

# Package ‘Rsubread’

November 18, 2017

**Version** 1.28.0

**Date** 2017-10-25

**Title** Subread sequence alignment for R

**Author** Wei Shi and Yang Liao with contributions from Gordon Smyth, Jenny Dai and Timothy Triche, Jr.

**Maintainer** Wei Shi <shi@wehi.edu.au>, Yang Liao <liao@wehi.edu.au> and Gordon K Smyth <smyth@wehi.EDU.AU>

**Description** Provides powerful and easy-to-use tools for analyzing next-gen sequencing read data. Includes quality assessment of sequence reads, read alignment, read summarization, exon-exon junction detection, fusion detection, detection of short and long indels, absolute expression calling and SNP calling. Can be used with reads generated from any of the major sequencing platforms including Illumina GA/HiSeq/MiSeq, Roche GS-FLX, ABI SOLiD and LifeTech Ion PGM/Proton sequencers.

**URL** <http://bioconductor.org/packages/release/bioc/html/Rsubread.html>

**License** GPL-3

**biocViews** Sequencing, Alignment, SequenceMatching, RNASeq, ChIPSeq, GeneExpression, GeneRegulation, Genetics, SNP, GeneticVariability, Preprocessing, QualityControl, GenomeAnnotation, Software

**NeedsCompilation** yes

## R topics documented:

align . . . . .	2
atgcContent . . . . .	8
buildindex . . . . .	9
createAnnotationFile . . . . .	10
detectionCall . . . . .	11
detectionCallAnnotation . . . . .	12
exactSNP . . . . .	12
featureCounts . . . . .	14
findCommonVariants . . . . .	22
getInBuiltAnnotation . . . . .	23

processExons . . . . .	24
promoterRegions . . . . .	25
propmapped . . . . .	26
qualityScores . . . . .	27
removeDupReads . . . . .	28
repair . . . . .	28
RsubreadUsersGuide . . . . .	29
sam2bed . . . . .	30
txUnique . . . . .	31

<b>Index</b>	<b>32</b>
--------------	-----------

---

align	<i>Align sequence reads to a reference genome via seed-and-vote</i>
-------	---

---

## Description

align function can align both DNA and RNA sequencing reads. Subjunc is an RNA-seq aligner and it reports full alignment of each read (align reports partial alignment for exon spanning reads).

## Usage

```
align(
# index for reference sequences
index,

# input reads and output
readfile1,
readfile2=NULL,
type="rna",
input_format="gzFASTQ",
output_format="BAM",
output_file=paste(as.character(readfile1),"subread",output_format,sep="."),

# offset value added to Phred quality scores of read bases
phredOffset=33,

# thresholds for mapping
nsubreads=10,
TH1=3,
TH2=1,
maxMismatches=3,

# unique mapping and multi-mapping
unique=FALSE,
nBestLocations=1,

# indel detection
indels=5,
complexIndels=FALSE,

# read trimming
```

```
nTrim5=0,
nTrim3=0,

# distance and orientation of paired end reads
minFragLength=50,
maxFragLength=600,
PE_orientation="fr",

# number of CPU threads
nthreads=1,

# read group
readGroupID=NULL,
readGroup=NULL,

# color space reads
color2base=FALSE,

# dynamic programming
DP_GapOpenPenalty=-1,
DP_GapExtPenalty=0,
DP_MismatchPenalty=0,
DP_MatchScore=2,

# detect structural variants
detectSV=FALSE,

# gene annotation
useAnnotation=FALSE,
annot.inbuilt="mm10",
annot.ext=NULL,
isGTF=FALSE,
GTF.featureType="exon",
GTF.attrType="gene_id",
chrAliases=NULL)

subjunc(
# index for reference sequences
index,

# input reads and output
readfile1,
readfile2=NULL,
input_format="gzFASTQ",
output_format="BAM",
output_file=paste(as.character(readfile1),"subjunc",output_format,sep="."),

# offset value added to Phred quality scores of read bases
phredOffset=33,

# thresholds for mapping
nsubreads=14,
```

```
TH1=1,
TH2=1,
maxMismatches=3,

# unique mapping and multi-mapping
unique=FALSE,
nBestLocations=1,

# indel detection
indels=5,
complexIndels=FALSE,

# read trimming
nTrim5=0,
nTrim3=0,

# distance and orientation of paired end reads
minFragLength=50,
maxFragLength=600,
PE_orientation="fr",

# number of CPU threads
nthreads=1,

# read group
readGroupID=NULL,
readGroup=NULL,

# color space reads
color2base=FALSE,

# dynamic programming
DP_GapOpenPenalty=-1,
DP_GapExtPenalty=0,
DP_MismatchPenalty=0,
DP_MatchScore=2,

# detect all junctions including gene fusions
reportAllJunctions=FALSE,

# gene annotation
useAnnotation=FALSE,
annot.inbuilt="mm10",
annot.ext=NULL,
isGTF=FALSE,
GTF.featureType="exon",
GTF.attrType="gene_id",
chrAliases=NULL)
```

### Arguments

`index` character string giving the basename of index file. Index files should be located in the current directory.

readfile1	a character vector including names of files that include sequence reads to be aligned. For paired-end reads, this gives the list of files including first reads in each library. File format is FASTQ/FASTA by default. See <code>input_format</code> option for more supported formats.
readfile2	a character vector giving names of files that include second reads in paired-end read data. Files included in <code>readfile2</code> should be in the same order as their mate files included in <code>readfile1</code> . NULL by default.
type	a character string or an integer giving the type of sequencing data. Possible values include <code>rna</code> (or 0; RNA-seq data) and <code>dna</code> (or 1; genomic DNA-seq data such as WGS, WES, ChIP-seq data etc.). Character strings are case insensitive.
input_format	character string specifying format of read input files. <code>gzFASTQ</code> by default (this also includes FASTQ, FASTA and gzipped FASTA formats). Other supported formats include SAM and BAM. Character values are case insensitive.
output_format	character string specifying format of output file. BAM by default. Acceptable formats include SAM and BAM.
output_file	a character vector specifying names of output files. By default, names of output files are set as the file names provided in <code>readfile1</code> added with an suffix string.
phredOffset	numeric value added to base-calling Phred scores to make quality scores (represented as ASCII letters). Possible values include 33 and 64. By default, 33 is used.
nsubreads	numeric value giving the number of subreads extracted from each read.
TH1	numeric value giving the consensus threshold for reporting a hit. This is the threshold for the first reads if paired-end read data are provided.
TH2	numeric value giving the consensus threshold for the second reads in paired-end data.
maxMismatches	numeric value giving the maximum number of mis-matched bases allowed in the alignment. 3 by default. Mis-matches found in soft-clipped bases are not counted.
unique	logical indicating if only uniquely mapped reads should be reported. A uniquely mapped read has one single mapping location that has less mis-matched bases than any other candidate locations. By default, multi-mapping reads will be reported in addition to uniquely mapped reads. Number of alignments reported for each multi-mapping read is determined by the <code>nBestLocations</code> parameter.
nBestLocations	numeric value specifying the maximal number of equally-best mapping locations that will be reported for a multi-mapping read. 1 by default. The allowed value is between 1 to 16 (inclusive). In the mapping output, 'NH' tag is used to indicate how many alignments are reported for the read and 'HI' tag is used for numbering the alignments reported for the same read. This argument is only applicable when <code>unique</code> option is set to FALSE.
indels	numeric value giving the maximum number of insertions/deletions allowed during the mapping. 5 by default.
complexIndels	logical indicating if complex indels will be detected. If TRUE, the program will try to detect multiple short indels that occurs concurrently in a small genomic region (indels could be as close as 1bp apart).
nTrim5	numeric value giving the number of bases trimmed off from 5' end of each read. 0 by default.
nTrim3	numeric value giving the number of bases trimmed off from 3' end of each read. 0 by default.

