BicARE
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BicARE-package BicARE

Description

Biclustering Analysis and Results Exploration

Details

Package: BicARE
Version: 0.1.0
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Further information is available in the following vignettes:

BiCARE  BiCARE (source, pdf)

Author(s)

Pierre Gestraud
Maintainer: Pierre Gestraud , <pierre.gestraud@curie.fr>

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bicluster

Extract a bicluster

**Description**

Extract a bicluster from an object of class biclustering

**Usage**

bicluster(biclustering, k, graph=TRUE)

**Arguments**

biclustering  an object of class "biclustering" created by function FLOC

k             the number of the bicluster considered in the "biclustering" object

graph         boolean, indicating whether the graph should be plotted or not

**Value**

Returns the bicluster as a matrix with the genes on rows and the samples on columns. Result matrix is of class "bicluster". The "graph" option allows to plot the expression profiles of the genes across the conditions in the bicluster.

**Author(s)**

Pierre Gestraud

**Examples**

```r
### extract the first bicluster
data(sample.biclustering)
sample.biclustering
bic <- bicluster(sample.biclustering, 1, graph=TRUE)
plot(bic)
```
FLOC

Performs the FLOC algorithm

Description

Find a given number of biclusters using the a modified version of the FLOC algorithm.

Usage

FLOC(Data, k = 20, pGene = 0.5, pSample = pGene, r = NULL, N = 8, M = 6, t = 500,)

Arguments

Data  
an ExpressionSet-class or a matrix (with genes on rows and conditions on columns)

k  
the number of biclusters searched

pGene  
genes initial probability of membership to the biclusters

pSample  
samples initial probability of membership to the biclusters

r  
the residue threshold

N  
minimal number of gene per bicluster

M  
minimal number of conditions per bicluster

t  
number of iterations

blocGene  
a matrix indicating the directed initialisation for the genes (see details)

blocSample  
a matrix indicating the directed initialisation for the conditions (see details)

Details

This biclustering algorithm is based on the FLOC algorithm (FLexible Overlapped biClustering) defined by Yang et al. (see references). It can discover a set of k, possibly overlapping, biclusters. If r is set to NULL, the residue threshold used in the analysis is the residue of Data divided by 10. blocGene and blocSample are matrix of 0 and 1 with the rows representing the features (gene or samples) and the columns the biclusters. A 1 on line i and column j indicates that the feature i (gene or sample) will be include in the bicluster j during the initialisation step and will not be removed from it during the analysis. If the number of columns in these matrices is different from the number of bicluster searched, k is set to the maximal value of these two.

See bicluster to extract a bicluster from the biclustering result.

Value

Returns an object of class `biclustering`, a list containing at least:

Call  
the matched call.

ExpressionSet  
the data used

param  
a data.frame with the algorithm parameters

bicRow  
a matrix of boolean indicating the belonging of the genes to the biclusters

bicCol  
the same as for bicRow but for the conditions

mat.resvol.bic  
a matrix describing the biclusters
makeReport

Author(s)

Pierre Gestraud (pierre.gestraud@curie.fr)

References


Examples

data(sample.bicData)  ## subset of sample.ExpressionSet from Biobase
residue(sample.bicData)  ## 0.3401921
resBic <- FLOC(sample.bicData, k=10, pGene=0.5, r=0.05, N=8, M=10, t=500)
resBic

## initialising samples of 2 biclusters
iniSample <- matrix(0, ncol=2, nrow=26)
## first bicluster initialised around Female cases
iniSample[pData(sample.bicData)$sex=='Female', 1] <- 1
## second bicluster initialised around control cases
iniSample[pData(sample.bicData)$type=='Control', 2] <- 1
resBic <- FLOC(sample.bicData, k=10, pGene=0.5, r=0.05, N=8, M=10, t=500, blocSample=iniSample)
resBic

makeReport  ## Export the results as html files

Description

Exports the results as html files.

Usage

makeReport(dirPath, dirName, resBic, browse=TRUE)

Arguments

dirPath  path to the directory
dirName  the name of the directory where the report will be created
resBic  a biclustering result
browse  logical. If TRUE the web browser will be opened

Details

makeReport produces an html report of biclustering results in a new directory named dirName. If the browse argument is set to TRUE the web browser will be opened on the "home.html" file.

