

Bioconductor serialization best practices

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19 September 2019



Scenarios

Personal

- Check-pointing (e.g., for fast computation) or reproducibility

Package

- Demonstration data, e.g., for vignettes & examples
- Package-specific resource, e.g., reference data
- Project-wide benefit, e.g., EnsDb

Where

< 1 Mb

- Package-specific resource? in the package

1 - 100 Mb

- AnnotationHub or ExperimentHub

> 100 Mb

- Hmm, time to reconsider

How?

R formats

- 'Rda' files containing `data.frame`, `GRanges`, `SummarizedExperiment`, ...

Pros

- Fast and easy to load

Cons

- Only useful in *R*
- Complex (e.g., S4) objects: updated class definitions require methods to update the objects

Community-standard formats

- `csv`, `bed`, `hdf5`, ...

Pros

- Constant format, so consistent import
- Useful outside *R*

Cons

- Cost of importing or constructing complex objects 'on the fly'

Unserializing *R* objects

`readRDS()` (better than `data()` / `load()`)

- Reads the object into R
- (S4) attaches the necessary package(s)
- No automatic validation or updating

`updateObject()`

- Convention
- `BiocGenerics::updateObject` generic
- Object-specific methods defined by the developer, e.g.,
`selectMethod("updateObject", "GRanges")`

Best practices

Where?

- <1 Mb, useful in a single package or package hierarchy: package
- >1 Mb, or useful across packages: AnnotationHub or ExperimentHub

How?

- Community standard formats, unless ingestion into *R* is (time or space) expensive

Conclusions and acknowledgements

Bioconductor core team & close collaborators

- Funded by US National Institutes of Health, European Union, Chan-Zuckerberg Initiative ...

World-wide community of users & developers

[Technical](#) and [scientific](#) advisory boards



Acknowledgements



National Human Genome
Research Institute

NATIONAL CANCER INSTITUTE
Informatics Technology for
Cancer Research



Research reported in this presentation was supported by the NHGRI and NCI of the National Institutes of Health under award numbers U41HG004059, U24CA180996, and U24CA232979. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

This work was performed on behalf of the SOUND Consortium and funded under the EU H2020 Personalizing Health and Care Program, Action contract number 633974.

A portion of this work is supported by the Chan Zuckerberg Initiative DAF, an advised fund of Silicon Valley Community Foundation.