<code>minFragLength</code>	numeric value giving the minimum fragment length. 50 by default.
<code>maxFragLength</code>	numeric value giving the maximum fragment length. 600 by default.
<code>PE_orientation</code>	character string giving the orientation of the two reads from the same pair. It has three possible values including <code>fr</code> , <code>ff</code> and <code>rf</code> . Letter <code>f</code> denotes the forward strand and letter <code>r</code> the reverse strand. <code>fr</code> by default (ie. the first read in the pair is on the forward strand and the second read on the reverse strand).
<code>nthreads</code>	numeric value giving the number of threads used for mapping. 1 by default.
<code>readGroupID</code>	a character string giving the read group ID. The specified string is added to the read group header field and also be added to each read in the mapping output. NULL by default.
<code>readGroup</code>	a character string in the format of <code>tag:value</code> . This string will be added to the read group (RG) header in the mapping output. NULL by default.
<code>color2base</code>	logical. If TRUE, color-space read bases will be converted to base-space bases in the mapping output. Note that the mapping itself will still be performed at color-space. FALSE by default.
<code>DP_GapOpenPenalty</code>	a numeric value giving the penalty for opening a gap when using the Smith-Waterman dynamic programming algorithm to detect insertions and deletions. The Smith-Waterman algorithm is only applied for those reads which are found to contain insertions or deletions. -1 by default.
<code>DP_GapExtPenalty</code>	a numeric value giving the penalty for extending the gap, used by the Smith-Waterman algorithm. 0 by default.
<code>DP_MismatchPenalty</code>	a numeric value giving the penalty for mismatches, used by the Smith-Waterman algorithm. 0 by default.
<code>DP_MatchScore</code>	a numeric value giving the score for matches used by the Smith-Waterman algorithm. 2 by default.
<code>detectSV</code>	logical indicating if structural variants (SVs) will be detected during read mapping. See below for more details.
<code>reportAllJunctions</code>	logical indicating if all discovered junctions will be reported. This includes exon-exon junctions and also gene fusions. Presence of donor/receptor sites is not required when <code>reportAllJunctions</code> is TRUE. This option should only be used for RNA-seq data.
<code>useAnnotation</code>	logical indicating if gene annotation information will be used in the mapping. FALSE by default.
<code>annot.inbuilt</code>	a character string specifying an in-built annotation used for read summarization. It has four possible values including <code>"mm10"</code> , <code>"mm9"</code> , <code>"hg38"</code> and <code>"hg19"</code> , corresponding to the NCBI RefSeq annotations for genomes 'mm10', 'mm9', 'hg38' and 'hg19', respectively. <code>"mm10"</code> by default. See <a href="#">featureCounts</a> function for more details on the in-built annotations.
<code>annot.ext</code>	A character string giving name of a user-provided annotation file or a data frame including user-provided annotation data. If the annotation is in GTF format, it can only be provided as a file. If it is in SAF format, it can be provided as a file or a data frame. If an annotation is provided via <code>annot.ext</code> , the <code>annot.inbuilt</code> parameter will be ignored. See <a href="#">featureCounts</a> function for more details about this parameter.

<code>isGTF</code>	logical indicating if the annotation provided via the <code>annot.ext</code> argument is in GTF format or not. FALSE by default. This option is only applicable when <code>annot.ext</code> is not NULL.
<code>GTF.featureType</code>	a character string denoting the type of features that will be extracted from a GTF annotation. "exon" by default. This argument is only applicable when <code>isGTF</code> is TRUE.
<code>GTF.attrType</code>	a character string denoting the type of attributes in a GTF annotation that will be used to group features. "gene_id" by default. The grouped features are called meta-features. For instance, a feature can be an exon and several exons can be grouped into a gene (meta-feature). This argument is only applicable when <code>isGTF</code> is TRUE.
<code>chrAliases</code>	a character string providing the name of a comma-delimited text file that includes aliases of chromosome names. This file should contain two columns. First column contains names of chromosomes included in the SAF or GTF annotation and second column contains corresponding names of chromosomes in the reference genome. No column headers should be provided. Also note that chromosome names are case sensitive. This file can be used to match chromosome names between the annotation and the reference genome.

## Details

The `align` function implements the Subread aligner (Liao et al., 2013) that uses a new mapping paradigm called "seed-and-vote". Subread is general-purpose aligner that can be used to align both genomic DNA-seq reads and RNA-seq reads.

Subjunc is designed for mapping RNA-seq reads. The major difference between Subjunc and Subread is that Subjunc reports discovered exon-exon junctions and it also performs full alignments for every read including exon-spanning reads. Subread uses the largest mappable regions in the reads to find their mapping locations. The seed-and-vote paradigm has been found to be not only more accurate than the conventional seed-and-extend (adopted by most aligners) in read mapping, but it is a lot more efficient (Liao et al., 2013).

Both Subread and Subjunc can be used to align reads generated from major sequencing platforms including Illumina GA/HiSeq, ABI SOLiD, Roche 454 and Ion Torrent sequencers. Note that to map color-space reads (e.g. SOLiD reads), a color-space index should be built for the reference genome (see [buildindex](#) for details).

Subread and Subjunc have adjustable memory usage. See [buildindex](#) function for more details.

Mapping performance is largely determined by the number of subreads extracted from each read `nsubreads` and the consensus threshold `TH1` (also `TH2` for paired-end read data). Default settings are recommended for most of the read mapping tasks.

Subjunc requires donor/receptor sites to be present when detecting exon-exon junctions. It can detect up to four junction locations in each exon-spanning read.

`detectSV` option should be used for SV detection in genomic DNA sequencing data. For RNA-seq data, users may use `subjunc` with the `reportAllJunctions` option to detect SVs (and also junctions). For each library, breakpoints detected from SV events will be saved to a file with suffix name `breakpoints.txt`, which includes chromosomal coordinates of SV breakpoints and numbers of supporting reads. The BAM/SAM output includes extra fields to describe the complete alignments of breakpoint-containing reads. For a breakpoint-containing read, mapping of its major sequence segment is described in the main fields of BAM/SAM output whereas mapping of its minor sequence segment, which does not map along with the major segment due to the presence of a breakpoint, is described in the extra fields including `'CC'` (Chr), `'CP'` (Position), `'CG'` (CIGAR) and `'CT'` (strand). Note that each breakpoint-containing read occupies only one row in BAM/SAM output.

**Value**

No value is produced but SAM or BAM format files are written to the current working directory. For Subjunc, BED files including discovered exon-exon junctions are also written to the current working directory.

**Author(s)**

Wei Shi and Yang Liao

**References**

Yang Liao, Gordon K Smyth and Wei Shi. The Subread aligner: fast, accurate and scalable read mapping by seed-and-vote. *Nucleic Acids Research*, 41(10):e108, 2013.

**Examples**

```
# Build an index for the artificial sequence included in file 'reference.fa'.
library(Rsubread)
ref <- system.file("extdata", "reference.fa", package="Rsubread")
buildindex(basename="./reference_index", reference=ref)

# align a sample read dataset ('reads.txt') to the sample reference
reads <- system.file("extdata", "reads.txt.gz", package="Rsubread")
align(index="./reference_index", readfile1=reads, output_file="./Rsubread_alignment.BAM", phredOffset=64)
```

---

atgcContent

---

*Calculate percentages of nucleotides A, T, G and C in a sequencing read datafile*


---

**Description**

Calculate percentages of nucleotides A, T, G and C

**Usage**

```
atgcContent(filename, basewise=FALSE)
```

**Arguments**

filename	character string giving the name of input FASTQ/FASTA file
basewise	logical. If TRUE, nucleotide percentages will be calculated for each base position in the read across all the reads. By default, percentages are calculated for the entire dataset.