Make sure to have rights to create the result directory.

Author(s)

Pierre Gestraud (pierre.gestraud@curie.fr)
Examples

data(sample.biclustering)
dirPath <- getwd()  ## report created in the current working directory
dirName <- "test"
makeReport(dirPath, dirName, sample.biclustering, browse=FALSE)

residue

Residue of a matrix

Description

Returns the residue of a matrix.

Usage

residue(Data)

Arguments

Data an ExpressionSet-class or a matrix

Details

This function computes the residue of a matrix as defined by Yang et al (see references).

Author(s)

Pierre Gestraud

References


See Also

FLOC

Examples

data(sample.bicData)
residue(sample.bicData)
sample.bicData

*Example data set for BicARE*

**Description**
A subset of sample.ExpressionSet from package Biobase. The data for 26 cases, labeled A to Z and 350 genes. Each case has three covariates: sex (male/female), type (case/control) and score (testing score).

**Usage**
sample.bicData

**Format**
An ExpressionSet

---

sample.biclustering

*Example biclustering object*

**Description**
A biclustering object created by the FLOC function on the sample.bicData with the following options: k=10, pGene = 0.3, pSample = 0.5, r = 0.025, N = 8, M = 8, t = 1000.

**Usage**
sample.biclustering

**Format**
a biclustering object

---

testAnnot

*Find samples annotations over-represented covariates in biclusters*

**Description**
Characterisation of the biclusters in term of over-representation of sample covariates.

**Usage**
testAnnot(resBic, annot=NULL, covariates="all")
Arguments

- **resBic**: a biclustering result from `FLOC`
- **annot**: annotation matrix, default value is set to NULL, then phenoData of the ExpressionSet is used
- **covariates**: the names of the covariates that should be tested, default value is set to "all"

Details

For each bicluster and each covariate a chi-squared test is performed to test the adequation between the distribution of the levels of the covariates in the bicluster and in the original dataset.

Multiple testing correction is performed by the Benjamini-Yekutieli procedure. The residuals of the tests indicate if the level is over or down represented in the bicluster.

Due to the amount of results it is advised to use the `makeReport` function to get a html report.

Value

A biclustering object containing `resBic` and updated with the results of the tests in `resBic$covar`.

The results are presented as a list with:

- **covar**: the samples covariates tested
- **pvalues**: a matrix with the p-values of the tests
- **adjpvalues**: a matrix with the p-values adjusted by the Benjamini Yekutieli procedure
- **index**: a list of matrices with the numbers of each level in each bicluster
- **residuals**: a list of matrices with the residuals of the tests for each modality in each bicluster

Author(s)

Pierre Gestraud

Examples

```r
data(sample.biclustering)
resBic <- testAnnot(sample.biclustering, annot=NULL, covariates=c("sex", "type"))
```

`testSet` Find gene sets that are enriched in a bicluster

Description

Test of the over-representation of gene sets in the biclusters

Usage

`testSet(resBic, geneSetCol)`

Arguments

- **resBic**: a biclustering object created by `FLOC`
- **geneSetCol**: a GeneSetCollection-class
Details
The over-representation of a gene set in a bicluster is evaluated by an hypergeometric test.
The genes identifiers of the gene sets will automatically be mapped to the same as those used in the
data.
Due to the amount of results it is advised to use the makeReport function to get a html report.

Value
A biclustering object containing resBic and updated with the results of the tests in resBic$geneSet.
The results are presented as a list with:
GeneSetCollection
the GeneSetCollection used
pvalues
a matrix containing the pvalues of the tests for each geneSet and each bicluster
adjpvalue
a matrix containing the p-values adjusted by the Benjamini Yekutieli procedure

Author(s)
Pierre Gestraud (pierre.gestraud@curie.fr)

Examples
data(sample.biclustering)
gss <- GeneSetCollection(sample.biclustering$ExpressionSet[1:50,, setType=GOCollection()]
resBic <- testSet(sample.biclustering, gss)
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