

Package ‘RegionalST’

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Type Package

Title Investigating regions of interest and performing cross-regional analysis with spatial transcriptomics data

Version 1.3.0

Description This package analyze spatial transcriptomics data through cross-regional analysis. It selects regions of interest (ROIs) and identifies cross-regional cell type-specific differential signals. The ROIs can be selected using automatic algorithm or through manual selection. It facilitates manual selection of ROIs using a shiny application.

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| | |
|--------|--|
| DoGSEA | <i>Perform GSEA analysis for cross-regional DE genes</i> |
|--------|--|

Description

Perform GSEA analysis for cross-regional DE genes

Usage

```
DoGSEA(considerRes, whichDB = "hallmark", gmdir = NULL, withProp = FALSE)
```

Arguments

| | |
|-------------|--|
| considerRes | A list of cross-regional DE genes. |
| whichDB | A character string to select the database names, e.g., "hallmark", "kegg", "reactome". |
| gmdir | Directory for external database gmt file location. |
| withProp | Whether deconvolution proportion is used in previous steps. |

Value

A list including GSEA results for all cell types.

Examples

```
data(exampleRes)
allCTres <- DoGSEA(exampleRes, whichDB = "hallmark", withProp = TRUE)
```

DrawDotplot*Draw dot plot for GSEA results of cross-regional DE genes*

Description

Draw dot plot for GSEA results of cross-regional DE genes

Usage

```
DrawDotplot(  
  allCTres,  
  CT = 1,  
  angle = 20,  
  vjust = 0.9,  
  hjust = 1,  
  padj_cutoff = 1,  
  topN = 20,  
  chooseP = "padj",  
  eachN = NULL  
)
```

Arguments

| | |
|--------------------------|---|
| <code>allCTres</code> | A list of GSEA results for all cell types. |
| <code>CT</code> | A number of the interested cell type, e.g., 1, 2, 3. |
| <code>angle</code> | A number of plotting parameter, angle of the x axis label. |
| <code>vjust</code> | A number of vertical adjustment in plotting. |
| <code>hjust</code> | A number of horizontal adjustment in plotting. |
| <code>padj_cutoff</code> | A cutoff number of adjusted p value. |
| <code>topN</code> | A number of the plotted top pathways. |
| <code>chooseP</code> | A character string for the p value that used in plotting, e.g., "padj" or "pval". |
| <code>eachN</code> | The maximum number of pathways in each cell type. |

Value

A plot object

Examples

```
data(exampleRes)
allCTres <- DoGSEA(exampleRes, whichDB = "hallmark", withProp = TRUE)
DrawDotplot(allCTres, CT = 1, angle = 15, vjust = 1, chooseP = "padj")
```

DrawRegionProportion *Draw regional cell type distribution with cell type annotation information*

Description

Draw regional cell type distribution with cell type annotation information

Usage

```
DrawRegionProportion(sce, label = "celltype", selCenter = seq_len(10))
```

Arguments

| | |
|-----------|--|
| sce | A single cell experiment object. |
| label | A string character for the cell type variable. |
| selCenter | A vector of the interested ROIs, e.g., 1:4. |

Value

A plot object.

Examples

```
data("example_sce")
DrawRegionProportion(example_sce, label = "celltype", selCenter = 1:3)
```

DrawRegionProportion_withProp *Draw regional cell type distribution with cellular proportion information*

Description

Draw regional cell type distribution with cellular proportion information

Usage

```
DrawRegionProportion_withProp(  
  sce,  
  label = "CARD_CellType",  
  selCenter = seq_len(10)  
)
```

Arguments

| | |
|-----------|--|
| sce | A single cell experiment object. |
| label | A string character for the cell type variable. |
| selCenter | A vector of the interested ROIs, e.g., 1:4. |

Value

A plot object.

Examples

```
data("example_sce")  
DrawRegionProportion_withProp(example_sce,  
  label = "Proportions",  
  selCenter = 1:3)
```

exampleRes

Example DE output

Description

A simulated example DE output file

Usage

```
data(exampleRes)
```

Format

A list object.

Value

A list object.

Examples

```
data(exampleRes)
```

`example_sce`*Example single cell experiment for input*

Description

A simulated example input data file

Usage

```
data(example_sce)
```

Format

A SingleCellExperiment object.

Value

A SingleCellExperiment object.