**Details**

Sequencing reads could contain letter "N" besides "A", "T", "G" and "C". Percentage of "N" in the read dataset is calculated as well.

The basewise calculation is useful for examining the GC bias towards the base position in the read. By default, the percentages of nucleotides in the entire dataset will be reported.

**Value**

A named vector containing percentages for each nucleotide type if `basewise` is `FALSE`. Otherwise, a data matrix containing nucleotide percentages for each base position of the reads.

**Author(s)**

Zhiyin Dai and Wei Shi

---

buildindex	<i>Build index for a reference genome</i>
------------	---

---

**Description**

An index needs to be built before read mapping can be performed. This function creates a hash table for the reference genome, which can then be used by Subread and Subjunc aligners for read alignment.

**Usage**

```
buildindex(basename, reference, gappedIndex=TRUE, indexSplit=TRUE, memory=8000,
           TH_subread=100, colorspace=FALSE)
```

**Arguments**

<code>basename</code>	character string giving the basename of created index files.
<code>reference</code>	character string giving the name of a FASTA-format file that includes sequences of all chromosomes and contigs.
<code>gappedIndex</code>	logical. If <code>FALSE</code> , 16mers (subreads) will be extracted from every chromosomal location of a reference genome and then they will be used to build a hash table index. By default( <code>TRUE</code> ), subreads are extracted in every three bases from the genome.
<code>indexSplit</code>	logical. If <code>TRUE</code> , the built index is allowed to be splitted into multiple segments. The number of such segments is determined by <code>memory</code> value, genome size and permitting of gaps between subreads( <code>gappedIndex</code> ). If <code>indexSplit</code> is set to <code>FALSE</code> , a single-segment index (no splitting) will be generated regardless of what value is chosen for <code>memory</code> .
<code>memory</code>	numeric value specifying the amount of memory to be requested in megabytes. 8000 MB by default.
<code>TH_subread</code>	numeric value specifying the threshold for removing highly repetitive subreads (16bp mers). 100 by default. Subreads will be excluded from the index if they occur more than threshold number of times in the genome.
<code>colorspace</code>	logical. If <code>TRUE</code> , a color space index will be built. Otherwise, a base space index will be built.

## Details

This function generates a hash table (an index) for a reference genome, in which keys are subreads (16mers) and values are their chromosomal locations in the reference genome. By default, subreads will be extracted in every three bases from a reference genome. However, if `gappedIndex` is set to `FALSE`, then subreads will be extracted from every chromosomal location of genome for index building. The built index can then be used by Subread ([align](#)) and [subjunc](#) aligners to map reads (Liao et al. 2013).

Highly repetitive subreads (or uninformative subreads) are excluded from the hash table so as to reduce mapping ambiguity. `TH_subread` specifies the maximal number of times a subread is allowed to occur in the reference genome to be included in hash table.

The built index might be splitted into multiple segments if its size is greater than memory value. The number of such segments is dependent on memory value, size of reference genome and whether gaps are allowed between subreads extracted from genome. Only one segment is loaded into memory at any time when read alignment is being carried out. The larger the memory value, the faster the read mapping will be. If `indexSplit` is set to `FALSE`, the index will not be splitted and this will enable maximum mapping speed to be achieved.

The index needs to be built only once and it can then be re-used in the subsequent alignments.

## Value

No value is produced but index files are written to the current working directory.

## Author(s)

Wei Shi and Yang Liao

## References

Yang Liao, Gordon K Smyth and Wei Shi. The Subread aligner: fast, accurate and scalable read mapping by seed-and-vote. *Nucleic Acids Research*, 41(10):e108, 2013.

## Examples

```
# Build an index for the artificial sequence included in file 'reference.fa'
library(Rsubread)
ref <- system.file("extdata", "reference.fa", package="Rsubread")
buildindex(basename="./reference_index", reference=ref)
```

---

`createAnnotationFile` *Create an annotation file from a GRanges object, suitable for featureCounts()*

---

## Description

Any of `rtracklayer::import.bed('samplesubjunc.bed')`, `unlist(spliceGraph(TxDb))`, `transcripts(TxDb)`, `exons(TxDb)`, or `features(FDB)` will produce a `GRanges` object containing usable features for read counting.

This function converts a suitably streamlined `GRanges` object into annotations which can then be used by `featureCounts()` to quickly count aligned reads.

The `GRanges` object must contain an `elementMetadata` column named `'id'`.

**Usage**

```
createAnnotationFile(GR)
write.Rsubread(GR)
```

**Arguments**

GR                    The GRanges object to convert to an Rsubread annotation file

**Value**

A data frame with five columns named GeneID, Chr, Start, End and Strand.

**Author(s)**

Tim Triche, Jr. and Wei Shi

**Examples**

```
## Not run:
library(TxDb.Hsapiens.UCSC.hg19.lincRNAsTranscripts)
hg19LincRNAs <- transcripts(TxDb.Hsapiens.UCSC.hg19.lincRNAsTranscripts)
names(values(hg19LincRNAs)) <- gsub('tx_id', 'id', names(values(hg19LincRNAs)))
annot_for_featureCounts <- createAnnotationFile(hg19LincRNAs)

## End(Not run)
```

---

detectionCall                    *Determine detection p values for each gene in an RNA-seq dataset*

---

**Description**

Use GC content adjusted background read counts to determine the detection p values for each gene

**Usage**

```
detectionCall(dataset, species="hg", plot=FALSE)
```

**Arguments**

dataset                a character string giving the filename of a SAM format file, which is the output of read alignment.

species                a character string specifying the species. Options are hg and mm.

plot                    logical, indicating whether a density plot of detection p values will be generated.

**Value**

A data frame which includes detection p values and annotation information for each genes.

**Author(s)**

Zhiyin Dai and Wei Shi

---

detectionCallAnnotation

*Generate annotation data used for calculating detection p values*

---

### Description

This is for internal use only.

### Usage

```
detectionCallAnnotation(species="hg", binsize=2000)
```

### Arguments

species	character string specifying the species to analyse
binsize	binsize of intergenic region

### Details

This is an internal function and should not be called by users directly.

It takes as input the annotation files produced by [processExons](#) function, calculates GC percentages for each exon of genes and also for intergenic regions and add GC info into the annotations. The new annotation data are then saved to files which can be used by [detectionCall](#) function for calling absolutely expressed genes.

### Value

Two annotation files, which contain GC content for exons of genes and for intergenic regions respectively, are written to the current working directory. This function returns a NULL object.

### Author(s)

Zhiyin Dai and Wei Shi

---

exactSNP

*exactSNP - an accurate and efficient SNP caller*

---

### Description

Measure background noises and perform Fisher's Exact tests to detect SNPs.

