

Package ‘cellxgenedp’

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Title Discover and Access Single Cell Data Sets in the cellxgene Data Portal

Version 1.3.3

Description The cellxgene data portal (<https://cellxgene.cziscience.com/>) provides a graphical user interface to collections of single-cell sequence data processed in standard ways to 'count matrix' summaries. The cellxgenedp package provides an alternative, R-based interface, allowing data discovery, viewing, and downloading.

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Collate db.R collections.R datasets.R files.R facets.R keys.R cellxgene.R utilities.R publisher_metadata.R cxg.R

Depends dplyr

Imports httr, curl, jsonlite, utils, tools, shiny, DT, rjsoncons

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Author Martin Morgan [aut, cre] (<<https://orcid.org/0000-0002-5874-8148>>),
Kayla Interdonato [aut]

Maintainer Martin Morgan <mtmorgan.bioc@gmail.com>

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collections	<i>Query cellxgene collections, datasets, and files</i>
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Description

`files_download()` retrieves one or more cellxgene files to a cache on the local system.

`links()`, `authors()` and `publisher_metadata()` are helper functions to extract 'nested' information from collections.

Usage

```
collections(cellxgene_db = db())
```

```
datasets(cellxgene_db = db())
```

```
datasets_visualize(tbl)
```

```
files(cellxgene_db = db())
```

```
files_download(tbl, dry.run = TRUE, cache.path = .cellxgene_cache_path())
```

```
links(cellxgene_db = db())
```

```
authors(cellxgene_db = db())
```

```
publisher_metadata(cellxgene_db = db())
```

Arguments

`cellxgene_db` an optional 'cellxgene_db' object, as returned by `db()`.

`tbl` a `tibble()` typically derived from `datasets(db)` or `files(db)` and containing columns `dataset_id` (for `datasets_visualize()`), or columns `dataset_id`, `file_id`, and `filetype` (for `files_download()`).

`dry.run` `logical(1)` indicating whether the (often large) file(s) in `tbl` should be downloaded to a local cache. Files are not downloaded when `dry.run = TRUE` (default).

`cache.path` `character(1)` directory in which to cache downloaded files. The directory must already exist. The default is `tools::R_user_dir("cellxgenedp", "cache")`, a package-specific path in the user home directory.

Value

Each function returns a tibble describing the corresponding component of the database.

`files_download()` returns a `character()` vector of paths to the local files.

`links()` returns a tibble of external links associated with each collection. Common links include DOI, raw data / data sources, and lab websites.

`authors()` returns a tibble of authors associated with each collection.

`publisher_metadata()` returns a tibble of publisher metadata (journal, publication date, doi) associated with each collection.

Examples

```
db <- db()

collections(db)

collections(db) |>
  dplyr::glimpse()

datasets(db) |>
  dplyr::glimpse()

## visualize the first dataset
datasets(db) |>
  dplyr::slice(1) |>
  datasets_visualize()

files(db) |>
  dplyr::glimpse()

## Not run:
files(db) |>
  dplyr::slice(1) |>
  files_download(dry.run = FALSE)

## End(Not run)

## common links to external data
links(db) |>
  dplyr::count(link_type)

## authors per collection
authors() |>
  dplyr::count(collection_id, sort = TRUE)

publisher_metadata() |>
  dplyr::glimpse()
```

cxg	<i>Shiny application for discovering, viewing, and downloading cellxgene data</i>
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Description

Shiny application for discovering, viewing, and downloading cellxgene data

Usage

```
cxg(as = c("tibble", "sce"))
```

Arguments

as	character(1) Return value when quitting the shiny application. "tibble" returns a tibble describing selected datasets (including the location on disk of the downloaded file). "sce" returns a list of dataset files imported to R as SingleCellExperiment objects.
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Value

cxg() returns either a tibble describing datasets selected in the shiny application, or a list of datasets imported into R as SingleCellExperiment objects.

Examples

```
cxg()
```

db	<i>Retrieve updated cellxgene database metadata</i>
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Description

Retrieve updated cellxgene database metadata

Usage

```
db(overwrite = .db_online() && .db_first())
```

Arguments

overwrite	logical(1) indicating whether the database of collections should be updated from the internet (the default, when internet is available and, in an interactive session, the user requests the update), or read from disk (assuming previous successful access to the internet). overwrite = FALSE might be useful for reproducibility, testing, or when working in an environment with restricted internet access.
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Details

The database is retrieved from the cellxgene data portal web site. 'collections' metadata are retrieved on each call; metadata on each collection is cached locally for re-use.

Value

db() returns an object of class 'cellxgene_db', summarizing available collections, datasets, and files.

Examples

```
db()
```

FACETS

Facets available for querying cellxgene data

Description

FACETS is a character vector of common fields used to subset cellxgene data.

facets() is used to query the cellxgene database for current values of one or all facets.

facets_filter() provides a convenient way to filter facets based on label or ontology term.

Usage

```
FACETS
```

```
facets(cellxgene_db = db(), facets = FACETS)
```

```
facets_filter(facet, key = c("label", "ontology_term_id"), value, exact = TRUE)
```

Arguments

cellxgene_db	an (optional) cellxgene_db object, as returned by db().
facets	a character() vector corresponding to one of the facets in FACETS.
facet	the column containing faceted information, e.g., sex in datasets(db).
key	character(1) identifying whether value is a label or ontology_term_id.
value	character() value of the label or ontology term to filter on. The value may be a vector with length(value) > 0 for exact matches (exact = TRUE, default), or a character(1) regular expression.
exact	logical(1) whether values match exactly (default, TRUE) or as a regular expression (FALSE).

Format

FACETS is an object of class character of length 8.

Value

`facets()` returns a tibble with columns `facet`, `label`, `ontology_term_id`, and `n`, the number of times the facet label is used in the database.

`facets_filter()` returns a logical vector with length equal to the length (number of rows) of `facet`, with `TRUE` indicating that the value of key is present in the dataset.

Examples

```
f <- facets()

## levels of each facet
f |>
  dplyr::count(facet)

## same as facets(, facets = "organism")
f |>
  dplyr::filter(facet == "organism")

db <- db()
ds <- datasets(db)

## datasets with African American females
ds |>
  dplyr::filter(
    facets_filter(self_reported_ethnicity, "label", "African American"),
    facets_filter(sex, "label", "female")
  )

## datasets with non-European, known ethnicity
facets(db, "self_reported_ethnicity")
ds |>
  dplyr::filter(
    !facets_filter(
      self_reported_ethnicity, "label", c("European", "na", "unknown")
    )
  )
```

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