Examples

```
data(example_sce)
```

`FindRegionalCells`*Identify regional cells given centers and radiuses*

Description

Identify regional cells given centers and radiuses

Usage

```
FindRegionalCells(  
  sce,  
  centerID,  
  enhanced = FALSE,  
  radius = 10,  
  avern = 5,  
  doPlot = FALSE,  
  returnPlot = FALSE  
)
```

Arguments

| | |
|------------|--|
| sce | A single cell experiment object. |
| centerID | One or a vector of spot IDs as centers of ROIs. |
| enhanced | A logical variable for plotting enhanced plot or now. Default is FALSE. |
| radius | A number of fixed ROI radius. |
| avern | A number of the average sites used to compute unit distance, default is 5. |
| doPlot | A logical variable to specify whether plot the figure or not. |
| returnPlot | a logical variable to specify whether output the plot or not. |

Value

A list including center spot ID and regional spot IDs.

Examples

```
# FindRegionalCells(sce, centerID = "ACGCCTGACACGCGCT-1")
```

GetCrossRegionalDE_raw

Identify cross-regional differential analysis

Description

Identify cross-regional differential analysis

Usage

```
GetCrossRegionalDE_raw(  
  sce,  
  twoCenter = c(3, 4),  
  enhanced = FALSE,  
  label = "celltype",  
  n_markers = 10,  
  logfc.threshold = 0.25,  
  angle = 30,  
  hjust = 0,  
  size = 3,  
  min.pct = 0.1,  
  padj_filter = 0.05,  
  doHeatmap = TRUE  
)
```

Arguments

| | |
|-----------------|---|
| sce | A single cell experiment object. |
| twoCenter | A vector of two numbers for the interested ROI numbers. |
| enhanced | A logical variable for using enhanced data or not. |
| label | A variable name that contains the cell type information. |
| n_markers | A number specifying the top DE gene number. |
| logfc.threshold | A number for the cutoff threshold of log fold change. |
| angle | A number for angle when plotting. |
| hjust | A number for horizontal justification when plotting. |
| size | A number for text font size. |
| min.pct | A number of minimum percentage specified in the Seurat DE function. |
| padj_filter | A number for filtering adjusted p values. |
| doHeatmap | Logical variable for whether drawing the heatmap. |

Value

A list including the top DE genes (topDE), and all DE genes (allDE).

Examples

```
data("example_sce")
example_sce <- mySpatialPreprocess(example_sce, platform="Visium")
# I used a very big padj filter here because this is just a toy data
GetCrossRegionalDE_raw(example_sce, twoCenter = c(1,2),
                        min.pct = 0.01, logfc.threshold = 0.01,
                        padj_filter = 0.5)
```

GetCrossRegionalDE_withProp

Identify cross-regional differential analysis with proportion

Description

Identify cross-regional differential analysis with proportion

Usage

```
GetCrossRegionalDE_withProp(  
  sce,  
  twoCenter = c(3, 4),  
  label = "celltype",  
  n_markers = 10,  
  angle = 30,  
  hjust = 0,  
  size = 3,  
  padj_filter = 0.05,  
  doHeatmap = TRUE  
)
```

Arguments

| | |
|-------------|--|
| sce | A single cell experiment object. |
| twoCenter | A vector of two numbers for the interested ROI numbers. |
| label | A variable name that contains the cell type information. |
| n_markers | A number specifying the top DE gene number. |
| angle | A number for angle when plotting. |
| hjust | A number for horizontal justification when plotting. |
| size | A number for text font size. |
| padj_filter | A number for filtering adjusted p values. |
| doHeatmap | Logical variable for whether drawing the heatmap. |

Value

A list including the top DE genes (topDE), and all DE genes (allDE).

Examples

```
data("example_sce")  
example_sce <- mySpatialPreprocess(example_sce, platform="Visium")  
# Since the example data is very small, I set padj filter as NULL. Default is 0.05.  
GetCrossRegionalDE_withProp(example_sce, twoCenter = c(1,2), padj_filter = NULL)
```

GetOneRadiusEntropy *Computer the entropy for a fixed radius*

Description

Computer the entropy for a fixed radius

Usage

```
GetOneRadiusEntropy(
  sce,
  selectN,
  enhanced = FALSE,
  weight = NULL,
  label = "celltype",
  radius = 10,
  doPlot = FALSE,
  mytitle = NULL
)
```

Arguments

| | |
|----------|--|
| sce | A single cell experiment object. |
| selectN | A total number for selected centers. Should be smaller than the total site number. |
| enhanced | A logical variable of whether using enhanced data. |
| weight | A data frame to specify the weights of all cell types. |
| label | A variable name that contains the cell type information. |
| radius | A number for fixed radius. |
| doPlot | Logical variable about whether draw the plot. |
| mytitle | A character string for the title of the plot. |

Value

A list including the selected centers, computed entropies, radius.