### Usage

```
exactSNP(readFile, isBAM=FALSE, refGenomeFile, SNPAnnotationFile=NULL,
outputFile=paste(readFile, ".exactSNP.VCF", sep=""), qvalueCutoff=12, minAllelicFraction=0,
minAllelicBases=1, minReads=1, maxReads=3000, minBaseQuality=13, nTrimmedBases=3, nthreads=1)
```

**Arguments**

readFile	a character string giving the name of a file including read mapping results. This function takes as input a SAM file by default. If a BAM file is provided, the isBAM argument should be set to TRUE.
isBAM	logical indicating if the file provided via readFile is a BAM file. FALSE by default.
refGenomeFile	a character string giving the name of a file that includes reference sequences (FASTA format).
SNPAnnotationFile	a character string giving the name of a VCF-format file that includes annotated SNPs. Such annotation files can be downloaded from public databases such as the dbSNP database. Incorporating known SNPs into SNP calling has been found to be helpful. However note that the annotated SNPs may or may not be called for the sample being analyzed.
outputFile	a character string giving the name of the output file to be generated by this function. The output file includes all the reported SNPs (in VCF format). It includes discovered indels as well.
qvalueCutoff	a numeric value giving the q-value cutoff for SNP calling at sequencing depth of 50X. 12 by default. The q-value is calculated as $-\log_{10}(p)$ , where p is the p-value yielded from the Fisher's Exact test. Note that this function automatically adjusts the q-value cutoff for each chromosomal location according to its sequencing depth, based on this cutoff.
minAllelicFraction	a numeric value giving the minimum fraction of allelic bases out of all read bases included at a chromosomal location required for SNP calling. Its value must be within 0 and 1. 0 by default.
minAllelicBases	a numeric value giving the minimum number of allelic (mis-matched) bases a SNP must have at a chromosomal location. 1 by default.
minReads	a numeric value giving the minimum number of mapped reads a SNP-containing location must have (ie. the minimum coverage). 1 by default.
maxReads	Specify the maximum number of mapped reads a SNP-containing location can have. 3000 by default. Any location having more than this threshold number of reads will not be considered for SNP calling. This option is useful for removing PCR artefacts.
minBaseQuality	a numeric value giving the minimum base quality score (Phred score) read bases should satisfy before being used for SNP calling. 13 by default (corresponding to base calling p value of 0.05). Read bases with quality scores less than 13 will be excluded from analysis.
nTrimmedBases	a numeric value giving the number of bases trimmed off from each end of the read. 3 by default.
nthreads	a numeric value giving the number of threads/CPU's used. 1 by default.

**Details**

This function takes as input a SAM/BAM format file, measures local background noise for each chromosomal location and then performs Fisher's exact tests to find statistically significant SNPs .

This function implements a novel algorithm for discovering SNPs. This algorithm is comparable with or better than existing SNP callers, but it is fast more efficient. It can be used to call SNPs for

individual samples (ie. no control samples are required). Detail of the algorithm is described in a manuscript which is currently under preparation.

**Value**

No value is produced but but a VCF format file is written to the current working directory. This file contains detailed information for discovered SNPs including chromosomal locations, reference bases, alternative bases, read coverages, allele frequencies and p values.

**Author(s)**

Yang Liao and Wei Shi

---

featureCounts	<i>featureCounts: a general-purpose read summarization function</i>
---------------	---

---

**Description**

This function assigns mapped sequencing reads to genomic features

**Usage**

```
featureCounts(files,  
  
# annotation  
annot.inbuilt="mm10",  
annot.ext=NULL,  
isGTFAnnotationFile=FALSE,  
GTF.featureType="exon",  
GTF.attrType="gene_id",  
chrAliases=NULL,  
  
# level of summarization  
useMetaFeatures=TRUE,  
  
# overlap between reads and features  
allowMultiOverlap=FALSE,  
minOverlap=1,  
fracOverlap=0,  
fracOverlapFeature=0,  
largestOverlap=FALSE,  
readExtension5=0,  
readExtension3=0,  
read2pos=NULL,  
  
# multi-mapping reads  
countMultiMappingReads=FALSE,  
  
# fractional counting  
fraction=FALSE,
```

```
# long reads
isLongRead=FALSE,

# read filtering
minMQS=0,
splitOnly=FALSE,
nonSplitOnly=FALSE,
primaryOnly=FALSE,
ignoreDup=FALSE,

# strandness
strandSpecific=0,

# exon-exon junctions
juncCounts=FALSE,
genome=NULL,

# parameters specific to paired end reads
isPairedEnd=FALSE,
requireBothEndsMapped=FALSE,
checkFragLength=FALSE,
minFragLength=50,
maxFragLength=600,
countChimericFragments=TRUE,
autosort=TRUE,

# number of CPU threads
nthreads=1,

# read group
byReadGroup=FALSE,

# miscellaneous
maxMOp=10,
reportReads=NULL,
tmpDir=".",
verbose=FALSE)
```

## Arguments

<code>files</code>	a character vector giving names of input files containing read mapping results. The files can be in either SAM format or BAM format. The file format is automatically detected by the function.
<code>annot.inbuilt</code>	a character string specifying an in-built annotation used for read summarization. It has four possible values including "mm10", "mm9", "hg38" and "hg19", corresponding to the NCBI RefSeq annotations for genomes 'mm10', 'mm9', 'hg38' and 'hg19', respectively. "mm10" by default. The in-built annotation has a SAF format (see below).
<code>annot.ext</code>	A character string giving name of a user-provided annotation file or a data frame including user-provided annotation data. If the annotation is in GTF format, it can only be provided as a file. If it is in SAF format, it can be provided as a file or a data frame. See below for more details about SAF format annotation.

annot.ext will override annot.inbuilt if they are both provided.

isGTFAnnotationFile	logical indicating whether the annotation provided via the annot.ext argument is in GTF format or not. FALSE by default. This option is only applicable when annot.ext is not NULL.
GTF.featureType	a character string giving the feature type used to select rows in the GTF annotation which will be used for read summarization. "exon" by default. This argument is only applicable when isGTFAnnotationFile is TRUE. Feature types can be found in the third column of a GTF annotation.
GTF.attrType	a character string giving the attribute type in the GTF annotation which will be used to group features (eg. exons) into meta-features (eg. genes). "gene_id" by default. This argument is only applicable when isGTFAnnotationFile is TRUE. Attributes can be found in the ninth column of a GTF annotation.
chrAliases	a character string giving the name of a chromosome name alias file. This should be a two-column comma-delimited text file. Chromosome name aliases included in this file are used to match chr names in annotation with those in the reads. First column in the file should include chr names in the annotation and second column should include chr names in the reads. Chr names are case sensitive. No column header should be included in the file.
useMetaFeatures	logical indicating whether the read summarization should be performed at the feature level (eg. exons) or meta-feature level (eg. genes). If TRUE, features in the annotation (each row is a feature) will be grouped into meta-features, using their values in the GeneID column in the SAF-format annotation file or using the GTF.attrType attribute in the GTF-format annotation file, and then reads will be assigned to the meta-features instead of the features. See below for more details.
allowMultiOverlap	logical indicating if a read is allowed to be assigned to more than one feature (or meta-feature) if it is found to overlap with more than one feature (or meta-feature). FALSE by default.
minOverlap	integer giving the minimum number of overlapped bases required for assigning a read to a feature (or a meta-feature). For assignment of read pairs (fragments), number of overlapping bases from each read in the same pair will be summed. If a negative value is provided, then a gap of up to specified size will be allowed between read and the feature that the read is assigned to. 1 by default.
fracOverlap	numeric giving minimum fraction of overlapping bases in a read that is required for read assignment. Value should be within range [0,1]. 0 by default. Number of overlapping bases is counted from both reads if paired end. Both this option and minOverlap option need to be satisfied before a read can be assigned.
fracOverlapFeature	numeric giving minimum fraction of bases included in a feature that is required for overlapping with a read or a read pair. Value should be within range [0,1]. 0 by default.
largestOverlap	If TRUE, a read (or read pair) will be assigned to the feature (or meta-feature) that has the largest number of overlapping bases, if the read (or read pair) overlaps with multiple features (or meta-features).
readExtension5	integer giving the number of bases extended upstream from 5' end of each read. 0 by default.

