

# Splice Graphs

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Work of Daniel Bindreither, Marc Carlson, Martin Morgan

## 1 Background: Gene models to splice graphs

Sammeth, 2010, Computation Biology 16: [1117-1140](#)

Starting point (for us): known gene model of exons-within-transcripts-within-genes

Vertices represent gene start, exon donor / acceptor, gene ends, counting bin boundaries

Edges represent paths between vertices

- zero or more exons
- sometimes shared by two or more transcripts

Splice codes

- Summarize order of vertices
- Familiar ('exon skip'; 'alternative promoter'; etc) and exotic

Bubbles

- Represent alternative splicing events
- simple
- compound – three or more transcripts, three-way and higher splicing

## 2 Benefits

Edges contain  $\geq 1$  counting bins, so...

Classification of splice events

Genome-wide assessment of splicing events

Testing bubbles (alternative splicing), in addition to edges

## 3 Example (partial)

### 3.1 Implementation

SpliceGraph package, in development; also spliceGraph function in GenomicFeatures.

Transform exons-within-transcripts-within-genes GRangesList to edges-within-transcripts-within-genes.

- includes DESeq 'counting bins' operation (disjoin)

```
library("SpliceGraph")
library(TxDb.Dmelanogaster.UCSC.dm3.ensGene)
txdb <- TxDb.Dmelanogaster.UCSC.dm3.ensGene
spliced <- spliceGraphs(txdb)
save(spliced, file="2012-07-06-pasilla-spliceGraph.rda")
```

Count via countSpliceGraphReads

Normalization, etc: like DEXSeq

Statistical assessment

- edges: analogous to / using DEXSeq
- ??? bubbles ???  
~ sample + edge + condition \*  $\sum_{bp} I(\text{edge} == bp_i)$

### 3.2 Bubbles

event	Nr
retain 1 intron	1302
alternative promotor usage	892
alternative transcript end	824
2 alternative donors	734
skip 1 exon	695
2 alternative acceptors	658
	...
Total	11913

- FBgn0011224 annotated with 42 exons and 22 transcripts, has 63 bubbles

### 3.3 Edges vs. exons

90249 non-zero count exons, median 25 reads / exon 22981 edges, 32 reads / edge  
Differential expression: stay tuned...

```
load("cD.exons-fb.Rda")
load("cD.sG-fb.Rda")
tbl <- read.table("phenoData.txt", header=TRUE)

exonCounts <- cD.ex[rowMeans(cD.ex) != 0,]
edgeCounts <- cD.sG[rowMeans(cD.sG) != 0,]

nrow(exonCounts)
mean(exonCounts)

nrow(edgeCounts)
mean(edgeCounts)
```

### 3.4 Differential representation (to come...)