Examples

```
data("example_sce")
weight <- data.frame(celltype = c("Cancer Epithelial", "CAFs",
                                "T-cells", "Endothelial",
                                "PVL", "Myeloid", "B-cells",
                                "Normal Epithelial", "Plasmablasts"),
                    weight = c(0.25,0.05,
                               0.25,0.05,
                               0.025,0.05,
                               0.25,0.05,0.025))
example_sce <- mySpatialPreprocess(example_sce, platform="Visium")
GetOneRadiusEntropy(example_sce, selectN = round(length(example_sce$spot)/2),
                    weight = weight, radius = 5, doPlot = TRUE,
                    mytitle = "Radius 5 weighted entropy")
```

 GetOneRadiusEntropy_withProp

Computer the entropy for a fixed radius with cell type proportion

Description

Computer the entropy for a fixed radius with cell type proportion

Usage

```
GetOneRadiusEntropy_withProp(
  sce,
  selectN,
  weight = NULL,
  label = "celltype",
  radius = 10,
  doPlot = FALSE,
  mytitle = NULL
)
```

Arguments

| | |
|---------|--|
| sce | A single cell experiment object. |
| selectN | A total number for selected centers. Should be smaller than the total site number. |
| weight | A data frame to specify the weights of all cell types. |
| label | A variable name that contains the cell type information. |
| radius | A number for fixed radius. |
| doPlot | Logical variable about whether draw the plot. |
| mytitle | A character string for the title of the plot. |

Value

A list including the selected centers, computed entropies, radius.

Examples

```
data("example_sce")
weight <- data.frame(celltype = c("Cancer Epithelial", "CAFs", "T-cells", "Endothelial",
  "PVL", "Myeloid", "B-cells", "Normal Epithelial", "Plasmablasts"),
  weight = c(0.25,0.05,
    0.25,0.05,
    0.025,0.05,
    0.25,0.05,0.025))
example_sce <- mySpatialPreprocess(example_sce, platform="Visium")
GetOneRadiusEntropy_withProp(example_sce, selectN = round(length(example_sce$spot)/10),
  weight = weight,
```

```
radius = 5,
doPlot = TRUE,
mytitle = "Radius 5 weighted entropy")
```

getProportion *Define an accessor method for Proportion_CARD*

Description

Define an accessor method for Proportion_CARD

Usage

```
getProportion(card)
```

Arguments

card A CARD object.

Value

A matrix containing the spot-level cell type proportion information

Examples

```
# getProportion(card)
```

ManualSelectCenter *Manually select top ROIs*

Description

Manually select top ROIs

Usage

```
ManualSelectCenter(sce)
```

Arguments

sce A single cell experiment object.

Value

An sce object with selected centers and radiuses.

Examples

```
data("example_sce")
example_sce <- mySpatialPreprocess(example_sce, platform="Visium")
# I commented this out because the shiny app will get stuck without input.
# example_sce <- ManualSelectCenter(example_sce)
```

mySpatialPreprocess *Perform Preprocessing for spatial data (tailored from BayesSpace function)*

Description

Perform Preprocessing for spatial data (tailored from BayesSpace function)

Usage

```
mySpatialPreprocess(
  sce,
  platform = c("Visium", "ST"),
  n.PCs = 15,
  n.HVGs = 2000,
  skip.PCA = FALSE,
  assay.type = "logcounts"
)
```

Arguments

| | |
|------------|--|
| sce | A SingleCellExperiment object. |
| platform | Which platform the data are from, Visium or ST. |
| n.PCs | Number of PCs used in the analysis. |
| n.HVGs | Number of highly variable genes used in the analysis. |
| skip.PCA | A boolean variable to choose whether skipping the PCA step or not. |
| assay.type | Which assay to use, default is logcounts. |

Value

A processed SingleCellExperiment object.

Examples

```
data(example_sce)
example_sce <- mySpatialPreprocess(example_sce, platform="Visium")
```

pathways_hallmark *Hallmark database*

Description

Hallmark database downloaded from MSigDB (Feb, 2023)

Usage

```
data(pathways_hallmark)
```

Format

A list object.

Value

A list object.

Source

MSigDB

References

Liberzon et al. (2015) Cell Syst. 1(6):417-425 ([PubMed](#))

Examples

```
data(pathways_hallmark)
```

pathways_kegg *KEGG database*

Description

KEGG database downloaded from MSigDB (Feb, 2023)

Usage

```
data(pathways_kegg)
```

Format

A list object.

Value

A list object.