readExtension3	integer giving the number of bases extended downstream from 3' end of each read. 0 by default.
read2pos	Specifying whether each read should be reduced to its 5' most base or 3' most base. It has three possible values: NULL, 5 (denoting 5' most base) and 3 (denoting 3' most base). The default value is NULL. With the default value, the whole read is used for summarization. When read2pos is set to 5 (or 3), read summarization will be performed based on the 5' (or 3') most base position. read2pos can be used together with readExtension5 and readExtension3 parameters to set any desired length for reads.
countMultiMappingReads	logical indicating if multi-mapping reads/fragments should be counted, FALSE by default. If TRUE, a multi-mapping read will be counted up to N times if it has N reported mapping locations. This function uses the 'NH' tag to find multi-mapping reads.
fraction	logical indicating if fractional counts are produced for multi-mapping reads and/or multi-overlapping reads. FALSE by default. See below for more details.
isLongRead	logical indicating if input data contain long reads. This option should be set to TRUE if counting Nanopore or PacBio long reads.
minMQS	integer giving the minimum mapping quality score a read must satisfy in order to be counted. For paired-end reads, at least one end should satisfy this criteria. 0 by default.
splitOnly	logical indicating whether only split alignments (their CIGAR strings contain letter 'N') should be included for summarization. FALSE by default. Example split alignments are exon-spanning reads from RNA-seq data. useMetaFeatures should be set to FALSE and allowMultiOverlap should be set to TRUE, if the purpose of summarization is to assign exon-spanning reads to all their overlapping exons.
nonSplitOnly	logical indicating whether only non-split alignments (their CIGAR strings do not contain letter 'N') should be included for summarization. FALSE by default.
primaryOnly	logical indicating if only primary alignments should be counted. Primary and secondary alignments are identified using bit 0x100 in the Flag field of SAM/BAM files. If TRUE, all primary alignments in a dataset will be counted no matter they are from multi-mapping reads or not (ie. countMultiMappingReads is ignored).
ignoreDup	logical indicating whether reads marked as duplicates should be ignored. FALSE by default. Read duplicates are identified using bit 0x400 in the FLAG field in SAM/BAM files. The whole fragment (read pair) will be ignored if paired end.
strandSpecific	integer indicating if strand-specific read counting should be performed. It has three possible values: 0 (unstranded), 1 (stranded) and 2 (reversely stranded). 0 by default.
juncCounts	logical indicating if number of reads supporting each exon-exon junction will be reported. Junctions are identified from those exon-spanning reads in input data. FALSE by default.
genome	a character string giving the name of a FASTA-format file that includes the reference sequences used in read mapping that produced the provided SAM/BAM files. NULL by default. This argument should only be used when juncCounts is TRUE. Note that providing reference sequences is optional when juncCounts is set to TRUE.
isPairedEnd	logical indicating if paired-end reads are used. If TRUE, fragments (templates or read pairs) will be counted instead of individual reads. FALSE by default.

requireBothEndsMapped	logical indicating if both ends from the same fragment are required to be successfully aligned before the fragment can be assigned to a feature or meta-feature. This parameter is only applicable when isPairedEnd is TRUE.
checkFragLength	logical indicating if the two ends from the same fragment are required to satisfy the fragment length criteria before the fragment can be assigned to a feature or meta-feature. This parameter is only applicable when isPairedEnd is TRUE. The fragment length criteria are specified via minFragLength and maxFragLength.
minFragLength	integer giving the minimum fragment length for paired-end reads. 50 by default.
maxFragLength	integer giving the maximum fragment length for paired-end reads. 600 by default. minFragLength and maxFragLength are only applicable when isPairedEnd is TRUE. Note that when a fragment spans two or more exons, the observed fragment length might be much bigger than the nominal fragment length.
countChimericFragments	logical indicating whether a chimeric fragment, which has its two reads mapped to different chromosomes, should be counted or not. TRUE by default.
autosort	logical specifying if the automatic read sorting is enabled. This option is only applicable for paired-end reads. If TRUE, reads will be automatically sorted by their names if reads from the same pair are found not to be located next to each other in the input. No read sorting will be performed if there are no such reads found.
nthreads	integer giving the number of threads used for running this function. 1 by default.
byReadGroup	logical indicating if read counting will be performed for each individual read group. FALSE by default.
maxMOp	integer giving the maximum number of 'M' operations (matches or mis-matches) allowed in a CIGAR string. 10 by default. Both 'X' and '=' operations are treated as 'M' and adjacent 'M' operations are merged in the CIGAR string.
reportReads	output detailed read assignment results for each read (or fragment if paired end). The detailed assignment results can be saved in three different formats including CORE, SAM and BAM (note that these values are case sensitive). Default value of this option is NULL, indicating not to report assignment results for reads. CORE format represents a tab-delimited text file that contains four columns including read name, status (assigned or the reason if not assigned), number of targets and target list. A target is a feature or a meta-feature. Items in the target lists is separated by comma. If a read is not assigned, its number of targets will be set as -1. For each input file, a text file is generated and its name is the input file name added with '.featureCounts'. When SAM or BAM format is specified, the detailed assignment results will be saved to SAM and BAM format files. Names of generated files are the input file names added with '.featureCounts.sam' or '.featureCounts.bam'. Three tags are used to describe read assignment results: XS, XN and XT. Tag XS gives the assignment status. Tag XN gives number of targets. Tag XT gives comma separated target list.
tmpDir	a character string specifying the directory under which intermediate files are saved (later removed). By default, current working directory is used.
verbose	logical indicating if verbose information for debugging will be generated. This may include information such as unmatched chromosomes/contigs between reads and annotation.

## Details

featureCounts is a general-purpose read summarization function, which assigns to the genomic features (or meta-features) the mapped reads that were generated from genomic DNA and RNA sequencing.

This function takes as input a set of files containing read mapping results output from a read aligner (e.g. [align](#) or [subjunc](#)), and then assigns mapped reads to genomic features. Both SAM and BAM format input files are accepted.

featureCounts accepts two annotation formats: SAF (Simplified Annotation Format) and GTF/GFF formats. Specification of GTF/GFF format can be found at <https://genome.ucsc.edu/FAQ/FAQformat.html>. SAF is a simplified annotation format and below shows an example of this format:

```
GeneID Chr Start End Strand
497097 chr1 3204563 3207049 -
497097 chr1 3411783 3411982 -
497097 chr1 3660633 3661579 -
100503874 chr1 3637390 3640590 -
100503874 chr1 3648928 3648985 -
100038431 chr1 3670236 3671869 -
...
```

SAF annotation includes the following five required columns: GeneID, Chr, Start, End and Strand. The GeneID column includes identifiers of features. These identifiers can be integer or character string. The Chr column includes chromosome names of features and these names should match the chromosome names included in the provided SAM/BAM files. The Start and End columns include start and end coordinates of features, respectively. Both start and end coordinates are inclusive. The Strand column indicates the strand of features ("+" or "-"). Column names in a SAF annotation should be the same as those shown in the above example (case insensitive). Columns can be in any order. Extra columns are allowed to be added into the annotation.

In-built annotations, which were generated based on NCBI RefSeq gene annotations, are provided to facilitate convenient read summarization. We provide in-built annotations for the following genomes: "hg38", "hg19", "mm10" and "mm9". The content of in-built annotations can be accessed via the [getInBuiltAnnotation](#) function. These annotations have a SAF format.

