- Zscore(u.spot)

```

---

Zscore.plot

*Z-score Data Visualization: R vs I or M vs A*


---

**Description**

This function allows to plot **R-values** vs **I-values** or **M-values** vs **A-values** for identifying differential expression.

**Usage**

```
Zscore.plot(dataSet.spot, Zscore.min, Zscore.max, all, col)
```

**Arguments**

dataSet.spot	Spot Object
Zscore.min	The lower value in a range, if Zscore.min = NULL then the file will contain all values bellow Zscore.max
Zscore.max	The greater value in a range, if Zscore.max = NULL then file will be contain all values above Zscore.min. Both values, Zscore.min and Zscore.max can not be NULL
all	Plot all the observations in four sets: $Z < 1$ , $1 < Z < 1.5$ , $1.5 < Z < 2$ , $Z > 2$
col	Color in which the pioints of the plot will be shown where only the points from center are plot. This argument must be quoted and the possible values it can take are the same from the colors function in the R base package.

**See Also**

colors()

**Examples**

```

data(WT.dataset)
Zscore.plot(WT.dataset, Zscore.min = 1, Zscore.max = 2)

```

---

`Zscore.points`*Z-score Window*

---

**Description**

This function display the window that show the results after the Z-score. This window allow:

1. Show the plots of the up and down generated with the function `Zscore.plot` regulated spots in: `Zscore < 1 sd` `1 sd < Zscore < 1.5 sd` `1.5 sd < Zscore < 2 sd` `Zscore > 2 sd` and All the points
2. Save the plots in pdf and save the results in an output file
3. Gene annotations. Denote any gene information beyond the expression level data.

This is just a function for the GUI, and can not be used in the command line.

**Usage**

```
Zscore.points(type,text,envir, swap)
```

**Arguments**

<code>type</code>	Type of analysis done: "ri" is for a R-I analysis and "ma" is for M-A analysis
<code>text</code>	The text for the text area of the history of the project
<code>envir</code>	Environment where the variables are stored
<code>swap</code>	Is this a swap analysis or an individual analysis

# Index

## \* IO

- back.gui, 5
- set.grid.properties, 24

## \* aplot

- cys.plot, 7
- ma.plot, 16
- ri.plot, 24
- Zscore.plot, 33
- Zscore.points, 34

## \* arith

- a.arise, 3
- alter.unique, 4
- create.project, 7
- filter.spot, 8
- genArise, 9
- get.values, 11
- global.norm, 12
- grid.norm, 13
- m.arise, 16
- ma.plot, 16
- note, 18
- r.arise, 21
- set.path.project, 25
- set.project.properties, 26
- Zscore, 32

## \* character

- trim, 29

## \* classes

- DataSet-class, 8
- Spot-class, 27

## \* color

- imageLimma, 15
- principal, 20

## \* datasets

- Simon, 26
- WT.dataset, 32

## \* data

- DataSet-class, 8
- Spot-class, 27

## \* documentation

- classes, 6
- help, 14

## \* file

- get.Zscore, 11
- make.swap, 17
- old.project, 19
- post.analysis, 19
- project.select, 20
- read.dataset, 22
- read.spot, 22
- reset.history, 23
- set.history.project, 25
- single.norm, 27
- swap.select, 28
- write.dataSet, 29
- write.spot, 30
- write.zscore, 31

## \* graphs

- cys.plot, 7

## \* math

- a.arise, 3
- alter.unique, 4
- analysis.window, 4
- annotations, 5
- bg.correct, 6
- filter.spot, 8
- genArise, 9
- genArise.init, 9
- genMerge, 10
- global.norm, 12
- graphic.choose, 12
- grid.norm, 13
- i.arise, 14
- m.arise, 16
- meanUnique, 18
- r.arise, 21
- spotUnique, 28
- Zscore, 32

a.arise, [3, 16](#)  
alter.unique, [4](#)  
analysis.window, [4](#)  
annotations, [5](#)

back.gui, [5](#)  
bg.correct, [6](#)

classes, [6](#)  
create.project, [7](#)  
cys.plot, [7](#)

DataSet, [6](#)  
DataSet-class, [8](#)

filter.spot, [8](#)

genArise, [9](#)  
genArise.init, [9](#)  
genMerge, [10](#)  
get.values, [11](#)  
get.Zscore, [11](#)  
global.norm, [12](#)  
graphic.choose, [12](#)  
grid.norm, [13](#)

help, [14](#)

i.arise, [14, 21](#)  
imageLimma, [15](#)

m.arise, [3, 16](#)  
ma.plot, [16](#)  
make.swap, [17](#)  
meanUnique, [18](#)

note, [18](#)

old.project, [19](#)

post.analysis, [19](#)  
principal, [20](#)  
project.select, [20](#)  
projects.select (project.select), [20](#)

r.arise, [14, 21](#)  
read.dataset, [22](#)  
read.spot, [8, 22, 27, 31](#)  
reset.history, [23](#)  
ri.plot, [24](#)  
set.grid.properties, [24](#)  
set.history.project, [25](#)  
set.path.project, [25](#)  
set.project.properties, [26](#)  
Simon, [26](#)  
single.norm, [27](#)  
Spot, [6](#)  
Spot-class, [27](#)  
spotUnique, [28](#)  
swap.select, [28](#)

trim, [29](#)

write.dataSet, [29](#)  
write.spot, [12, 17, 23, 27, 30](#)  
write.zscore, [22, 31](#)  
WT.dataset, [32](#)

Zscore, [32](#)  
Zscore.plot, [33](#)  
Zscore.points, [34](#)