Source

[MSigDB](#)

References

Kanehisa and Goto (2000) Nucleic Acids Research 28(1):27-30 ([PubMed](#))

Examples

```
data(pathways_kegg)
```

| | |
|-------------------|--------------------------|
| pathways_reactome | <i>REACTOME database</i> |
|-------------------|--------------------------|

Description

REACTOME database downloaded from MSigDB (Feb, 2023)

Usage

```
data(pathways_reactome)
```

Format

A list object.

Value

A list object.

Source

[MSigDB](#)

References

Jassal et al. (2020) Nucleic Acids Research 28(1):27-30 ([PubMed](#))

Examples

```
data(pathways_reactome)
```

PlotOneSelectedCenter *Plot one selected ROI*

Description

Plot one selected ROI

Usage

```
PlotOneSelectedCenter(sce, ploti, enhanced = FALSE)
```

Arguments

sce A single cell experiment object.
ploti A number of indicate which ROI to plot.
enhanced A logical variable for using enhanced data or not.

Value

A figure object for the selected ROI.

Examples

```
data("example_sce")  
example_sce <- mySpatialPreprocess(example_sce, platform="Visium")  
PlotOneSelectedCenter(example_sce, ploti = 1)
```

RankCenterByEntropy *Automatically rank ROI centers based on entropy*

Description

Automatically rank ROI centers based on entropy

Usage

```
RankCenterByEntropy(  
  sce,  
  weight,  
  enhanced = FALSE,  
  selectN = round(length(sce$spot)/10),  
  label = "celltype",  
  topN = 10,  
  min_radius = 10,  
  avern = 5,
```



```

    radius_vec = c(10, 15, 20),
    doPlot = TRUE
  )

```

Arguments

| | |
|------------|--|
| sce | A single cell experiment object. |
| weight | A data frame to specify the weights of all cell types. |
| enhanced | A logical variable of whether using enhanced data. |
| selectN | A total number for selected centers. Should be smaller than the total site number. |
| label | A variable name that contains the cell type information. |
| topN | A number to specify the total amount of top ranked ROIs. |
| min_radius | The minimum repellent radius. |
| avern | A number of the average sites used to compute unit distance, default is 5. |
| radius_vec | A vector of numbers for candidate radiuses. |
| doPlot | Logical variable about whether draw the plot. |

Value

An sce object with selected ROI information.

Examples

```

data("example_sce")
example_sce <- mySpatialPreprocess(example_sce, platform="Visium")
weight <- data.frame(celltype = c("Cancer Epithelial", "CAFs", "T-cells", "Endothelial",
  "PVL", "Myeloid", "B-cells", "Normal Epithelial", "Plasmablasts"),
  weight = c(0.25,0.05,
    0.25,0.05,
    0.025,0.05,
    0.25,0.05,0.025))
example_sce <- RankCenterByEntropy(example_sce, weight, label = "celltype",
  selectN = round(length(example_sce$spot)/10),
  topN = 3, min_radius = 10,
  radius_vec = c(10,15),
  doPlot = TRUE)

```

RankCenterByEntropy_withProp

Automatically rank ROI centers based on entropy with proportions

Description

Automatically rank ROI centers based on entropy with proportions

Usage

```
RankCenterByEntropy_withProp(
  sce,
  weight,
  selectN = round(length(sce$spot)/10),
  topN = 10,
  min_radius = 10,
  avern = 5,
  radius_vec = c(10, 15, 20),
  doPlot = TRUE
)
```

Arguments

| | |
|------------|--|
| sce | A single cell experiment object. |
| weight | A data frame to specify the weights of all cell types. |
| selectN | A total number for selected centers. Should be smaller than the total site number. |
| topN | A number to specify the total amount of top ranked ROIs. |
| min_radius | The minimum repellent radius. |
| avern | A number of the average sites used to compute unit distance, default is 5. |
| radius_vec | A vector of numbers for candidate radiuses. |
| doPlot | Logical variable about whether draw the plot. |

Value

An sce object with selected ROI information.

Examples

```
data("example_sce")
weight <- data.frame(celltype = c("Cancer Epithelial", "CAFs", "T-cells", "Endothelial",
  "PVL", "Myeloid", "B-cells", "Normal Epithelial", "Plasmablasts"),
  weight = c(0.25,0.05,
    0.25,0.05,
    0.025,0.05,
    0.25,0.05,0.025))
example_sce <- mySpatialPreprocess(example_sce, platform="Visium")
## I set our min_raius as 10 and radius vector as 10 and 15 as the example dataset is very small
example_sce <- RankCenterByEntropy_withProp(example_sce, weight,
  selectN = round(length(example_sce$spot)/10),
  topN = 3, min_radius = 10,
  radius_vec = c(10,15),
  doPlot = TRUE)
```

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