The in-built annotations are a modified version of NCBI RefSeq gene annotations. We downloaded the RefSeq gene annotations from NCBI ftp server (eg. RefSeq annotation for mm10 was downloaded from [ftp://ftp.ncbi.nlm.nih.gov/genomes/M\\_musculus/ARCHIVE/BUILD.38.1/mapview/seq\\_gene.md.gz](ftp://ftp.ncbi.nlm.nih.gov/genomes/M_musculus/ARCHIVE/BUILD.38.1/mapview/seq_gene.md.gz)). We then used these annotations to create our in-built annotations. For each gene, we used its CDS (coding DNA sequence) and UTR (untranslated) regions provided in the original annotation to construct a list of exons, by merging and concatenating all CDSs and UTRs belonging to the same gene. Exons within each gene include all chromosomal bases included in the original CDS and UTR regions, but they include each base only once (they might be included multiple times in the original CDSs and UTRs). Also, exons within the same gene do not overlap with each other.

Users may provide an external annotation for read summarization via the `annot.ext` argument. If the external annotation is in SAF format, it can be provided as either a `data.frame` or a tab-delimited text file with proper column names included. If it is in GTF/GFF format, it should be provided as a file only (and `isGTFAnnotationFile` should be set to TRUE).

featureCounts function uses the `GTF.attrType` attribute in a GTF/GFF annotation to group features to form meta-features when performing read summarization at meta-feature level.

The argument `useMetaFeatures` specifies whether read summarization should be performed at feature level or at meta-feature level. A feature represents a continuous genomic region and a

meta-feature is a group of features. For instance, an exon is a feature and a gene comprising one or more exons is a meta-feature. To assign reads to meta-features, featureCounts firstly groups into meta-features the features that have the same gene identifiers. featureCounts looks for gene identifiers in GeneID column of a SAF annotation or by using GTF .attrType attribute in a GTF/GFF annotation. Then for each read featureCounts searches for meta-features that have at least one feature that overlaps with the read. A read might be found to overlap with more than one feature within the same meta-feature (eg. an exon-spanning read overlaps with more than one exon from the same gene), however this read will still be counted only once for the meta-feature.

RNA-seq reads are often summarized to meta-features to produce read counts for genes. Further downstream analysis can then be carried out to discover differentially expressed genes. Feature-level summarization of RNA-seq data often yields exon-level read counts, which is useful for investigating alternative splicing of genes.

featureCounts provides multiple options to count multi-mapping reads (reads mapping to more than one location in the reference genome). Users can choose to ignore such reads by setting countMultiMappingReads to FALSE, or fully count every alignment reported for a multi-mapping read by setting countMultiMappingReads to TRUE (each alignment carries 1 count), or count each alignment fractionally by setting both countMultiMappingReads and fraction to TRUE (each alignment carries  $1/x$  count where  $x$  is the total number of alignments reported for the read).

featureCounts also provides multiple options to count multi-overlapping reads (reads overlapping with more than one meta-feature when conducting meta-feature-level summarization or overlapping with more than one feature when conducting feature-level summarization). Users can choose to ignore such reads by setting allowMultiOverlap to FALSE, or fully count them for each overlapping meta-feature/feature by setting allowMultiOverlap to TRUE (each overlapping meta-feature/feature receives a count of 1 from a read), or assign a fractional count to each overlapping meta-feature/feature by setting both allowMultiOverlap and fraction to TRUE (each overlapping meta-feature/feature receives a count of  $1/y$  from a read where  $y$  is the total number of meta-features/features overlapping with the read).

If a read is both multi-mapping and multi-overlapping, then each overlapping meta-feature/feature will receive a fractional count of  $1/(x*y)$  if countMultiMappingReads, allowMultiOverlap and fraction are all set to TRUE.

When isPairedEnd is TRUE, fragments (pairs of reads) instead of reads will be counted. featureCounts function checks if reads from the same pair are adjacent to each other (this could happen when reads were for example sorted by their mapping locations), and it automatically reorders those reads that belong to the same pair but are not adjacent to each other in the input read file.

## Value

A list with the following components:

counts	a data matrix containing read counts for each feature or meta-feature for each library.
counts_junction (optional)	a data frame including primary and secondary genes overlapping an exon junction, position information for the left splice site ('Site1') and the right splice site ('Site2') of a junction (including chromosome name, coordinate and strand) and number of supporting reads for each junction in each library. Both primary and secondary genes overlap at least one of the two splice sites of a junction. Secondary genes do not overlap more splice sites than the primary gene. When the primary and secondary genes overlap same number of splice sites, the gene with the smallest leftmost base position is selected as the primary gene. For each junction, no more than one primary gene is reported but there might be more

	than one secondary genes reported. If a junction does not overlap any genes, it has a missing value in the fields of primary gene and secondary gene. Note that this data frame is only generated when <code>juncCounts</code> is <code>TRUE</code> .
<code>annotation</code>	a data frame with six columns including <code>GeneID</code> , <code>Chr</code> , <code>Start</code> , <code>End</code> and <code>Length</code> . When read summarization was performed at feature level, each row in the data frame is a feature and columns in the data frame give the annotation information for the features. When read summarization was performed at meta-feature level, each row in the data frame is a meta-feature and columns in the data frame give the annotation information for the features included in each meta feature except the <code>Length</code> column. For each meta-feature, the <code>Length</code> column gives the total length of genomic regions covered by features included in that meta-feature. Note that this length will be less than the sum of lengths of features included in the meta-feature when there are features overlapping with each other. Also note the <code>GeneID</code> column gives Entrez gene identifiers when the in-built annotations are used.
<code>targets</code>	a character vector giving sample information.
<code>stat</code>	a data frame giving numbers of unassigned reads and the reasons why they are not assigned (eg. ambiguity, multi-mapping, secondary alignment, mapping quality, fragment length, chimera, read duplicate, non-junction and so on), in addition to the number of successfully assigned reads for each library.

**Author(s)**

Wei Shi and Yang Liao

**References**

Yang Liao, Gordon K Smyth and Wei Shi. featureCounts: an efficient general-purpose program for assigning sequence reads to genomic features. *Bioinformatics*, 30(7):923-30, 2014.

**See Also**

[getInBuiltAnnotation](#)

**Examples**

```
## Not run:
library(Rsubread)

# Summarize SAM format single-end reads using built-in RefSeq annotation for mouse genome mm10:
featureCounts(files="mapping_results_SE.sam",annot.inbuilt="mm10")

# Summarize single-end reads using a user-provided GTF annotation file:
featureCounts(files="mapping_results_SE.sam",annot.ext="annotation.gtf",
isGTFAnnotationFile=TRUE,GTF.featureType="exon",GTF.attrType="gene_id")

# Summarize single-end reads using 5 threads:
featureCounts(files="mapping_results_SE.sam",nthreads=5)

# Summarize BAM format single-end read data:
featureCounts(files="mapping_results_SE.bam")

# Perform strand-specific read counting (strandSpecific=2 if reversely stranded):
featureCounts(files="mapping_results_SE.bam",strandSpecific=1)
```

```
# Summarize paired-end reads and counting fragments (instead of reads):
featureCounts(files="mapping_results_PE.bam",isPairedEnd=TRUE)

# Count fragments satisfying the fragment length criteria, eg. [50bp, 600bp]:
featureCounts(files="mapping_results_PE.bam",isPairedEnd=TRUE,
checkFragLength=TRUE,minFragLength=50,maxFragLength=600)

# Count fragments that have both ends successfully aligned without checking the fragment length:
featureCounts(files="mapping_results_PE.bam",isPairedEnd=TRUE,requireBothEndsMapped=TRUE)

# Exclude chimeric fragments from fragment counting:
featureCounts(files="mapping_results_PE.bam",isPairedEnd=TRUE,countChimericFragments=FALSE)

## End(Not run)
```

---

findCommonVariants      *Finding the common variants among all input VCF files*

---

## Description

The common variants (inc. SNPs and indels) among all the input files are found. A data frame containing these common variants is returned. The data frame has a similar format as VCF files.

## Usage

```
findCommonVariants(VCF_files)
```

## Arguments

VCF\_files      a character vector giving the names of VCF format files.

## Details

This function loads all variants (SNPs and indels) from the input VCF files, and find the common variants that are reported in all the VCF files. If a variant record in a input VCF file has multiple alternative sequences (split by ‘;’), each alternative sequence is treated as a single variant. Two variants in two VCF files are the same only if their genomic locations, their reference sequences, their alternative sequences and their variant types are identical.

This function currently does not support other types of variants other than SNPs and indels.

There are eight columns in the returned data frame: chromosome name, position, identity, reference sequence, alternative sequence, quality, filter and extra information. The input may have more columns; these columns are not included in the data frame. If the identity, the quality, the filter and the extra information for the same variant are different among the input VCF files, those information associated with the lowest quality value of this variant among the VCF files is reported in the resulted data frame. For example, if an SNP on chrX:12345 (A=>G) is reported in all the three input VCF files, and the quality scores in the three VCF files are 100, 10, 50 respectively, the identity, the quality, the filter and the extra information in the second VCF file are reported in the resulted data frame for this SNP.

**Value**

A data frame containing the common variants among all the input VCF files is returned. The first eight columns are: chromosome name, position, identity, reference sequence, alternative sequence, quality, filter and extra information.

If there are not any common variants, this function returns an NA value.

**Author(s)**

Yang Liao and Wei Shi

**Examples**

```
## Not run:
# finding the common variants between to input VCF files: a.vcf and b.vcf
library(Rsubread)
findCommonVariants(c('a.vcf', 'b.vcf'))

## End(Not run)
```

---

getInBuiltAnnotation    *Retrieve in-built annotations provided by featureCounts function*

---

**Description**

Retrieve an in-built annotation and save it to a data frame

**Usage**

```
getInBuiltAnnotation(annotation="mm10")
```

**Arguments**

annotation	a character string specifying the in-built annotation to be retrieved. It has four possible values including mm10, mm9, hg38 and hg19, corresponding to the NCBI RefSeq annotations for genomes 'mm10', 'mm9', 'hg38' and 'hg19', respectively. mm10 by default.
------------	--

**Details**

The [featureCounts](#) read summarization function provides in-built annotations for conveniently summarizing reads to genes or exons, and this function allows users to have access to those in-built annotations.

For more information about these annotations, please refer to the help page for [featureCounts](#) function.

**Value**

A data frame with five columns including GeneID, Chr, Start, End and Strand.

**Author(s)**

Wei Shi

**See Also**[featureCounts](#)**Examples**

```
library(Rsubread)
x <- getInBuiltAnnotation("hg38")
x[1:5,]
```

---

**processExons***Obtain chromosomal coordinates of each exon using NCBI annotation*

---

**Description**

This is for internal use.

**Usage**

```
processExons(filename="human_seq_gene.md", species="hg")
```

**Arguments**

filename	a character string giving the name of input .md file (NCBI annotation file)
species	a character string specifying the species

**Details**

This is an internal function and should not be called by users directly.

It processes the NCBI mapview annotation data downloaded from the following links: (these annotations include chromosomal coordinates of UTR and CDS regions of genes).

[ftp://ftp.ncbi.nlm.nih.gov/genomes/H\\_sapiens/mapview/seq\\_gene.md.gz](ftp://ftp.ncbi.nlm.nih.gov/genomes/H_sapiens/mapview/seq_gene.md.gz)

[ftp://ftp.ncbi.nlm.nih.gov/genomes/M\\_musculus/mapview/seq\\_gene.md.gz](ftp://ftp.ncbi.nlm.nih.gov/genomes/M_musculus/mapview/seq_gene.md.gz)

This function finds the chromosomal coordinates of intergenic regions (regions between neighbouring genes) and then outputs them to a file. It also outputs to a file chromosomal coordinates of exons of genes by concatenating UTRs with CDSs and merging overlapping CDSs within each gene. The generated annotation files will then be used by [detectionCallAnnotation](#) function to produce annotation data required by [detectionCall](#) function.

**Value**

Two annotation files are written to the current working directory. This function returns a NULL object.

**Author(s)**

Zhiyin Dai and Wei Shi

---

promoterRegions	<i>Generate annotation for promoter region of genes</i>
-----------------	---

---

**Description**

Create a SAF data.frame of genewise promoter regions

**Usage**

```
promoterRegions(annotation="mm10", upstream=3000L, downstream=2000L)
```

**Arguments**

annotation	a character string giving the name of a genome with built-in annotation or a SAF format gene annotation. If using built-in annotation, the character string should be one of the following: mm10, mm9, hg38 or hg19. They correspond to the NCBI RefSeq annotations for genomes 'mm10', 'mm9', 'hg38' and 'hg19', respectively. mm10 by default.
upstream	an integer giving the number of upstream bases that will be included in the promoter region generated for each gene. These bases are taken immediately upstream (5' end) from transcriptional start site of each gene.
downstream	an integer giving the number of downstream bases that will be included in the promoter region generated for each gene. These bases are taken immediately downstream (3' end) from transcriptional start site of each gene.

**Details**

This function takes as input a SAF format gene annotation and produces a SAF format annotation that contains chromosomal coordinates of promoter region of each gene. See [featureCounts](#) for definition of the SAF format.

Built-in annotations can be accessed via function [getInBuiltAnnotation](#).

**Value**

A SAF format data.frame with columns GeneID, Chr, Start, End and Strand.

**Author(s)**

Gordon K Smyth

**See Also**

[featureCounts](#), [getInBuiltAnnotation](#)

**Examples**

```
library(Rsubread)
x <- promoterRegions()
head(x)
```

---

propmapped	<i>Calculate the proportion of mapped reads/fragments in SAM/BAM files</i>
------------	--

---

### Description

Number of mapped reads/fragments will be counted and fraction of such reads/fragments will be calculated.

### Usage

```
propmapped(files, countFragments=TRUE, properlyPaired=FALSE, verbose=FALSE)
```

### Arguments

files	a character vector giving the names of SAM/BAM format files. Format of input files is automatically determined by the function.
countFragments	logical, indicating whether reads or fragments (read pairs) should be counted. If TRUE, fragments will be counted when paired-end read data are provided. This function automatically detects if the data are single end or paired end. For single end data, each read is treated as a fragment and therefore the value of this parameter should be set to TRUE.
properlyPaired	logical, indicating if only properly paired reads will be counted. This is only applicable for paired end data. FALSE by default.
verbose	display verbose information.

### Details

This function uses the FLAG field in the SAM/BAM to look for mapped reads and count them. Reads/fragments, which have more than one reported location, will be reported only once.

When counting single end reads, counting reads has the same meaning with counting fragments (the results are identical).

### Value

A data frame containing the total number of reads, number of mapped reads and proportion of mapped reads for each library.

### Author(s)

Wei Shi and Yang Liao

### Examples

```
# build an index using the sample reference sequence provided in the package
# and save it to the current directory
library(Rsubread)
ref <- system.file("extdata", "reference.fa", package="Rsubread")
buildindex(basename="./reference_index", reference=ref)

# align the sample read data provided in this packge to the sample reference
```

```
# and save the mapping results to the current directory
reads <- system.file("extdata","reads.txt.gz",package="Rsubread")
align(index="./reference_index",readfile1=reads,output_file="./Rsubread_alignment.BAM")

# get the percentage of successfully mapped reads
propmapped("./Rsubread_alignment.BAM")
```

---

qualityScores	<i>Extract quality score data in a sequencing read dataset</i>
---------------	--

---

## Description

Extract quality strings and convert them to Phred scores

## Usage

```
qualityScores(filename, input_format="gzFASTQ", offset=33, nreads=10000)
```

## Arguments

filename	character string giving the name of an input file containing sequence reads.
input_format	character string specifying format of the input file. gzFASTQ (gzipped FASTQ) by default. Acceptable formats include gzFASTQ, FASTQ, SAM and BAM. Character string is case insensitive.
offset	numeric value giving the offset added to the base-calling Phred scores. Possible values include 33 and 64. By default, 33 is used.
nreads	numeric value giving the number of reads from which quality scores are extracted. 10000 by default.

## Details

Quality scores of read bases are represented by ASCII characters in next-gen sequencing data. This function extracts the quality characters from each base in each read and then converts them to Phred scores using the provided offset value (`offset`).

If the total number of reads in a dataset is  $n$ , then every  $n/nreads$  read is extracted from the input data.

## Value

A data matrix containing Phred scores for read bases. Rows in the matrix are reads and columns are base positions in each read.

## Author(s)

Wei Shi, Yang Liao and Zhiyin Dai

## Examples

```
library(Rsubread)
reads <- system.file("extdata","reads.txt.gz",package="Rsubread")
x <- qualityScores(filename=reads,offset=64,nreads=1000)
x[1:10,1:10]
```

---

removeDupReads	<i>Remove sequencing reads which are mapped to identical locations</i>
----------------	--

---

### Description

Remove reads which are mapped to identical locations, using mapping location of the first base of each read.

### Usage

```
removeDupReads(SAMfile, threshold=50, outputFile)
```

### Arguments

SAMfile	a character string giving the name of a SAM format input file.
threshold	a numeric value giving the threshold for removing duplicated reads, 50 by default. Reads will be removed if they are found to be duplicated equal to or more than threshold times.
outputFile	a character string giving the base name of output files.

### Details

This function uses the mapping location of first base of each read to find duplicated reads. Reads are removed if they are duplicated more than threshold number of times.

### Value

A SAM file including the remaining reads after duplicate removal.

### Author(s)

Yang Liao and Wei Shi

---

repair	<i>Re-order paired-end reads to place reads from the same pair next to each other</i>
--------	---

---

### Description

Fast re-ordering of paired-end reads using read names and mapping locations.

### Usage

```
repair(inFiles, inFormat="BAM", outFiles=paste(inFiles, "repair", sep="."),
addDummy=TRUE, fullData=TRUE, compress=FALSE, nthreads=8)
```

**Arguments**

<code>inFiles</code>	a character vector giving names of input files. These files are typically location-sorted BAM files.
<code>inFormat</code>	a character string specifying format of input files. Supported formats include BAM and SAM.
<code>outFiles</code>	a character string giving names of output files. Re-ordered reads are saved to BAM-format files.
<code>addDummy</code>	logical indicating if a dummy read will be added to each singleton read which has a missing pair in the input. TRUE by default.
<code>fullData</code>	logical indicating if sequences and base-calling quality scores of reads will be included in the output. TRUE by default.
<code>compress</code>	logical indicating if compression should be turned on when generating BAM output. FALSE by default.
<code>nthreads</code>	a numeric value giving number of CPU threads. 8 by default.

**Details**

This function takes as input paired-end BAM or SAM files, re-orders reads to make reads from the same pair be adjacent to each other and then outputs the re-ordered reads into BAM files.

The function makes use of both read names and mapping information of reads (eg. mapping coordinates) to identify reads belonging to the same pair. This makes sure that all paired-end reads are correctly re-ordered, especially those multi-mapping read pairs that include more than one reported alignment in the input.

The BAM files produced by this function are compatible with [featureCounts](#), meaning that no read re-ordering will be performed when providing these files to [featureCounts](#).

**Value**

No value is produced but BAM files with re-ordered reads are written to the current working directory.

**Author(s)**

Wei Shi and Yang Liao

---

RsubreadUsersGuide      *View Rsubread Users Guide*

---

**Description**

Users Guide for Rsubread and Subread

**Usage**

```
RsubreadUsersGuide()
```

**Details**

The Subread/Rsubread Users Guide provides detailed description to the functions and programs included in the Subread and Rsubread software packages. It also includes case studies for analyzing next-gen sequencing data.

The Subread package is written in C and it can be downloaded from <http://subread.sourceforge.net>. The Rsubread package provides R wrappers functions for many of the programs included in Subread package.

**Value**

Character string giving the file location.

**Author(s)**

Wei Shi

**See Also**

[vignette](#)

---

sam2bed

*Convert a SAM format file to a BED format file*

---

**Description**

SAM to BED conversion

**Usage**

```
sam2bed(samfile, bedfile, readlen)
```

**Arguments**

samfile	character string giving the name of input file. Input format should be in SAM format.
bedfile	character string giving the name of output file. Output file is in BED format.
readlen	numeric value giving the length of reads included in the input file.

**Details**

This function converts a SAM format file to a BED format file, which can then be displayed in a genome browser like UCSC genome browser, IGB, IGV.

**Value**

No value is produced but a BED format file is written to the current working directory. This file contains six columns including chromosomal name, start position, end position, name('.'), mapping quality score and strandness.

**Author(s)**

Wei Shi

---

txUnique	<i>Count number of bases unique to each transcript</i>
----------	--

---

**Description**

For each transcript, number of unique bases and total number of bases will be reported.

**Usage**

```
txUnique(GTF_Annotation_File, Feature_Type = "exon", Gene_ID_Attribute = "gene_id", Transcript_ID
```

**Arguments**

**GTF\_Annotation\_File**  
character string giving the name of a GTF file containing the transcript-level annotations.

**Feature\_Type**  
character string specifying the type of annotations. Only the annotations of the given type will be loaded from the GTF file. exon by default. The third column in the GTF file contains the type of the annotation in each line

**Gene\_ID\_Attribute**  
character string specifying the attribute name in the 9-th column in the GTF file for the gene identifier. gene\_id by default.

**Transcript\_ID\_Attribute**  
character string specifying the attribute name in the 9-th column in the GTF file for the transcript identifier. transcript\_id by default.

**Details**

This function compares the transcripts belonging to the same gene and then counts the number of bases unique to each transcript. It also reports the total number of bases for each transcript. When a transcript is found to contain overlapping exons, the overlapping exons will be merged and each overlapping base will be counted only once.

**Value**

A data matrix in which each row represents a transcript. The matrix includes four columns: gene identifier, transcript identifier, number of bases unique to each transcript and total number of bases each transcript has.

**Author(s)**

Yang Liao and Wei Shi

# Index

## \*Topic **documentation**

RsubreadUsersGuide, 29

align, 2, 10, 19

atgcContent, 8

buildindex, 7, 9

createAnnotationFile, 10

detectionCall, 11, 12, 24

detectionCallAnnotation, 12, 24

exactSNP, 12

featureCounts, 6, 14, 23–25, 29

findCommonVariants, 22

getInBuiltAnnotation, 19, 21, 23, 25

processExons, 12, 24

promoterRegions, 25

propmapped, 26

qualityScores, 27

removeDupReads, 28

repair, 28

RsubreadUsersGuide, 29

sam2bed, 30

subjunc, 10, 19

subjunc (align), 2

txUnique, 31

vignette, 30

write.Rsubread (createAnnotationFile),  
10